



1      13 May 2016  
2      EMA/332805/2016  
3      Committee for Medicinal Products for Human use (CHMP)  
4      Committee for Medicinal Products for Veterinary use (CVMP)  
5      Quality Working Party (QWP)

6      **Reflection paper on the dissolution specification for**  
7      **generic oral immediate release products**  
8      Draft

Draft agreed by the QWP	March 2016
Draft adopted by the CHMP for release for consultation	March 2016
Draft adopted by the CVMP for release for consultation	April 2016
Start of public consultation	13 May 2016
End of consultation (deadline for comments)	13 August 2016

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<b>Keywords</b>	Dissolution specification, Generic, Oral immediate release product
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## 37 **Introduction**

38 During the last few years the suitability of dissolution specifications has been discussed in marketing  
39 authorisation procedures. Some referrals concerning this topic have been raised through the CMD(h).

40 A decision tree is proposed to make the evaluation process more transparent. However there may be  
41 some drugs with very narrow therapeutic ranges or products where there is prior knowledge of critical  
42 dissolution behaviour (e.g. sublingual or orodispersible tablets with some buccal absorption), which still  
43 have to be evaluated on a case by case basis.

## 44 **Scope**

45 In the context of this reflection paper immediate release is identified as at least 75% of the active  
46 substance is dissolved within 45 minutes. This derives from the Ph. Eur. (5.17.1) recommendation for  
47 conventional release dosage forms.

48 This paper discusses the suitability of the dissolution method and the specifications for *in vitro*  
49 dissolution of orally administered generic drug products with immediate release characteristics. Where  
50 applicable, this reflection paper should be read in connection with the principles of relevant guidelines  
51 listed as references.

52 The dissolution specification should ensure batch to batch consistency and, ideally, signal potential  
53 problems with *in vivo* bioavailability.

54 This reflection paper does not discuss the dissolution tests in three different buffers required as  
55 complementary to bioequivalence studies, those tests required in support of biowaiver of strengths or  
56 BCS biowaiver as defined in 4.2.1 and 4.2.2 and Appendix III respectively of the (human) Guideline on  
57 the Investigation of Bioequivalence (CPMP/EWP/QWP/1401/98 Rev. 1 / Corr\*\*) or in the Guideline on  
58 the conduct of bioequivalence studies for veterinary medicinal products (EMA/CVMP/016/00-Rev.2).

## 59 **Definitions**

### 60 **Dissolution specification**

61 The dissolution specification is expressed in terms of the quantity ( $Q$ ) of active substance dissolved in a  
62 specified time, expressed as a percentage of the content stated on the product label.

### 63 **Discriminatory Power**

64 The discriminatory power is the ability of a test procedure to discriminate between batches with  
65 respect to critical process parameters and /or critical material attributes which may have an impact on  
66 the bioavailability. Ideally all non-bioequivalent batches should be detected by the *in vitro* dissolution  
67 test results.

### 68 **Biobatch**

69 Biobatch is a batch used in a bioavailability/bioequivalence study or in clinical testing. In the context of  
70 this Reflection Paper the biobatch is the batch of the applied product, which has been shown to be  
71 bioequivalent in a bioequivalence study of a generic vs. a reference drug product.

72 **Discussion**

73 **1. Test method**

74 **1.1. Development of dissolution method**

75 A dissolution procedure intended to be used as a routine control test for immediate release drug  
76 products should be robust, reproducible and discriminatory in order to assure a consistent product  
77 quality and to detect altered product quality attributes, which may affect the *in vivo* performance. For  
78 the development of such a dissolution procedure, the following aspects in particular should be  
79 considered:

- 80 • Selection of a suitable dissolution medium should be based on the physico-chemical characteristics  
81 of the active substance(s) and the intended dose range of the drug product to be tested. It should  
82 be ensured that sink conditions are met.
- 83 • In general, an aqueous medium should be used and the pH should first be evaluated in the  
84 physiological pH range. The addition of surfactants should be avoided. When surfactants are used,  
85 for instance to achieve sink conditions for poorly aqueous-soluble active substances, the type of  
86 surfactant should be justified. The concentration of the surfactant should be as low as possible and  
87 be justified by relevant solubility and dissolution data and an accompanying scientific discussion.
- 88 • The development of methods using the paddle apparatus should start with a stirring speed of 50  
89 rpm. Higher stirring speeds may be applied with an appropriate justification. A higher stirring  
90 speed may be justified by high variability of the results (e.g. > 20% RSD at time-points  $\leq 10$   
91 minutes, > 10% RSD in the later phase for a sample size of 12) observed at lower speed rates due  
92 to hydrodynamic effects (e.g. coning) or other factors (e.g. tablet sticking). However, it is known  
93 that methods with increased stirring speeds may be less discriminatory. Increasing the stirring  
94 speed at the expense of the discriminatory power simply to reduce variability of the results or to  
95 obtain complete dissolution in a shorter time should be avoided. An increase of the stirring speed  
96 may be considered in case of over-discriminatory conditions towards *in vivo* performance.  
97 However, in all cases the dissolution profiles at increased stirring speeds should have sufficient  
98 discriminatory power for drug product quality control.
- 99 • During development, the contribution of method parameters to the variability of the results should  
100 be investigated and reduced to a minimum.
- 101 • The discriminatory power should be discussed (see also section 1.2).

102 Further procedural recommendations on dissolution testing are provided in the European  
103 Pharmacopoeia.

104 **1.2. Test conditions and discriminatory power**

105 To allow extrapolation of the results of a bioequivalence study from the biobatch to commercial  
106 batches, it is necessary to have a suitable specification of the amount of active substance released at a  
107 specified time-point. The test conditions should be chosen to allow discrimination between batches with  
108 different *in vitro* release characteristics. In an optimal case the *in vitro* results can mimic the *in vivo*  
109 situation; the next best approach is to reproduce the rank order between batches and discrimination of  
110 batches with different quality attributes without knowing about the *in vivo* relevance of these  
111 differences. Both approaches may be used for routine batch control.

112 The suitability of the test conditions for routine batch testing should be demonstrated using batches  
113 with different quality attributes. To achieve this, batches with meaningful changes compared to the  
114 applied finished product should be manufactured. Such changes may relate to the quantitative  
115 formulation, input parameters and/or using slightly modified process parameters. Current knowledge  
116 of both the characteristics derived from the Biopharmaceutics Classification System (BCS) and the  
117 finished product must be taken into account when choosing the quality attributes to change. For  
118 instance, for a finished product where the *in vivo* absorption (rate and/or extent) is expected to be  
119 limited by solubility / intrinsic dissolution of the active substance, i.e. BCS 2 and 4, suitable quality  
120 attributes may be particle size of the active substance or other attributes that would have an impact on  
121 the *in vivo* dissolution. For a finished product where the *in vivo* absorption is expected to be limited by  
122 gastric emptying or intestinal permeability, i.e. containing BCS 1 or 3 class active substance with rapid  
123 or very rapid dissolution (refer to BE Guideline), suitable quality attributes may be factors in the  
124 formulation and/or manufacturing process that will have an impact on the disintegration of the finished  
125 product and significantly affect the rate of *in vitro* dissolution.

126 Changes to the composition of the drug product to create a "bad batch" should be covered by the  
127 proposed qualitative batch formula and only the proportions of the employed excipients might be  
128 changed. The complete omission of one or more specific excipients from the formulation (e.g. binder,  
129 disintegrant) is not supported. The dissolution test conditions should be able to detect these changes  
130 by setting a suitable specification.

131 However, for drug products containing a BCS class 1 or class 3 active substances with very high  
132 solubility over the physiological pH range, it may not always be possible to detect any differences in  
133 dissolution behaviour after meaningful changes in relevant formulation and/or manufacturing  
134 parameters have been made.

### 135 **1.2.1. Batches with different *in vivo* behaviour included in pharmaceutical 136 development**

137 In cases where several batches of the drug product have been tested during development *in vivo*  
138 leading to batches with acceptable pharmacokinetic parameters and those with non-acceptable  
139 pharmacokinetic parameters, dissolution test conditions should be chosen which allow discrimination  
140 between acceptable and non-acceptable batches by setting a suitable specification.

### 141 **1.2.2. Only batches with acceptable *in vivo* behaviour included in 142 pharmaceutical development**

143 Batches representing different *in vitro* dissolution profiles, derived from the defined manufacturing  
144 process by setting process parameters within the range of maximum variability expected from process  
145 validation studies, are so-called "side-batches". The dissolution profiles of the side-batches can be used  
146 to set a suitable dissolution specification, when bioequivalence with the reference product is  
147 demonstrated. If the batches with the extreme range of *in vitro* dissolution profiles (i.e., fastest and  
148 slowest) are found to be bioequivalent to the reference product, then future batches with dissolution  
149 profiles within this range are also expected to be bioequivalent (when using the same manufacturing  
150 process). Thus, a suitable specification may be set based on the *in vitro* dissolution profile of the side  
151 batch with the slowest dissolution, using the methodology described in section 2.

152 For a marketing authorisation application for a generic medicinal product, a bioequivalence study  
153 between a representative batch of the generic product (test) series versus the originator product on  
154 the market (reference) has to be performed. The acceptance criteria for bioequivalence are set for the

155 pharmacokinetic parameters AUC and  $C_{max}$ . The latter is a measure of dissolution speed *in vivo*; in case  
156 of the same AUC, a larger  $C_{max}$  indicates faster *in vivo* dissolution. In a bioequivalence study design a  
157 comparison of the dissolution profiles ( $n=12$ ) of test and reference products is required using the  
158 proposed test conditions of the generic drug product.

159 To estimate the discriminatory power of the dissolution test it may be helpful to look at the *in vivo* data  
160 (point estimates and the respective confidence intervals) of the bioequivalence study. Due to the  
161 acceptance criteria for bioequivalence the point estimates for  $C_{max}$  plus the respective 90% confidence  
162 interval of the generic product have to be between 80% and 125% of the  $C_{max}$  of the reference  
163 product. According to the equivalence rules (opposite to a superiority test with the objective of detect  
164 statistical significant differences) small differences without clinical relevance will be accepted as long  
165 the 90% confidence interval fulfils these criteria.

166 In such a case the rank order of the *in vivo* and *in vitro* results should be compared. If a test product  
167 with significantly larger  $C_{max}$  shows faster *in vitro* dissolution than the reference product, this may be  
168 used as an indicator for suitability of the chosen test conditions. The larger the difference of the *in vivo*  
169 point estimates is, the greater the chance that this difference may also be reflected *in vitro*. In case of  
170 an opposite rank order, i.e. a test product with significantly larger  $C_{max}$  shows slower *in vitro*  
171 dissolution behaviour or vice versa, the test conditions should be further optimised in order to reflect  
172 the *in vivo* trend.

### 173 **1.2.3. No batches with *in vivo* behaviour included in pharmaceutical 174 development**

175 In certain instances the need for a bioequivalence study is waived based on fulfilling the criteria of the  
176 so called BCS-biowaiver. In such instances there is no batch used in a bioavailability/bioequivalence  
177 study or in clinical testing (biobatch) and by analogy, the batch that has been shown to be equivalent  
178 with a reference product based on satisfactory *in vitro* discriminatory dissolution data in at least three  
179 different pH media is considered to be the test batch.

180 For more information see Appendix III in the Guideline on the Investigation of Bioequivalence  
181 (CPMP/EWP/QWP/1401/98 Rev. 1/ Corr\*\*) and /or Guideline on the conduct of bioequivalence studies  
182 for veterinary medicinal products (EMA/CVMP/016/00-Rev.2).

## 183 **2. Setting Specifications**

184 When the dissolution test conditions have been chosen a suitable dissolution specification should be  
185 set. The dissolution specification is defined by a  $Q$  value, i.e. mean value, at a given time point, which  
186 allows discrimination between acceptable and non-acceptable batches. Batch results showing  
187 compliance with stage  $S_1$ ,  $S_2$  and  $S_3$  (Ph. Eur. 2.9.3.) are acceptable. The specification should be set in  
188 such a way so that during routine manufacture and testing it would be expected that compliance with  
189  $S_2$  is attained.

190 Before setting the  $Q$  value, the time range allowing discrimination should be considered from the  
191 dissolution profile of the biobatch. Sampling time points should be sufficient to obtain a meaningful  
192 dissolution profile (c.f. human BE guideline, Appendix I).

193 To ensure that the results of the bioequivalence study may be extrapolated to the drug product  
194 administered to the patient, all commercial batches should show similar behaviour compared to the  
195 biobatch. The dissolution profile of the biobatch, using test conditions providing discriminatory power  
196 should be used to set a suitable specification. Similar dissolution of two batches may be assumed in

197 case of differences of less than 10% in their mean results. Therefore, the  $Q$  value is recommended to  
198 be set on the basis of the biobatch dissolution result (mean value) minus 10%.

199 The acceptance criteria the  $Q$  value is usually set in the range between 75-85% (5% intervals) to  
200 demonstrate discriminatory power and satisfactory dissolution. It is not considered relevant to have a  
201 limit of more than 85%. Usually the time points 15, 30 or 45 minutes would be sufficient, but other  
202 time points may be used if justified. It is not considered relevant to choose a time point before 15  
203 minutes.

204 The recommendations in Annex 1 are meant as guidance for setting the specification. The  
205 discriminatory power is closely linked to the time point and  $Q$  value chosen. If time points/ $Q$  values  
206 other than proposed in the decision tree would lead to discriminatory power, this is also acceptable.

207 ***How to read the recommendations in Annex 1:***

- 208 • If the dissolution of the biobatch is larger than or equal to 95% in 15 minutes, the specification  
209 may be set to  $Q=85\%$  after 15 minutes<sup>1</sup>;
- 210 • If the dissolution of the biobatch is less than 95% but larger than or equal to 85% in 15 minutes,  
211 the specification ( $Q$ ) may be set to 75%, 80% or 85% whichever is closer to  $Q=\text{biobatch result } -10\%$   
212 % at 15 minutes<sup>1</sup>;
- 213 • If dissolution of the biobatch is larger than or equal to 85% after 30 minutes, the specification ( $Q$ )  
214 may be set to 75%, 80% or 85% whichever is closer to  $Q=\text{biobatch result } -10\%$  at 30 minutes;
- 215 • If dissolution is larger than or equal to 85% after 45 minutes, the specification may be set to 75%,  
216 80% or 85% after 45 minutes.

217 In case dissolution of the biobatch is less than or equal to 85% after 45 minutes, a minimum of 75%  
218 after 45 minutes should be specified if possible. Otherwise, if the dissolution specification ( $Q$ ) is less  
219 than 75% after 45 minutes, the drug product is not inside the recommendation of the Ph. Eur. of an  
220 immediate release dosage form (see Annex 1: Decision tree for the principles for setting  
221 specifications). Therefore, the dissolution specification should be based on more than one time point.

222 In case there is no biobatch (e.g. BCS biowaiver), the specification limit with a fixed  $Q$  value within 15  
223 min (for BCS class I and III) or 30 minutes (applicable only for human BCS class I products) can be  
224 established. This  $Q$  value should be 80% using discriminatory test conditions, irrespective of the  
225 dissolution results of the test batch observed in the study used to claim the BCS biowaiver. The  
226 conditions for the dissolution test in the specification should be chosen as the most discriminatory  
227 between those used in the comparative dissolution study.

228 **Conclusion**

229 This reflection paper should facilitate congruent decisions on setting specifications for *in vitro*  
230 dissolution of generic drug products with immediate release characteristics. The principle is to derive  
231 the specification of the drug product on the basis of the quality characteristics of the biobatch. Similar  
232 principles may be considered for deriving the specification for innovator products.

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<sup>1</sup> Both these bullet points are captured in the first decision box in the decision tree in Annex 1.

234 **References**

- 235 European Pharmacopoeia (Ph. Eur.), 5.17.1, Recommendations on Dissolution Testing ;  
236 Guideline on the Investigation of Bioequivalence (CPMP/EWP/QWP/1401/98 Rev. 1/ Corr \*\*);  
237 Guideline on the conduct of bioequivalence studies for veterinary medicinal products  
238 (EMA/CVMP/016/00-Rev.2);  
239 European Pharmacopoeia (Ph. Eur.) 8<sup>th</sup> edition, 2.9.3, Dissolution Test for Solid Dosage Forms;  
240 Guideline on quality of oral modified release products (EMA/CHMP/QWP/428693/2013);  
241 ICH guideline Q8 (R2) on pharmaceutical development (EMA/CHMP/ICH/167068/2004);  
242 Note for Guidance Specifications: Test Procedures and Acceptance Criteria for new Drug Substances  
243 and new Drug Products – Chemical Substances (CPMP/ICH/367/96);  
244 VICH GL52 on Bioequivalence: blood level bioequivalence study (EMA/CVMP/VICH/751935/2013 –  
245 Corr.1).  
246

247 **Annex 1: Decision tree for the principles for setting**  
248 **specifications based on the dissolution results of the biobatch**

