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4 **Guideline on the conduct of efficacy studies for**
5 **intramammary products for use in cattle**
6 **Draft**

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7 This guideline replaces the CVMP guideline "Conduct of efficacy studies for intramammary products for
8 use in cattle" ([CVMP/344/1999-Rev.1](#))

Comments should be provided using this [template](#). The completed comments form should be sent to vet-guidelines@ema.europa.eu.



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45 **Executive summary**

46 This revised guideline is intended to provide guidance on the conduct of efficacy studies and their
47 evaluation for veterinary medicinal products that are administered via the teat canal to cattle. It
48 therefore addresses the treatment of clinical and subclinical mastitis during the lactation period, the
49 treatment of subclinical mastitis at drying off, and the prevention of new intramammary infections
50 during the dry period. The scope of the guideline has been extended in order to include
51 recommendations on pre-clinical data, in addition to those on clinical field studies for the
52 demonstration of efficacy. Moreover, information is included for generic intramammary products.

53 **1. Introduction (background)**

54 This guideline addresses data requirements for demonstrating pre-clinical and clinical efficacy of
55 products for intramammary use in cattle.

56 The majority of products for treatment and prevention of intramammary infections contain
57 antimicrobial substances, and the recommendations in this guideline focus on such products and their
58 use. It is recognised that acceptable methods other than those referred to in this guideline might be
59 capable of providing adequate information, provided they are sufficiently justified.

60 Since the principles for demonstrating clinical efficacy of a product intended for treatment and/or
61 prevention of intramammary infections are the same for antimicrobials as for other types of
62 substances, recommendations made in this guideline also apply to intramammary products containing
63 other types of active substances.

64 SPC recommendations made for the use of intramammary products should be evidence-based,
65 meaning a rationale with respect to active substance, dose, frequency of administration and treatment
66 length should be given, and the anticipated efficacy of the product should be demonstrated and
67 confirmed by appropriate pre-clinical and clinical studies.

68 **2. Scope**

69 This guideline is intended to provide guidance on design, conduct and reporting of pre-clinical and
70 clinical studies submitted in support of a new application for a marketing authorisation for a product for
71 intramammary use in dairy cattle, or to vary the conditions for use of an already authorised product.

72 Recommendations concern intramammary products for use during lactation and at drying off. This
73 guideline also includes recommendations for generics of authorised intramammary products.

74 For intramammary products containing antimicrobial substances, recommendations made in the
75 guideline for the Demonstration of Efficacy for Veterinary Medicinal Products containing Antimicrobial
76 Substances (EMA/CVMP/627/2001) and in the guideline for the conduct of pharmacokinetic studies in
77 target animal species (EMA/CVMP/133/1999-final) apply, where relevant. As appropriate the
78 Guidance on pre-approval information for registration of new veterinary medicinal products for food
79 producing animals with respect to antimicrobial resistance (CVMP/VICH/644/01-Final, 2004) should be
80 considered. With regard to tolerance please see VICH GL 43 guideline on target animal safety for
81 veterinary pharmaceutical products (CVMP/VICH/393388/2006) and the guideline (on) local tolerance
82 of intramammary preparations in cows (7AE21a, Volume 7, 1993).

83 **3. Legal basis**

84 This guideline replaces the current CVMP guideline for the conduct of efficacy studies for
85 intramammary products for use in cattle (CVMP/344/99- final-rev.1) and should be read in conjunction
86 with Directive 2001/82, as amended. Furthermore, in accordance with the provisions of the European
87 Convention for the Protection of Vertebrate Animals Used for Experimental and Other Scientific
88 Purposes and Directive 2010/63/EU on protection of animals used for scientific purposes, the 3R
89 principles (replacement, reduction and refinement) should be applied whenever possible.

90 Applicants should also refer to other relevant European and VICH guidelines, including those listed in
91 the reference list of this document.

92 **4. Pharmacology**

93 For the demonstration of pharmacodynamic properties and pharmacokinetics also the Guideline for the
94 demonstration of efficacy for veterinary medicinal products containing antimicrobial substances
95 (EMA/CVMP/ 261180/2012-rev) should be considered, as appropriate.

96 **4.1. Pharmacodynamic properties**

97 Studies on pharmacodynamics should be performed according to validated and/or internationally
98 accepted methods, if available.

99 As a general rule, the mode and mechanism of action underlying the desired therapeutic effect(s) of
100 the active substance(s) should be described, and any possible secondary effects relevant for the target
101 species/indication should be reported. Furthermore, the influence of milk on the pharmacological
102 activity of the active substance(s) should be investigated, where appropriate.

103 Studies may include *in vitro* and/or *in vivo* designs. The experimental design employed and the method
104 of measuring the pharmacodynamic effect should be fully described by the applicant, unless they are
105 known as standard procedures.

106 **4.2. Pharmacokinetics**

107 For lactating cow products, the concentration of the active substance(s) in plasma as a function of time
108 should be determined to investigate the potential systemic absorption. Furthermore, the concentration
109 of the active substance(s) in milk as a function of time should be investigated to allow an estimation of
110 the therapeutic concentration-time profile at the infection sites in the udder.

111 For dry cow products, the concentration time profile in plasma should be investigated in order to
112 determine the extent of systemic absorption.

113 In addition, factors like release of the active substance(s) from the formulation, and the physico-
114 chemical properties of the active substance(s) and the excipients should be considered, as these may
115 have influence on the availability of the product in the milk or dry udder secretion, as well as in udder
116 tissue. In this respect parameters like composition, particle size distribution, viscosity and dissolution
117 in milk should be discussed with regard to the claimed indication.

118 **5. Clinical studies**

119 It is recommended to conduct clinical studies according to Good Laboratory Practice (GLP) and/or Good
120 Clinical Practice (GCP). In case GLP and/or GCP are not applied, traceability and integrity of data
121 should be adequately guaranteed by other means. For clinical field trials, GCP status is required.

122 **5.1. Dose selection principles**

123 The principles of dose selection aim at finding an optimal dose and dosing regimen, taking the target
124 pathogen species into account and minimising the risk for development of resistance.

125 When selecting the appropriate dosing regimen the following aspects should be considered:

- 126 • quantity/activity of the active substance(s) and volume of the product, administered to a single
127 quarter,
- 128 • number of administrations per day (dosing interval),
- 129 • number of administrations needed to achieve complete cure (duration of treatment).

130 The rationale for the dosing regimen should be provided. Published literature on the disease may be
131 used as supportive information.

132 **5.2. Dose determination studies**

133 Dose determination studies should be performed with the final formulation of the test product where
134 possible.

135 For defining the target dose usually three dose levels need to be tested. Dose determination studies
136 should always include a negative control, which consequently requires the implementation of an
137 adequate rescue protocol for animal welfare reasons. With regard to clinical and subclinical mastitis
138 during lactation, investigation of different treatment durations is recommended in order to identify an
139 optimal dosing strategy. Dosing intervals should be aligned with usual milking intervals.

140 Dose determination studies could be performed either in naturally or experimentally infected cows
141 preferably under controlled conditions. In the absence of experimental models for dry cow therapy
142 dose determination should be conducted under field conditions.

143 **5.2.1. Experimental studies in lactating cows**

144 In lactating cows, dose determination should be studied under controlled conditions in experimentally
145 infected animals using suitable and well documented models.

146 The experimental infection should be performed with an udder pathogen which is relevant for the
147 claimed indication, and which can induce a disease pattern of clinical and/or subclinical mastitis similar
148 to natural infection. Information with regard to origin and *in vitro* susceptibility of the challenge strain
149 of the target pathogen to the proposed substance(s) should be provided. The choice of the challenge
150 strain should be justified. The design of an experimental study (e.g. time point for initiation of
151 treatment, sampling procedure, observation period, efficacy criteria etc.) should mimic field conditions.
152 If an experimental infection study is not feasible, dose determination may also be performed in
153 naturally infected animals.

154 **5.3. Dose confirmation**

155 Confirmation of the selected dosing regimen should be performed with the final formulation, preferably
156 in naturally infected animals. The evaluation can be performed under field conditions or under well-
157 controlled clinical conditions (e.g. laboratory conditions). It may also be appropriate to use dose
158 confirmation studies to investigate different treatment durations if this cannot be explored in dose
159 determination studies.

160 Preferably the study should include a negative control group (see also 7.1); this may require
161 appropriate measures with regard to animal welfare. Where study conditions do not allow inclusion of a
162 negative control group (e.g. in clinical mastitis cases with low spontaneous cure rates) it may be
163 acceptable to use a suitable positive control. In such a case the internal validity of the study needs to
164 be ensured.

165 Dose confirmation studies may be waived in circumstances where dose-finding data are available that
166 provide convincing support that the selected dosing regimen is appropriate for the treatment of
167 naturally occurring infections. This option requires all the following criteria to be fulfilled: the conditions
168 of the dose determination study and the susceptibility of any challenge strain are representative of the
169 field situation; a clear dose effect relationship is documented by dose determination data which allows
170 the selection of one appropriate dose; the dosing interval and the number of administrations is
171 adequately justified.

172 **5.4. Field studies**

173 **5.4.1. General considerations**

174 Field studies should be carried out to confirm the efficacy (and target animal safety) of the test product
175 at the selected dosage regimen under practical conditions. The final formulation of the test product
176 should be used.

177 The studies should be multicentric and representative for European conditions, taking into account
178 differences in animal husbandry systems, geographical location and climate, and they should be
179 performed in line with GCP. Appropriate statistical methods should be applied (see CVMP guideline on
180 statistical principles for veterinary clinical trials, CVMP/EWP/81976/2010).

181 **5.4.2. Study design and population**

182 Field studies should be blinded (whenever feasible), controlled and animals should be allocated
183 randomly to test and control groups. The details of the blinding method used should be provided.

184 The number of cows selected from a single herd should not exceed 20% of the total number of cases
185 included in the complete study to avoid that treatment outcome evaluation is dominated by the results
186 in one single herd.

187 The study should be designed so as to ensure that blinding is not jeopardised in circumstances where
188 the withdrawal periods differ between test and control treatment. The **positive control** should be an
189 intramammary product with the same indications as the test product and should be authorised in
190 accordance with Council Directive 2001/82/EC as amended. The applicant should justify the choice of
191 the positive control in relation to the indication and the target population for treatment. Products for
192 which recent susceptibility data suggest that posology may be inadequate for the infection under

193 study, or products where posology differs between Member States should be avoided. In the absence
194 of a suitable positive control the applicant should seek scientific advice from the authorities.

195 A **negative control** is considered necessary for demonstration of efficacy for preventive treatments at
196 drying off, implying that an untreated group of cows with non-infected animals/quarters needs to be
197 included. Comparison with a negative control is also considered necessary for infections with a high
198 spontaneous cure rate (e.g. some subclinical infections, *Escherichia coli* infections in lactating cows),
199 since a non-inferiority study design is unlikely to yield conclusive information for this situation.
200 Appropriate measures with regard to animal welfare should be taken into account.

201 The choice of the type of control should be justified by the applicant.

202 **5.4.3. Pathogens**

203 A claim for efficacy should be demonstrated for each target pathogen separately or for a target
204 pathogen group if scientifically justified (e.g. Coagulase-negative staphylococci). The choice of the
205 claimed pathogens should be justified with regard to the intended use of the product (either during
206 lactation or at drying off), and with regard to the spectrum of activity of the substance under study. In
207 general, the clinical study should be sufficiently powered to demonstrate a statistically significant effect
208 for each claimed bacteria species separately. For pathogens less common in the field, it may be
209 difficult to recruit sufficient cases. In such a situation, a lower number of cases may be justifiable
210 provided the overall data base can support conclusions on efficacy.

211 **5.4.4. Bacteriological diagnostic procedures**

212 Milk sampling and microbiological investigations should be carried out in accordance with standard (or
213 accepted) methods, for example, those recommended by the National Mastitis Council or by other
214 adequate references.

215 For recruitment of cows with subclinical mastitis, bacteriological examinations of milk samples should
216 be performed from all udder quarters of any cow in order to meet the inclusion criteria. In case of
217 clinical mastitis, pre-treatment bacteriological examination can be performed from the affected udder
218 quarter only, based on clinical signs. After treatment, bacteriological examinations of milk samples
219 should be performed from all included quarters.

220 For mastitis pathogens isolated from pre-treatment milk samples, *in vitro* antimicrobial susceptibility to
221 the antimicrobial(s) used should be determined according to recognized procedures. For animals which
222 are classed as clinical failures, susceptibility testing should be performed as well.

223 **5.4.5. Relevant parameters for efficacy evaluation**

224 **Bacteriological status**

225 Bacteriological status is the primary parameter for evaluating success of treatment and should be
226 evaluated for each included udder quarter. Only cases of clinical and subclinical mastitis in which the
227 claimed target pathogens are isolated in the pre-treatment sample should be used in calculating cure
228 rates.

229 **Clinical status**

230 In clinical mastitis cases the clinical cure is the co-primary parameter. The clinical cure should be
231 evaluated for each infected quarter and based on the return to normal of the parameters concerning
232 the cow's general condition, the appearance of the milk and the local clinical signs of the udder.

233 **Somatic cell counts (SCC)**

234 In clinical and subclinical mastitis trials, individual quarter milk SCC is determined from one pre-
235 treatment sample and from the second post-treatment sample. The same applies to cases in which
236 prevention of new infections during the dry period is studied. Mean SCCs are calculated from the
237 results for each treatment group and – in case of clinical and subclinical mastitis - separately for
238 bacteriologically cured and not cured quarters unless otherwise indicated. The SCC results for each
239 treatment group may be used as a secondary endpoint.

240 **5.4.6. Herd and cow information**

241 Study cows should be selected from herds with proper cow identification and health records. To the
242 extent possible, the history of the herd and cows should be recorded after the inclusion of a cow in the
243 trial and before the commencement of the treatment.

244 Farm:

- 245 • Name and address or farm code and district/region of herd owner;
- 246 • Location of the herd;
- 247 • Number of dairy cows;
- 248 • Methods of herd management, milking, and dry cow management;
- 249 • Teat disinfection procedures if practised;
- 250 • Bulk milk SCC in the herd over preceding months.

251 Cows:

- 252 • Name or identification number;
- 253 • Breed;
- 254 • Number of lactations;
- 255 • Date of calving;
- 256 • Estimated or measured milk yield at time of treatment;
- 257 • Cow milk SCC during preceding months;
- 258 • History of previous mastitis treatments;
- 259 • In clinical mastitis: carefully recorded clinical signs at the time of treatment;
- 260 • In dry cow treatment: the milk yields of cows at drying off and the method of drying off.

261 **5.4.7 Inclusion criteria**

262 With regard to inclusion criteria, please refer to the following sections which address special
263 considerations for the respective indications:

- 264 Section 5.4.9: Special considerations for clinical mastitis in lactating cows.
- 265 Section 5.4.10: Special considerations for subclinical mastitis in lactating cows.
- 266 Section 5.4.11: Special considerations for subclinical mastitis at drying off and prevention of
267 new infections during the dry period.

268 **5.4.8 Exclusion criteria**

269 The following cows are to be excluded from the trial:

- 270 • Cows with concurrent disease;
- 271 • Cows given systemic or intramammary anti-infectious or anti-inflammatory treatments within a
272 30-day period before the trial;
- 273 • Cows treated with products inducing an immune-mediated response against mastitis
274 pathogens.
- 275 • Cows with visible teat damage;
- 276 • In clinical mastitis: cows with severe systemic clinical signs requiring systemic treatment;
- 277 • In clinical mastitis: cows with clinical signs of mastitis in two or more udder quarters;
- 278 • In subclinical mastitis: cows with signs of subclinical mastitis in two or more udder quarters;
- 279 • In clinical and subclinical mastitis: cows with a daily milk yield less than 5 litres of milk prior to
280 onset of clinical signs.

281 **5.4.9 Special considerations for clinical mastitis in lactating cows**

282 **Treatment unit**

283 In clinical mastitis the treatment unit and the statistical unit is the individual udder quarter.

284 **Inclusion criteria**

285 In clinical mastitis trials, all lactating cows with clinical mastitis limited to 1 quarter which can be
286 treated with intramammary treatment only are eligible. The pre-treatment milk sample should be
287 bacteriologically positive regarding the target pathogen(s) as claimed.

288 **Pre-treatment sampling**

289 Before treatment one milk sample from the affected udder quarter should be taken for bacteriological
290 analysis and determination of quarter milk SCC and the cow should be clinically examined (general
291 condition, appearance of milk, udder consistency).

292 **Treatment**

293 In any included cow only the single affected quarter will be treated. A cow developing clinical mastitis
294 in additional quarters during the experimental period should be withdrawn from the study post
295 inclusion (please, refer to section 5.4.12). With regard to controls please refer to section 5.4.2. In
296 addition clinical examination should be made when considered necessary

297 **Post-treatment sampling**

298 After treatment two milk samples should be taken for bacteriological analysis. These samples should
299 be taken between day 14 and day 28 from the last treatment, at least 7 days apart. Clinical
300 examination should be performed at the first bacteriological post treatment sampling. If clinical cure
301 has not been achieved by this sampling time point, the case should be excluded from further sampling
302 (see below for assessment of success/failure). Quarter milk SCC should be determined from the second

303 post-treatment sample meaning that only cows with clinical cure at the first post treatment sampling
304 are concerned.

305 **Assessment of success/failure**

306 Cases of success and failure which are to be included in the final data analysis:

307 A case is regarded a treatment success if there is clinical cure at the first post-treatment sampling
308 (normal appearance of the milk, normal condition of the udder, normal general condition) as well as
309 bacteriological cure in both post-treatment milk samples (absence of the udder pathogen species which
310 was present at the time of inclusion).

311 Cows with new infections in the originally infected, treated quarter (i.e. detection of an udder pathogen
312 which is different from that isolated at inclusion in one or both post-treatment milk samples) can be
313 classified as a bacteriological cure for the original pathogen. The number and type of new infections in
314 each treatment group should be included in the final study report. A high frequency of these
315 occurrences requires a thorough analysis.

316 A case is regarded a failure

- 317 • If the criteria for clinical cure are not met in the clinical examination at the first post-treatment
318 sampling (the cow should then have been excluded from further sampling).
- 319 • If the original pathogen detected at the time of inclusion is present in one or both post-
320 treatment samples.
- 321 • If additional antimicrobial treatment associated with the mastitis case enrolled is necessary
322 during the study period.

323 **5.4.10 Special considerations for subclinical mastitis in lactating cows**

324 **Treatment unit**

325 In subclinical mastitis during lactation, the treatment unit and the statistical unit is the individual
326 quarter.

327 **Inclusion criteria**

328 In subclinical mastitis trials, all lactating cows with the presence of the same target pathogen(s) in two
329 pre-treatment milk samples in conjunction with elevated quarter somatic cell count (SCC) > 200 000
330 cells/ml in one pre-treatment milk sample are eligible for a study. Only cows with 1 sub-clinically
331 infected quarter should be included.

332 **Pre-treatment sampling**

333 Before treatment two quarter milk samples from all udder quarters should be taken one to three days
334 apart for bacteriological analysis; if a pathogen can only be isolated from one out of these two
335 samples, diagnosis should be confirmed with a third sample. Quarter milk SCC should be determined
336 from one of the pre-treatment samples.

337 **Treatment**

338 In any included cow only the single confirmed positive quarter will be treated. With regard to controls,
339 please refer to section 5.4.2.

340 **Post-treatment sampling**

341 After treatment two milk samples should be taken for bacteriological analysis. These samples should
342 be taken between day 14 and day 28 from the cessation of the treatment from the included quarter,
343 and should be separated by a period of at least 7 days. Quarter milk SCC should be determined from
344 the second post-treatment sample.

345 **Assessment of success/failure**

346 Cases of success and failure which are to be included in the final data analysis:

347 A case is regarded a treatment success if the original pathogen is not detected in either of the post-
348 treatment milk samples. A marked decrease in the somatic cell count is considered supportive.

349 With regard to new infections the same evaluation as defined for clinical mastitis cases will apply.

350 A case is regarded a failure

- 351 • If the original pathogen detected at the time of inclusion is present in one or both post-
352 treatment samples.
- 353 • If additional antimicrobial treatment associated with the subclinical mastitis is necessary during
354 the experimental period.

355 **5.4.11 Special considerations for subclinical mastitis at drying off and**
356 **prevention of new infections during the dry period**

357 Both treatment of subclinical infections at drying off and prevention of new infections during the dry
358 period can be studied in the same animal; however, treatment and prevention should not be studied in
359 the same quarter.

360 **Treatment unit**

361 For dry cow treatment, the treatment unit is the cow but the statistical unit is the individual quarter.

362 **Inclusion criteria**

363 For dry cow treatment, lactating cows which are approaching the end of lactation and ready for drying-
364 off are eligible for the trial.

365 Cows with subclinically infected quarters (presence of the same target pathogen(s) in two pre-
366 treatment milk samples, SCC > 200 000 cells/ml in one of these samples) are eligible for studying
367 treatment effect on subclinical infections.

368 For assessment of prevention of new infections during the dry period, only non-infected healthy
369 quarters are eligible at drying-off. Two pre-treatment milk samples should be bacteriologically negative
370 and SCC values, examined in one of these samples, should be < 200 000 cells/ml.

371 Only animals with dry periods of sufficient length (approximately 35 days or more) should be included.

372 **Pre-treatment sampling**

373 Within one week prior to drying-off, two pre-treatment quarter milk samples should be taken one to
374 three days apart from all quarters for bacteriological analysis. For subclinical mastitis cases in which a
375 pathogen can only be isolated from one out of two milk samples, a third sample may be necessary for
376 confirmation of diagnosis (see also inclusion criteria).

377 The same sampling strategy applies with respect to prevention of new infections during the dry period.
378 In cases where only one out of two pre-treatment milk samples is free of pathogens, a third sample is
379 needed to confirm the diagnosis.

380 Quarter milk SCC should be determined from one of the pre-treatment samples.

381 **Treatment**

382 At drying-off, all four quarters of animals should be treated. This may include treatment of infected
383 and non-infected quarters of one cow. With regard to controls please refer to section 5.4.2.

384 **Post-treatment sampling**

385 After calving two post-treatment milk samples should be taken for bacteriological analysis. The first
386 milk sample should be taken before the first regular milking after calving following the colostrum
387 stage, and the second post-treatment sample 4 to 7 days later. Quarter milk SCC should be
388 determined from the second post-treatment sample.

389 In addition the cow should be clinically examined after calving at appropriate times and intervals, for
390 any pathological changes of the udder or of the appearance of the milk.

391 **Assessment of success/failure**

392 Cases of success and failure which are to be included in the final data analysis:

393 Subclinical mastitis

394 A case is regarded a treatment success if the original pathogen is not detected in either of the two
395 post-treatment milk samples.

396 With regard to new infections the same evaluation as defined for clinical mastitis cases will apply.

397 A case is regarded a failure

- 398 • If the original pathogen detected at the time of inclusion is present in one or both post-
399 treatment samples.
- 400 • If additional antimicrobial treatment associated with mastitis is necessary during the
401 experimental period.

402 Prevention of new infections

403 A case is regarded a prevention success if no udder pathogens can be detected in either of the post-
404 treatment milk samples after calving.

405 A case is regarded a prevention failure

- 406 • If any target udder pathogen can be detected in either or both post-treatment milk samples
407 (corresponding to a new infection).

- 408 • If additional antimicrobial treatment related to mastitis is necessary during the study period.

409 **5.4.12 Withdrawals**

410 Animals/quarters which are to be excluded from the final data analysis should be recorded as follows:

- 411 • Cases which are not interpretable due to lack or loss of information (e.g. quarters with no
412 pathogens in the pre-treatment samples, contaminated pre-treatment milk samples) shall be
413 listed in the final report, and their distribution in each group shall be analysed.
- 414 • Data from cows with clinical mastitis in which additional quarters had to be treated during the
415 experimental period shall be excluded from the final analysis and listed separately for each
416 treatment group. The reasons and the potential impact of the withdrawals on the study results
417 should be discussed.
- 418 • Cows treated with antibiotics due to intercurrent diseases during the experimental period
419 should be excluded from the trial and indicated in the final report.
- 420 • Any other cases in which the exclusion from final data analysis is justified should be indicated as
421 well.

422 **5.4.13 Presentation of data - reporting**

423 A record from each individual case should be presented in the dossier. The data on the bacteriological
424 results and the bacteriological response for each organism for each treated quarter should be
425 summarized and tabulated separately for each bacterial species and treatment group. *In vitro*
426 susceptibility results should be enclosed in the dossier.

427 As appropriate, the data should be expressed as number of quarters and number of cows cured
428 clinically, bacteriologically, and based on individual quarter milk SCC (subclinical mastitis only), see
429 table 1 and 2 as examples.

430 For subclinical mastitis studies, it is preferable to present combined cure rates based on individual
431 quarter data (bacteriological cure + quarter milk SCC < 200 000 cells/ml).

432 *Table 1.* An example for data presentation for each treatment group in clinical mastitis (further details
433 are given in the text).

		Post-treatment cure in clinical mastitis		
		Clinical cure	Bacteriological cure	Bacteriological + clinical cure
Treatment groups	No of quarters	n %	n %	n %
Test product				
Positive control				
and/or				
Negative control				

434

435

436 Table 2. An example for data presentation for each treatment group in subclinical mastitis (further
 437 details are given in the text).

		Post-treatment cure in subclinical mastitis	
		Bacteriological cure	Bacteriological cure + SCC < 200 00 cells/ml
Treatment groups	No of quarters or - at drying off -number of quarters/cows	n %	n %
Test product			
Positive control			
and/or			
Negative control			

438 Cases of clinical mastitis occurring during the dry period and during the post-calving investigational
 439 period should be recorded.

440 6. Generic products – data requirements

441 The overarching principle is that generics of intramammary products should be therapeutically
 442 equivalent to an originator, the reference product being a product with a complete documentation for
 443 marketing authorisation. However, the Guideline “Conduct of bioequivalence studies for veterinary
 444 medicinal products (CVMP/016/2000/Rev. 2)” is not applicable for locally acting products such as
 445 intramammary products. In consequence, Art. 13 (3) of the Directive 2001/82/EC as amended applies,
 446 i.e. data demonstrating the efficacy should be provided. In such cases comparable efficacy between
 447 test and reference product should be demonstrated by an appropriate clinical trial, e.g. by a non-
 448 inferiority field study.

449 Differences in product formulation may influence penetration and distribution of the active substance in
 450 the mastitic udder. Taking into account the different locations of mastitis pathogens, it may therefore
 451 not be possible to predict that efficacy of a generic product will be non-inferior for all target pathogens
 452 based only on efficacy for the pathogen that is the least susceptible *in vitro*. Therefore, in order to gain
 453 all the claims for the reference product, the study should be conducted using the target pathogen that
 454 is justified as the most difficult to treat *in vivo* based on pharmacokinetic properties, pathophysiological
 455 characteristics and susceptibility of the target pathogen(s), as appropriate. The parameters for
 456 evaluation of efficacy (and tolerance) in field trials apply. It is recognised that large numbers of cases
 457 will be required to satisfy statistical requirements. If adequate safety parameters are also recorded in
 458 the clinical trial, it may be possible to waive a dedicated local tolerance study.

459 Efficacy and tolerance studies may be waived for generic products that are fully identical to the
 460 reference product.

461 Efficacy studies may also be waived if the following conditions are fulfilled: the generic product has the
 462 same pharmaceutical form and contains qualitatively and quantitatively the same active substance(s)
 463 (same salts), the excipients of the generic are qualitatively and quantitatively very similar compared to
 464 the reference product, and the physicochemical properties (e.g. crystalline form, particle size
 465 distribution, viscosity, relative density, dissolution profile) of the generic product are similar to those of
 466 the reference product (please, see Annex). Local tolerance data might be requested.

467

468 **Definitions**

469 **Mastitis:** Inflammation of one or more quarters of the mammary gland, almost always caused by
470 infecting microorganism(s).

471 **Clinical mastitis:** Clinical signs in one or more quarters (swelling, heat, pain, redness) and/or
472 changes in the appearance of milk (clots or flakes, watery appearance, discoloration), with or without
473 general signs (fever, loss of appetite).

474 **Subclinical mastitis:** Elevated milk somatic cell count in a quarter and isolation of an udder pathogen
475 from the milk, but no clinical signs.

476 **New infection:** Isolation of a pathogen from a mammary gland that has not previously been isolated
477 from that mammary gland or has not been isolated for some predetermined period of time.

478 **References**

479 CVMP guideline: Demonstration of efficacy for veterinary medicinal products containing antimicrobial
480 substances (EMA/CVMP/627/2001) – *currently under revision*

481 CVMP guideline on fixed combination products (EMA/CVMP/83804/2005)

482 CVMP guideline on statistical principles for veterinary clinical trials for veterinary medicinal products
483 (pharmaceuticals) (CVMP/EWP/81976/2010)

484 CVMP guideline on Conduct of pharmacokinetic studies in target animal species
485 (EMA/CVMP/133/1999)

486 Directive 2010/63/EU of the European Parliament and of the Council on the protection of animals used
487 for scientific purposes. Official Journal of the European Union L 276, 20/10/2010, p. 33-79

488 Good Laboratory Practice (GLP) (see Council Directive 88/320/EEC as amended)

489 Local tolerance of intramammary preparations in cows (7AE21a Volume 7)

490 VICH GL9: Guideline on good clinical practices (CVMP/VICH/595/1998)

491 VICH GL43: Guideline on target animal safety for veterinary pharmaceutical products
492 (CVMP/VICH/393388/2006), section 3.4. Mammary Gland Safety Studies

493

494 **ANNEX – Biowaivers for intramammary generics**

495 **I Introduction**

496 Efficacy studies for intramammary products may be waived if generic and reference product are very
497 similar, that is, if the generic product has the same pharmaceutical form and contains qualitatively and
498 quantitatively the same active substance(s) (same salt), the excipients of the generic are qualitatively
499 and quantitatively very similar compared to the reference product, and the physicochemical properties
500 (e.g. crystalline form, particle size distribution, viscosity, relative density, dissolution profile) of the
501 generic product are similar to those of the reference product.

502 This annex explains what quality requirements for applications for generic intramammary products
503 could be provided to demonstrate that the generic and referenced /originator product are very similar
504 in order to provide a biowaiver.

505 It is noted that the annex only refers to quality requirements and not to any *in vivo*/efficacy testing. It
506 should also be noted that extrapolation of withdrawal periods between products was not considered.

507 **II Summary Requirements**

508 Generally, biowaivers can only be granted on a case by case basis and when justified by the
509 appropriate supporting data.

510 **IN VITRO TESTS**

511 The following tests may be appropriate to demonstrate similarity of the products but is not an
512 exhaustive list. Some tests might not be relevant depending on the pharmaceutical form of the
513 intramammary preparations (e.g. solutions).

514 Selection of the tests to establish formulation similarity should be justified.

515 **Composition**

516 Investigational analytical studies should be presented in order to establish that the generic product has
517 an identical or very similar qualitative and quantitative formulation as the reference product.

518 Due consideration should be given to the grade of excipients and the properties, e.g. rheological that
519 they impart and whether or not these could influence release of the active substance from the
520 formulation. Where the properties of the product formulation could be influenced, these should be
521 investigated during pharmaceutical development and criteria established on the specification of the
522 excipient to control the relevant parameter.

523 **Crystalline form**

524 Data should be presented to demonstrate that the same crystalline form(s) of the active substance(s)
525 is used in the generic and reference products.

526 If there is more than one active substance in the product then the crystalline form of each active
527 substance should be investigated separately.

528 **Pharmaceutical form**

529 The pharmaceutical form should be the same, and the appearance of the generic and the reference
530 products should be similar.

531 **Particle size distribution**

532 Data should be provided to demonstrate that the generic and the reference product are similar in
533 terms of particle size distribution of the active substance(s) and, if relevant, the excipients.

534 If there is more than one active substance in the product then each active substance should be
535 considered separately. In case of excipients not dissolved, the particle size of these should also be
536 considered.

537 **Viscosity**

538 The viscosity of the products should be measured over a justified temperature range, including the
539 physiological temperature of the target species. The rheological profiles of the generic and reference
540 products should be similar.

541 **Relative density**

542 Data should be provided demonstrating that the generic and the reference products are similar in
543 terms of relative density.

544 ***In vitro* dissolution test**

545 *In vitro* dissolution studies may be used to provide evidence of the similarity of the quality of the
546 generic and reference products.

547 The *f*₂ statistic is widely used for comparison of dissolution profiles, but may not be appropriate in all
548 cases. When the *f*₂ statistic is not suitable, then the similarity may be compared using model-
549 dependent or model-independent methods e.g. by statistical multivariate comparison of the
550 parameters of the Weibull function or the percentage dissolved at different time points.

551 Alternative methods to the *f*₂ statistic to demonstrate dissolution similarity are considered acceptable,
552 if statistically valid and satisfactorily justified.

553 The similarity acceptance limits should be pre-defined and justified and not be greater than a 10%
554 difference. In addition, the dissolution variability of the test and reference product data should also be
555 similar, however, a lower variability of the test product may be acceptable.

556 Evidence that the statistical software has been validated should also be provided.

557 A clear description and explanation of the steps taken in the application of the procedure should be
558 provided, with appropriate summary tables.

559 Lactating cow products

560 Investigations should demonstrate that the generic and reference products have a similar *in vitro*
561 dissolution. Comparative *in vitro* dissolution experiments should follow current compendial standards
562 and a thorough description of experimental settings and analytical methods should be provided. This
563 should include a study protocol, batch information of the generic and reference batches, detailed
564 experimental conditions, validation of experimental methods, individual and mean results and if
565 necessary respective summary statistics. It is recommended to use 12 units of the product for each
566 experiment to enable statistical evaluation. Dissolution profiles should be compared considering
567 physiologically relevant experimental temperatures and pHs and the profile should be characterised
568 using a sufficient number of timepoints. The use of surfactants should be avoided unless their use is
569 unavoidable. Where surfactants are required their concentration should be minimised.

570 Whilst it is acknowledged that determination of the similarity of dissolution profiles using the *f*₂
571 statistic is applicable to immediate release products, in the absence of other statistical tests it can be

572 useful as a quantitative measure of the similarity of the in-vitro dissolution profiles of the generic and
573 reference intramammary products.

574 When using this approach, a sufficient number of timepoints prior to the plateau of the dissolution
575 curve, must be used in the calculations.

576 Where more than 85% of the drug is dissolved within 15 minutes, dissolution profiles may be accepted
577 as similar without further mathematical calculation.

578 In case more than 85% is not dissolved at 15 minutes but within 30 minutes, at least three time points
579 are required: the first time point before 15 minutes, the second one at 15 minutes and the third time
580 point when the release is close to 85%. In these cases mathematical evaluation such as calculation of
581 similarity factor f_2 (see below) may be required to demonstrate comparable dissolution.

582 In case more than 85% is not dissolved within 30 minutes, more than three time points may be
583 required.

584 Dissolution similarity may be determined using the f_2 statistic as follows:

$$f_2 = 50 \cdot \log \left[\frac{100}{\sqrt{1 + \frac{\sum_{t=1}^n [R(t) - T(t)]^2}{n}}} \right]$$

585
586 In this equation f_2 is the similarity factor, n is the number of time points, $R(t)$ is the mean percent
587 reference drug dissolved at time t after initiation of the study; $T(t)$ is the mean percent test drug
588 dissolved at time t after initiation of the study. For both the reference and test formulations, percent
589 dissolution should be determined.

590 The evaluation of the similarity factor is based on the following conditions:

- 591
- A minimum of three time points (zero excluded)
 - The time points should be the same for the two formulations
 - Twelve individual values for every time point for each formulation
 - Not more than one mean value of > 85% dissolved for any of the formulations.
 - The relative standard deviation or coefficient of variation of any product should be less than
596 20% for the first point and less than 10% from second to last time point.

597 An f_2 value between 50 and 100 suggests that the two dissolution profiles are similar.

598 Dry cow products

599 This category of intramammary preparations is characterised by prolonged release profiles.

600 In this context reference is made to the chapters/monographs of the European Pharmacopoeia relating
601 to relevant test apparatus, test conditions and test requirements with respect to dissolution testing of
602 prolonged release dosage forms

603 In the absence of appropriate guidance on *in vitro* dissolution testing of dry cow preparations
604 applicants are in any case strongly recommended to seek scientific advice from the authorities.

605

606 **SIMILARITY BETWEEN THE FORMULATIONS**

607 The results of the above tests should be obtained with 3 different (at least pilot) batches of both the
608 generic and the reference products, unless otherwise justified. Methods used should be relevant and
609 appropriate. Where relevant, validation data of the test methods used should be provided.

610 The generic product batches used in the study should be representative of the product to be marketed
611 and this should be justified by the applicant.

612 The applicant should document how representative batches of the reference product have been
613 selected.

614 To consider the reference and the generic products very similar, the difference in results between the
615 reference and the generic product should not be greater than the variability of different batches of the
616 reference product.

617 If comparison studies are not considered conclusive to demonstrate the similarity of the formulations,
618 no biowaiver can be claimed, and efficacy studies should be provided in the dossier.