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3 Committee for Medicinal Products for Veterinary Use (CVMP)

4 **Guideline on the procedure to be followed when a batch**  
5 **of a vaccine finished product is suspected to be**  
6 **contaminated with bovine viral diarrhoea virus**  
7 **Draft**

Draft agreed by Immunologicals Working Party	October 2014
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9 This guideline replaces 'Guideline on the procedure to be followed when a batch of a vaccine  
10 finished product is suspected to be contaminated with bovine viral diarrhoea (BVD) virus'  
11 (EMA/CVMP/IWP/205351/2006).<sup>1</sup>

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13 Comments should be provided using this [template](#). The completed comments form should be sent  
to [vet-guidelines@ema.europa.eu](mailto:vet-guidelines@ema.europa.eu)

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<sup>1</sup> The guideline was revised to remove the provision for an in vivo test, with the view to ensuring the current best practice with regard to implementation of 3Rs (replacement, reduction and refinement) principles.



14 Guideline on the procedure to be followed when a batch  
15 of a vaccine finished product is suspected to be  
16 contaminated with bovine viral diarrhoea virus

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## 27 **1. Introduction (background)**

28 Starting materials of animal origin, including bovine serum are still essential ingredients of the cell  
29 culture media used in the production of many immunological veterinary medicinal products.  
30 Different risks are associated with the use of such starting materials. The presence of extraneous  
31 agents in bovine serum certainly represents a major risk to the quality of the finished product. One  
32 of the specific risks associated with the use of bovine serum is the contamination of the finished  
33 vaccine with Bovine Viral Diarrhoea Virus (BVDV).

34 The challenge when suspecting contamination of a vaccine batch with a BVDV is the confirmation of  
35 this contamination. It is therefore important to agree on an approach to this confirmation of  
36 contamination, which can be followed and mutual recognised by all the Competent Authorities.

## 37 **2. Scope**

38 The aim of this guideline is to outline the procedure to be followed by the Competent Authorities  
39 when a batch of a vaccine is suspected to be contaminated with BVDV. Considering the risk of  
40 BVDV in bovine serum, the highest risk will be with live and inactivated vaccines indicated for use  
41 in pestivirus susceptible species (bovine, porcine, ovine, caprine. Of these, the greatest risk is  
42 associated with the use of live vaccines in pregnant BVDV susceptible females.

## 43 **3. Legal basis**

44 This guideline has to be read in conjunction with the introduction and general principles and Title II  
45 of Annex I to Directive 2001/82/EC.

## 46 **4. Definition of a suspicion**

47 Contamination of a vaccine batch with BVDV (type 1 or 2) can be suspected when Bovine Viral  
48 Diarrhoea (BVD) or Classical Swine Fever (CSF) antibodies are detected in animals without  
49 vaccination or field infection history after the use of inactivated or live vaccines not containing  
50 BVDV vaccine antigens or in unexplained conditions (e.g. CSF diagnostic in pigs; not associated  
51 with a CSF epizootic or when a CSF outbreak is definitely excluded). Reproductive disorders  
52 (abortion, mummification, stillborn neonates etc.) may be associated with the use of a batch of  
53 vaccine contaminated with live BVDV in pregnant females susceptible to the virus.

54 The risk of non-specific amplification should be taken into account when selecting primers for  
55 detection of BVD genome by reverse transcription polymerase chain reaction (RT-PCR).  
56 Furthermore, the principles stated in the CVMP guideline on Requirements and controls applied to  
57 bovine serum used in the production of immunological veterinary medicinal products  
58 (EMA/CVMP/743/00) which indicates clearly (4.3.1) that the applicant should be able to clarify  
59 whether or not any nucleic acid detected originates from infectious BVDV particles, should also  
60 apply.

61 The use of bovine serum containing inactivated BVDV particles should not (in some cases, albeit  
62 rare, serum is added as a stabiliser to the finished vaccine) induce any serological response in the  
63 animals inoculated with the finished product of the vaccine as the dilution of the bovine serum is  
64 too high during the manufacturing process. However, if the bovine serum contains live BVD viral

65 particles, they will multiply in the cells used for the vaccine production and the viral burden will be  
66 high in the finished product.

67 • If a live vaccine is contaminated with live BVDV (type 1 or 2) and is administered to  
68 pregnant females (bovine, ovine, caprine, porcine), an iatrogenic disease may be induced  
69 in the foetuses which can be similar to BVDV, CSF or border disease. The disease could be  
70 of particular severity in the case of a vaccine contamination caused by a BVDV type 2  
71 strain (haemorrhagic disease).

72 • If an inactivated vaccine is contaminated with BVDV (type 1 or 2) and is administered to  
73 bovines, a seroconversion may be induced, which depends on the concentration of the  
74 contaminant.

75 In pigs, sheep or goats, a vaccine contaminated with BVDV may interfere with the  
76 diagnostic measures for CSF or border disease due to induction of BVD antibodies.

77 For example, after several injections of the vaccine, alleged CSF antibodies can be detected  
78 as the repeated administration of the contaminated vaccine induces the appearance of BVD  
79 antibodies which cross react with CSF antigens. This can interfere with CSF surveillance  
80 programmes based on serological sampling, particularly in breeders which are usually given  
81 numerous vaccines against several viral diseases e.g. Aujeszky's disease, parvovirus.

82 In countries applying eradication or control programmes against BVD, interference with  
83 national surveillance programmes can also occur when cattle are repeatedly vaccinated  
84 with BVDV contaminated, inactivated vaccines such as Infectious Bovine Rhinotracheitis  
85 (IBR).

## 86 5. Detection of BVDV contamination

87 *In vitro* diagnostic methods are currently available which can usefully be used to detect BVDV  
88 contamination. The sensitivity and specificity of these tests need to be demonstrated. When PCR is  
89 used, the manufacturer should be able to clarify whether or not any nucleic acid detected  
90 originates from infectious particles.

91 *In vitro* tests can be used either as first choice or confirmatory tests to detect BVDV contamination  
92 of finished products. When BVDV contamination is suspected and PCR is used for the diagnosis, it  
93 is preferable to use a semi-quantitative RT-PCR with an internal control, which will allow  
94 quantification of the number of detected BVDV RNA copies in each sample. This should already  
95 allow discrimination between background signals and serious BVDV contamination. In order to  
96 determine if the detected sequences are the signals of infectious or non-infectious particles, a  
97 confirmatory *in vitro* test has to be carried out on the original sample.

98 As an example of a first choice or confirmatory (for PCR) diagnostic method to detect BVDV  
99 contamination, an *in vitro* test can be carried out as follows.

100 After neutralisation of the active ingredient(s) of the vaccine (in the case of live vaccines) with  
101 mono specific antiserum (free of BVD antibodies), the finished product has to be inoculated into  
102 sensitive cells, free of BVDV.

103 It is advisable to incubate as many wells as possible (such as twenty wells on a 24 wells microtitre  
104 plate), and perform at least three passages of the cells inoculated. Monolayers are observed for the  
105 appearance of cellular changes caused by replicating cytopathogenic strains of BVDV. The  
106 presence of non-cytopathogenic strains of BVDV should be screened by using an  
107 immunoperoxidase monolayer/linked assay (IPMA/IPLA) or an immunofluorescent assay or PCR.

## 108 **6. Measures to be taken**

109 Measures have to be taken (alert, withdrawal of the contaminated batch) only when positive  
110 results are obtained with the *in vitro* test.

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