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4 Guideline on the adventitious agent safety of urine- 5 derived medicinal products

6 Draft

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8 This guideline replaces the document on Biological products derived from human urine (CPMP/118/95).
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11 **Guideline on the adventitious agent safety of urine-**
12 **derived medicinal products**

13 **Table of contents**

14 **1. Introduction (background)..... 3**
15 **2. Scope..... 3**
16 **3. Legal basis 3**
17 **4. Adventitious agents safety 3**
18 4.1. Selection of donors3
19 4.2. Processes4
20 4.3. Investigational studies of inactivation/reduction capacity of the manufacturing processes
214
22 4.4. Overall viral and TSE safety5
23 **References 6**
24

25 **1. Introduction (background)**

26 Human urine is used to prepare several products indicated in the field of endocrinology, such as human
27 chorionic gonadotropin (hCG), human menopausal gonadotropin or menotropin (HMG) and follicle-
28 stimulating hormone (FSH) and urokinase products used for thrombolysis. These hormones and
29 urokinase extracted from pooled human urine were available on the market as early as the 1970s.
30 Significant improvements in the manufacturing processes of these products have been introduced in
31 the 1990s in order to reach a higher purity profile. In parallel, marketing authorization dossiers have
32 been updated as regards the viral safety standards set during this decade.¹ Urine may be naturally
33 contaminated with viruses harboured in the urinary tract. Other viruses arising from the genital or
34 intestinal tract during urine collection may be present in urine donation. Assessment of the viral
35 clearance capacity of manufacturing processes has shown that the purification processes of these
36 medicinal products contain several steps able to remove/inactivate adventitious agents. These data
37 provide support that the viral safety record for this class of products is largely due to the extraction
38 and purification processes.

39 The emergence of variant Creutzfeldt-Jakob disease (vCJD) in the 1990s and more recently the cases
40 of apparent iatrogenic vCJD infection by blood transfusion in man in the UK prompted EMA to assess
41 the risk linked to the use of urine-derived products as regards this new form of CJD. Expert meetings
42 addressed this question in 2002 and 2007² and the results of these assessments were included in the
43 Position statement on Creutzfeldt-Jakob disease and plasma-derived and urine-derived medicinal
44 products and its revisions (February 2003, June 2004 and June 2011).³

45 **2. Scope**

46 Medicinal products derived from human urine fall under the definition of Article 1(2b) of Directive
47 2001/83/EC⁴ as follows: "Any substance or combination of substances which may be used in or
48 administered to human beings either with a view to restoring, correcting or modifying physiological
49 functions by exerting a pharmacological, immunological or metabolic action, or to making a medical
50 diagnosis."

51 This document addresses specific aspects which should be taken into consideration in the evaluation of
52 viral and TSE safety of medicinal products derived from human urine.

53 **3. Legal basis**

54 This guideline should be read in conjunction with the introduction and general principles (4) and Annex
55 I to Directive 2001/83/EC as amended.⁴

56 **4. Adventitious agents safety**

57 **4.1. Selection of donors**

58 Generally, donations of urine are from volunteers, who are instructed to refrain from donating in case
59 of illness. In addition, for enrolment in a donor panel, manufacturers should establish exclusion criteria
60 with respect to the general status of health as far as this is feasible. Accordingly, for urine-derived
61 hormones, which are sourced from a relatively small well-defined donor population, manufacturers
62 have put in place limited exclusion criteria for the selection of a donor. However, for other products
63 manufactured from very large donor pools (e.g. urokinase), such measures are difficult to apply.

64 As urine collection takes place outside of professional supervision these criteria would not be checked
65 at each donation unlike blood/plasma donors. Therefore manufacturers should follow up the donor
66 criteria at defined intervals.

67 **4.2. Processes**

68 Two classes of drug substance are derived from human urine - hormones (hCG, hMG, FSH) and
69 urokinase. Manufacturing strategies vary according to product and manufacturer. They generally
70 consist of extraction, precipitation and purification steps, which are applied after individual urine
71 collections, with or without preservative, have been pooled.

72 Urine may be contaminated with viruses harboured in the urinary tract or with viruses originating from
73 the genital or intestinal tract. Taking into consideration limitations associated with testing of large
74 urine pools used as starting material, virus safety mainly relies on the potential of the production
75 process to inactivate or remove viruses. Manufacturers are therefore required to investigate the
76 capacity of their manufacturing processes to inactivate/remove a broad range of viruses representing
77 various physico-chemical properties. The available data suggest efficient clearance of viruses, which
78 may contaminate the urine pool, by defined steps in the manufacturing process. More specifically, for
79 urokinase dedicated viral clearance steps often consist of a pasteurisation step and nanofiltration. As
80 regards the urine-derived hormones, virus clearance is attributed to a combination of process steps,
81 which are specific for the individual manufacturing processes, such as alkali treatment, precipitation or
82 chromatographic steps. Manufacturers of urinary-derived hormones have been encouraged to
83 incorporate nanofiltration to further improve clearance of highly resistant, small non-enveloped viruses
84 and several manufacturing processes include such a virus filtration step.

85 Due to the number of places where starting materials are sourced, particular attention should be given
86 by manufacturers to the overall Quality Assurance System in place for the whole collection system and
87 to the validation/control of the early production steps of the manufacturing process.⁷

88 **4.3. Investigational studies of inactivation/reduction capacity of the** 89 **manufacturing processes**

90 General guidance on choice of viruses is given in the *Note for Guidance on virus validation studies: The*
91 *Design, Contribution and Interpretation of Studies Validating the Inactivation and Removal of Viruses*
92 (CPMP/BWP/268/95, revised).¹ This section contains further guidance relevant to urine derivatives. The
93 viruses that are the more frequently found in human urine are hepatitis B virus (HBV), human
94 cytomegalovirus (HCMV), and those from papillomavirus and polyomavirus genus. The presence of
95 other viruses brought by faecal contamination cannot be excluded (e.g. hepatitis A virus (HAV) or
96 other enteroviruses, hepatitis E virus (HEV), adenoviruses, noroviruses, astroviruses, coronavirus-like
97 particles, rotaviruses).

98 Viruses to be used in validation studies on urine-derived medicinal products should include:

99 Enveloped viruses

- 100 • enveloped RNA viruses (e.g. bovine viral diarrhoea virus (BVDV))

101 Enveloped RNA viruses such as rubella virus, mumps virus or measles virus are shed into human urine
102 during acute infection. RNA from numerous additional enveloped viruses has been detected in human
103 urine. Even if the presence of infective enveloped virus particles in urine is unlikely in many cases, it
104 cannot be totally excluded. Various enveloped RNA-models have been used to validate virus
105 inactivation methods. However, to date, the pestivirus bovine viral diarrhoea virus (BVDV) is
106 considered as a worst-case model for other RNA enveloped viruses.

107 • enveloped DNA viruses (e.g. herpesvirus, pseudorabies virus (PRV))

108 Human cytomegalovirus (HCMV) can be transmitted via urine. It is recommended to include a
109 herpesvirus such as pseudorabies virus (PRV) in the panel to model DNA enveloped viruses. For the
110 validation of steps based on size exclusion (virus filtration) studies with herpesviruses are not
111 necessary. Currently, there is no practical test system for hepatitis B virus titration. The duck hepatitis
112 B virus (DHBV) may be used as a model of human HBV. However, it requires the use of its natural
113 animal host (duck or primary duck cells) for titration. In consequence, there is no general requirement
114 to include DHBV in the virus panel.

115 Non-enveloped viruses SV40 and animal parvovirus

116 Infectious polyomaviruses, adenoviruses and enteroviruses can be found in human urine. SV40 as a
117 member of the polyomaviridae virus family should be used in validation studies. SV40 is also relevant
118 to represent HBV in size exclusion steps. Viruses which can be excreted at high titers in human stool
119 include many non-enveloped DNA or RNA viruses such as adenoviruses, hepatitis A virus (HAV) and
120 other enteroviruses, hepatitis E virus, noroviruses and astroviruses. An appropriate model for highly
121 resistant small non-enveloped viruses should be incorporated in the panel. This may be chosen among
122 porcine, canine, bovine and murine parvoviruses. In some specific cases, it may be justified to include
123 HAV in the panel to model enteroviruses (for example when one step is not expected to be efficient on
124 a more resistant virus like porcine parvovirus).

125 **4.4. Overall viral and TSE safety**

126 Urine-derived medicinal products have been used in the treatment of a number of conditions for
127 several years without any suspicion that they are responsible for the transmission of any infectious
128 agents. It is nevertheless fundamental to perform risk assessments for the overall transmission risks
129 for urine-derived medicinal products.

130 The following are likely to be the main components of each overall risk analysis. Estimates of the
131 robustness of the analysis in each case might usefully accompany each component.

- 132
- 133 1. Viral epidemiology for the region where collection takes place, and for the specific donor
134 population there (i.e. on the basis of age, gender, and endocrinal status).
 - 135 2. On the basis of the epidemiology data and taking into consideration the capacity for human
136 kidneys, urinary and genital tracts to harbour pathogens, agents which are most likely to be
137 relevant for the product could be identified.
 - 138 3. Donor selection criteria, encompassing donor briefing strategies with an estimate of how
139 effective they might be in particular populations, and donor motivation factors.
 - 140 4. The donation and collection system up to the start of pooling, and including the security and
141 hygienic measures in place.
 - 142 5. Any information available on the Quality Assurance System, Audits and Procedures followed by
143 the manufacturers to control the collection system and early production steps of the different
144 manufacturers/suppliers.
 - 145 6. Pooling strategies with a consideration of screening tests performed.
 - 146 7. The extraction and purification methodologies, including a consideration of any further
147 pathogen screening tests applied, and the indication of the point at which GMP starts.
 - 148 8. The effectiveness of each TSE agent and virus elimination step applied and the relevance of the
149 results obtained with model viruses used in validation studies with regards to the virus that
150 may be found in the starting material.

151 Where practicable, consideration should be given by companies to presenting estimates of the
152 probabilities of individual doses of a urine-derived medicinal product being contaminated with a
153 pathogen. Such risk analyses should follow the methodologies developed for plasma derived medicinal

154 products and should take into account viral safety aspects described in the plasma derived medicinal
155 products guideline⁵ and the guidance concerning reduction of TSE agents discussed in the “CHMP
156 Position statement on Creutzfeldt-Jakob Disease and Plasma-derived and Urine-derived Medicinal
157 products”³ and in the guideline on “Investigation of Manufacturing Processes for Plasma-derived
158 Medicinal products with regard to vCJD risk”.⁶ Risk analyses of this nature should appear in 3.2.A.2
159 Adventitious Agent Safety Evaluation of Marketing Authorisation applications.

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