



1 16 February 2017
2 EMA/CVMP/VICH/176637/2014
3 Committee for Medicinal Products for Veterinary Use (CVMP)

4 **VICH GL56 Studies to evaluate the metabolism and**
5 **residue kinetics of veterinary drugs in food-producing**
6 **species: study design recommendations for residue**
7 **studies in honey for establishing MRLs and withdrawal**
8 **periods**
9 Draft

Draft agreed by VICH Steering Committee	January 2017
Adopted by CVMP for release for consultation	16 February 2017
Start of public consultation	24 February 2017
End of consultation (deadline for comments)	31 July 2017

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Comments should be provided using this [template](#). The completed comments form should be sent to vet-guidelines@ema.europa.eu

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STUDIES TO EVALUATE THE METABOLISM AND RESIDUE KINETICS OF VETERINARY DRUGS IN FOOD-PRODUCING SPECIES: STUDY DESIGN RECOMMENDATIONS FOR RESIDUE STUDIES IN HONEY FOR ESTABLISHING MRLS AND WITHDRAWAL PERIODS

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Recommended for Consultation at Step 4 of the VICH Process
in January 2017
by the VICH Steering Committee

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This Guideline has been developed by the appropriate VICH Expert Working Group and will be subject to consultation by the parties, in accordance with the VICH Process. At Step 7 of the Process the final draft will be recommended for adoption to the regulatory bodies of the European Union, Japan and USA.

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58 **1. INTRODUCTION**

59 **1.1. Objective of guidance**

60 The objective of this guidance is to provide study design recommendations which will
61 facilitate the universal acceptance of the generated residue depletion data to fulfill the
62 national/regional requirements in order to establish appropriate Maximum Residue Limits
63 (MRLs) or other safe limits in honey following the treatment of honeybees with veterinary
64 drug products, or to justify withdrawal periods in honey for registration purposes when an
65 MRL already exists.

66 Use of veterinary drug products in honeybee production is considered as a minor use in minor
67 species in most jurisdictions.

68 **1.2. Background**

69 This guidance is one of a series developed to facilitate the mutual acceptance of residue
70 chemistry data for veterinary drug products used in food-producing animals. This guidance
71 was prepared after consideration of the current national/regional requirements and
72 recommendations for evaluating veterinary drug residues in the VICH regions.

73 **2. GUIDANCE**

74 **2.1. Purpose**

75 Residue studies in honey are recommended for registration or approval, as applicable, of a
76 veterinary drug product for use in honeybees.

77

78 These studies may be used to:

79 • measure the residues in honey

80 • generate data suitable for establishment of appropriate maximum residue limits.

81 • justify the withdrawal period for a veterinary drug product in accordance with an existing
82 MRL and/or generate data suitable for the establishment of risk-management measures
83 (*e.g.* use restrictions) in order to address consumer safety concerns. It is generally accepted
84 that the most practical withdrawal period in honey is a zero-day withdrawal, meaning that
85 all harvested honey should be safe for human consumption. (Honey is harvested when at
86 least 75% of the honeycells within a frame are filled and capped.) However, additional
87 measures (*e.g.* the time interval between end of treatment and start of honey flow) may
88 need to be followed.

89

90 Design elements for residue studies in honey differ in many respects from those in
91 commodities from other food producing species because honey is a unique food of animal
92 origin. There is minimal pharmacokinetic depletion of residues in honeybees following
93 treatment. When present in honey, residue concentrations are reduced mainly by dilution as
94 more honey is produced during the honey flow. Residue concentrations might also be
95 influenced by thermal degradation (as temperature inside the hive reaches 32-36 °C), acidic
96 hydrolysis (honey pH ranges 3.4 – 6.1) or other chemical reactions with honey matrix

97 components. Honey production rate depends on factors such as temperature, rain, season of
98 the year, climatic zone, food source/type and honeybee species/subspecies.

99 **2.2. Scope**

100 The intention is that one set of residue studies, conducted in multiple locations within one or
101 more regions, would satisfy the data recommendations for establishment of appropriate safe
102 limits for a veterinary drug (*i.e.*, the specific active substance)/veterinary drug product in
103 honey.

104 Studies should be conducted in conformity with the applicable principles of Good Laboratory
105 Practice (GLP).

106 **2.3. Residue Studies**

107 **2.3.1. General considerations**

108 When conducting residue studies for honey, treatments are typically applied to honeybee
109 colonies in accordance with Good Beekeeping Practice. Treatments are generally applied once
110 per year after honey harvesting and should be completed before honey flow commences.

111 **2.3.2. Test Article**

112 The test article (the veterinary drug product) used for the study should be representative of the
113 commercial formulation. Final Good Manufacturing Practices (GMP) manufactured material
114 (pilot scale or commercial scale) is the preferred source of test article; however, laboratory
115 scale preparations characterized with respect to GLP could also be appropriate.

116 **2.3.3. Residues to monitor**

117 Metabolic or total residue studies using radiolabelled drugs are not requested for MRL
118 assessment/approval of veterinary drug products used in honeybees. It is anticipated that in
119 most cases the residue to monitor would be the parent drug. However, data on
120 physicochemical properties of the active drug substance and other scientific information might
121 be useful to reveal the identity of putative transformation and/or degradation products. If data
122 indicate transformation or degradation of parent drug, an alternative residue or combination of
123 residues may need to be monitored. For substances that are prone to
124 transformation/degradation, prior to conducting residue studies, additional (*in-vitro*) studies
125 may be used to determine their stability in honey (during its production and up to its harvest).
126 Variables to be tested include pH, temperature, time and exposure to (UV)-light. The selected
127 conditions should be sufficiently justified.

128 **2.3.4. Bees and beekeeping conditions**

129 Healthy and strong colonies should be used (See Glossary for 'colony strength'). The
130 honeybee species/subspecies should be recorded. The colonies per site (See 2.3.6.) should be
131 uniform in adult honeybee population. The hives per site should be uniform in the number of
132 frames and box size and should be uniquely identified. Hive construction should be adequately
133 described. The hives should consist of one brood box only. A super box with frames should be
134 added at the start of honey flow. The number of frames in each box should be recorded.

135

136 Neither the colonies, boxes, nor frames should have a history of exposure to the veterinary
137 drug.

138 The study should be conducted at locations that mimic the conditions found at the time of the
139 year when apiarists would normally treat colonies with the particular veterinary drug product.

140 **2.3.5. Dosing and method of Administration**

141 The design should cover the maximum treatment regimen. The method of applying the
142 veterinary drug product in the study should be representative of the intended commercial
143 use. The route of administration should be described in detail.

144 If the veterinary drug product is intended to be applied in more than one method, a separate
145 residue study for each method of administration is recommended. Alternatively, a single study
146 representing the worst-case scenario can be conducted with the resulting safety
147 parameters (*i.e.*, proposed MRLs, use restrictions) being applicable to all methods. Full
148 justification of the worst-case scenario should be provided.

149 **2.3.6. Study design**

150 Residue studies should be conducted in four sites of differing agro-ecological areas within one
151 or more regions. If the residue studies are intended to support an application for a national
152 license, then depending on the country of application (size, variety of landforms and climatic
153 conditions), two to three sites (of differing agro-ecological areas) may be considered
154 sufficient. In such cases the national authorities could be consulted. For the duration of the
155 studies, information on climate (temperature, rainfall and any other parameter considered
156 relevant for the performance of the veterinary drug product) and beekeeping management
157 practices should be provided. In addition, data on the plants in the area in which honeybees
158 forage during the study as well as data on any supplemental feed given to the honeybees
159 should be reported.

160 The time of treatment, the approximate time of the start of honey flow and the time of sample
161 collection/colony should be provided.

162 **2.3.6.1. Colonies**

163 Six colonies per site should be treated, resulting in 24 colonies per residue study. Depending
164 on the type of application fewer treatment sites may be sufficient (See 2.3.6 referring to
165 application for a national license). A single sampling timepoint per colony is considered
166 appropriate. This is the first timepoint when honey from each colony is ready to be harvested
167 for human consumption (super honey from one or more frames). Honey harvest refers to the
168 collection of honey from the honeycombs once they are filled with capped honey; at least 75%
169 of the honeycells of the selected honeycombs should be filled and capped. Figure 1 outlines a
170 theoretical scheme (example) of sample collection per site.

171 **2.3.6.2. Residues in Comb Honey for Lipophilic Drug Substances Only**

172 Residue studies should be conducted as described in 2.3.6.1. In addition, for the last colony
173 harvested per site, a pooled wax sample (all available from the single colony) should be
174 collected and analysed following honey extraction (Figure 1). To determine residues in comb

175 honey from the separate concentrations in honey and wax, consider that 1 kg of comb honey
176 consists of 22/23 kg honey and 1/23 kg wax¹.

177 **2.3.6.3. Treatment during Honey Flow**

178
179 In cases of veterinary drug products that could be used during the honey flow, the basic study
180 design should be followed. The sponsor should provide justification for modifications to the
181 basic study design. Points to consider are transfer of the veterinary drug to existing and newly
182 produced honey.

183 **2.3.7. Sampling**

184 **2.3.7.1. Sample Preparation**

185 Super honey

186 The honey from all honeycombs in the collected frames from each colony should be harvested.
187 The honey should be extracted, filtered and thoroughly mixed to produce a pooled sample to
188 represent that particular colony. Extraction of honey from a honeycomb may be facilitated by
189 centrifugation. Sample processing (all activities after sampling and up to analysis) should take
190 into account the stability properties of the residues. The amount of the pooled honey produced
191 should be provided. The pH and moisture content of all pooled honey samples should be
192 measured and reported.

193 Wax

194 For wax samples, the combs should be homogenized after honey extraction. The amount of the
195 bulk wax sampled should be provided.

196 **2.3.7.2. Sample storage**

197 If the chemical analysis cannot be completed immediately following sample collection, the
198 samples should be stored appropriately. If samples are stored after collection, the Sponsor
199 bears the responsibility for demonstrating residue stability through to the time of assay. The
200 parameters and recommendations to assess sample storage are discussed in the VICH GL 49,
201 “Studies to Evaluate the Metabolism and Residue Kinetics of Veterinary Drugs in Food
202 Producing Animals: Validation of Analytical Methods Used in Residue Depletion Studies”².

203 **2.4. Analytical Method for Assay of Residues**

204 The Sponsor should submit a validated analytical method for the determination of the residues
205 in the samples generated. The method(s) should be capable of reliably determining
206 concentrations of the residues that encompass the appropriate proposed reference point for
207 honey (*i.e.*, MRL).

208 The parameters to be included in the method validation are fully discussed in the VICH GL
209 49, “Studies to Evaluate the Metabolism and Residue Kinetics of Veterinary Drugs in Food
210 Producing Animals: Validation of Analytical Methods Used in Residue Depletion Studies”
211 (See Footnote 2).

¹Beeswax Production, Harvesting, Processing and Products, Coggshall and Morse. Wicwas Press. 1984-06-01.
p. 41. ISBN 1878075063.

² VICH GL 49: Studies to Evaluate the Metabolism and Residue Kinetics of Veterinary Drugs in Food Producing Animals:
Validation of Analytical Methods Used in Residue Depletion Studies, EMA/CVMP/VICH/463202/2009.

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213 3. GLOSSARY

214 The following definitions are applied for purposes of this document.

215 **Beehive** is a place used for housing a colony of bees, commonly stackable wooden boxes
216 consisting of a bottom board, brood box/es and super box/es containing movable frames.

217 **Bee colony** is the aggregate of worker bees, drones, queen, and developing brood living
218 together as a family unit in a hive or other dwelling.

219 **Brood box (brood chamber)** is a box in which the queen is confined and brood is reared.
220 Brood refers to eggs, embryo's larval and pupal stages of bees.

221 **Colony strength** is evaluated by estimating the adult honey bee population in the hive and
222 depends on the time of the year and colony management practices.

223 **Comb honey** is honey stored by bees in the cells of freshly built broodless combs or thin comb
224 foundation sheets made solely of beeswax and sold in sealed whole combs or sections of such
225 combs (Revised Codex Standard for honey, 2001).

226 **Frame** is a rectangular wooden support designed to hold combs, usually 10 to each (brood or
227 super) box. In frames, a wax foundation is usually installed.

228 **Good Beekeeping Practice** refers to best practice recommendations found in numerous
229 references, for example: Hygiene in the apiary (A manual for hygienic beekeeping) Ed.
230 Dalibor Titera, BRI Dol 2009; National best management practice for beekeeping in the
231 Australian environment, The Australian honey bee industry council, 2007.

232 **Honey** is 'the natural sweet substance produced by honeybees from the nectar of plants or
233 from secretions of living parts of plants or excretions of plant sucking insects on the living
234 parts of plants, which the bees collect, transform by combining with specific substances of
235 their own, deposit, dehydrate, store and leave in the honey comb to ripen and mature'
236 according to the Codex definition (Revised Codex Standard for honey, 2001)³.

237 **Honey flow** is a period of time when one or more nectar or honeydew sources are in
238 abundance such that honeybees can store a surplus of honey.

239 **Honey harvest** refers to the collection of honey from the honeycombs once they are filled with
240 capped honey; at least 75% of the honeycells in a frame should be filled and capped.

241

242 **Honeybees** are a subset of bees in the genus *Apis*, primarily distinguished by the production
243 and storage of honey and the construction of perennial, colonial nests out of wax. Although
244 there are 7 species of honeybees (*A. andreniformis*, *A. cerana*, *A. dorsata*, *A. florea*, *A.*
245 *koschevnikovi*, *A. mellifera*, *A. nigrocincta*) only two of them *A. mellifera* (Western or
246 European honeybee) and *A. cerana* are maintained by beekeepers, with the former being the
247 most commonly domesticated species. *A. mellifera* is native to Europe, Asia and Africa and
248 was introduced to North America in the early 1600s. Since then it has spread worldwide.
249 There are many subspecies that have adapted to the local geographic and climatic
250 environments, and in addition hybrid strains have been bred [*e.g.* the Africanized bee (*A.*
251 *mellifera lingustica* X *A. m. scutellata*)].

³ Revised Codex Standard for honey (2001). [CODEX STAN 12-1981](#), Rev.1 (1987), Rev.2 (2001).

252 **Honeycomb** is a mass of hexagonal wax cells built by honeybees to contain their stores of
253 honey and pollen (honeycomb in the super box) or for raising brood (broodcomb in the brood
254 box).

255 **Lipophilic substance** refers to a chemical substance having high ($\log K_{ow} \geq 3$) octanol/water
256 partition coefficient.

257 **Maximum residue limit (MRL)** is the maximum concentration of a veterinary drug residue
258 that is legally permitted or recognized as acceptable in or on a food as set by a national or
259 regional regulatory authority. The term ‘tolerance,’ used in some countries, can be, in many
260 instances, synonymous with MRL.

261 **Residue** means the veterinary drug (parent) and/or its metabolites. In the case of honey this
262 may include transformation and degradation products.

263 **Super box** is a box in which the honeybees store honey (super honey) and that is placed above
264 a queen excluder and the brood chamber.

265 **Wax (or comb) foundation** is a plate made of wax with the base of the honeycomb on which
266 honeybees will construct a complete comb.

267 **Withdrawal period** is the period necessary between the last administration of a veterinary drug
268 product to animals and the production of foodstuffs from such animals, in order to protect
269 public health by ensuring that such foodstuffs do not contain residues in quantities in excess of
270 the maximum residue limits established.

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272 **Zero-day withdrawal** refers to a label indication that allows entry of edible tissues/animal
273 products into the food chain without regard to the time of last drug administration.

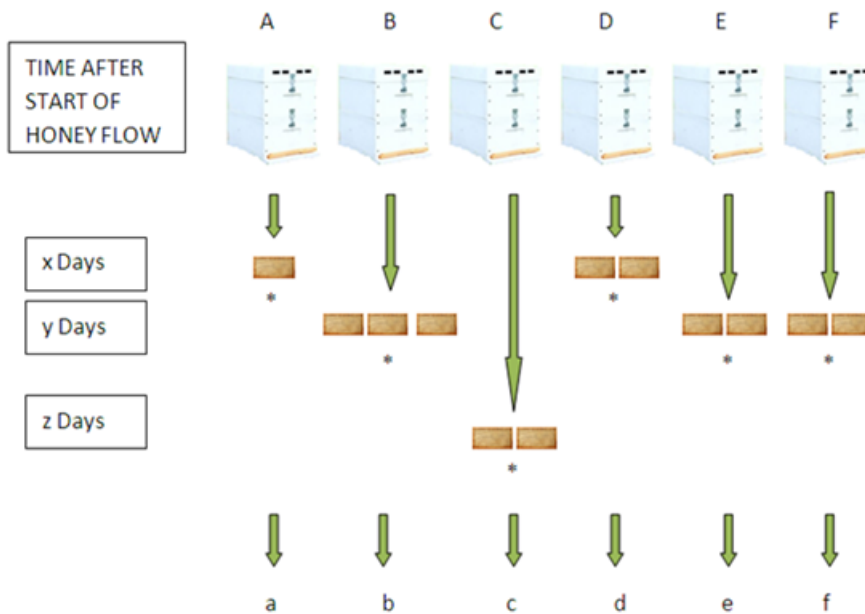
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x days < y days < z days

*: Number of frames collected per hive

a-f: Pooled honey samples from the collected frames of each hive

Under 2.3.6.2. (lipophilic compounds) a pooled wax sample should be collected following honey extraction of c honey sample, as in this figure C colony is the last colony harvested

279

280 **Figure 1.** A theoretical scheme of sample collection per site.

281 *The figure outlines a theoretical example of sample collection per site. A single sampling timepoint per colony is considered*
 282 *to be appropriate. This is the first timepoint when honey from each colony can be harvested for human consumption*
 283 *(only super honey from one or more frames). Honey harvest refers to the collection of honey from the honeycombs*
 284 *which are filled with capped honey. The figure illustrates that the time points when honey is mature (at least 75% of the*
 285 *honeycells in one or more frames are filled and capped) and the number of mature honeycombs per colony usually varies*
 286 *from hive to hive. The first honey harvest is considered the worst case in terms of residues.*

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