



1 18 October 2013
2 EMEA/CVMP/EWP/141272/2011
3 Committee for Medicinal products for Veterinary Use (CVMP)

4 **Guideline on the conduct of efficacy studies for**
5 **intramammary products for use in cattle**

6 Draft

Draft agreed by Efficacy Working Party (EWP-V)	September 2013
Adopted by CVMP for release for consultation	10 October 2013
Start of public consultation	18 October 2013
End of consultation (deadline for comments)	30 April 2014

7 *This guideline replaces* the CVMP guideline "Conduct of efficacy studies for intramammary products for
8 use in cattle "[\(CVMP/344/1999 Rev.1\)](#)

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10 **intramammary products for use in cattle**

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40 **Executive summary**

41 This revised guideline is intended to provide guidance on the conduct of efficacy studies and their
42 evaluation for veterinary medicinal products that are administered via the teat canal to cattle. It
43 therefore addresses the treatment of clinical and subclinical mastitis during the lactation period, the
44 treatment of subclinical mastitis at drying off, and the prevention of new intramammary infections
45 during the dry period. The scope of the guideline has been extended in order to include
46 recommendations on pre-clinical data, in addition to those on clinical field studies for the
47 demonstration of efficacy. Moreover, information is included for generic intramammary products.

48 **1. Introduction (background)**

49 This guideline addresses data requirements for demonstrating pre-clinical and clinical efficacy of
50 products for intramammary use in cattle.

51 The majority of products for treatment and prevention of intramammary infections contain
52 antimicrobial substances, and the recommendations in this guideline focus on such products and their
53 use. It is recognised that acceptable methods other than those referred to in this guideline might be
54 capable of providing adequate information, provided they are sufficiently justified.

55 Since the principles for demonstrating clinical efficacy of a product intended for treatment and/or
56 prevention of intramammary infections are the same for antimicrobials as for other types of
57 substances, recommendations made in this guideline also apply to intramammary products containing
58 other types of active substances.

59 SPC recommendations made for the use of intramammary products should be evidence-based,
60 meaning a rationale with respect to active substance, dose, frequency of administration and treatment
61 length should be given, and the anticipated efficacy of the product should be demonstrated and
62 confirmed by appropriate pre-clinical and clinical studies.

63 **2. Scope**

64 This guideline is intended to provide guidance on design, conduct and reporting of pre-clinical and
65 clinical studies submitted in support of a new application for a marketing authorisation for a product for
66 intramammary use in dairy cattle, or to vary the conditions for use of an already authorised product.

67 Recommendations concern intramammary products for use during lactation and at drying off. This
68 guideline also includes recommendations for generics of authorised intramammary products.

69 For intramammary products containing antimicrobial substances, recommendations made in the
70 guideline for the Demonstration of Efficacy for Veterinary Medicinal Products containing Antimicrobial
71 Substances (EMA/CVMP/627/2001) and in the guideline for the conduct of pharmacokinetic studies in
72 target animal species (EMA/CVMP/133/1999-final) apply, where relevant. With regard to tolerance
73 please see VICH GL 43 guideline on target animal safety for veterinary pharmaceutical products
74 (CVMP/VICH/393388/2006) and the guideline (on) local tolerance of intramammary preparations in
75 cows (7AE21a, Volume 7, 1993).

76 **3. Legal basis**

77 This guideline replaces the current CVMP guideline for the conduct of efficacy studies for
78 intramammary products for use in cattle (CVMP/344/99- final-rev.1) and should be read in conjunction
79 with Directive 2001/82, as amended.

80 Applicants should also refer to other relevant European and VICH guidelines, including those listed in
81 the reference list of this document.

82 **4. Pharmacology**

83 **4.1. Pharmacodynamic properties**

84 Studies on pharmacodynamics should be performed according to validated and/or internationally
85 accepted methods, if available.

86 As a general rule, the mode and mechanism of action underlying the desired therapeutic effect(s) of
87 the active substance(s) should be described, and any possible secondary and adverse effects relevant
88 for the target species/indication should be reported. Furthermore, the influence of milk on the
89 pharmacological activity of the active substance(s) should be investigated, where appropriate.

90 Studies may include *in vitro* and/or *in vivo* designs. The experimental design employed and the method
91 of measuring the pharmacodynamic effect should be fully described by the applicant, unless they are
92 known as standard procedures.

93 **4.2. Pharmacokinetics**

94 For lactating cow products, the concentration of the active substance(s) in plasma as a function of time
95 should be determined to investigate the potential systemic absorption. Furthermore, the concentration
96 of the active substance(s) in milk as a function of time should be investigated to allow an estimation of
97 the therapeutic concentration-time profile at the infection sites in the udder.

98 For dry cow products, the concentration profile in plasma should be investigated in order to determine
99 the extent of systemic absorption.

100 In addition, factors like release of the active substance(s) from the formulation, and the physico-
101 chemical properties of the active substance(s) and the excipients should be considered, as these may
102 have influence on the availability of the product in the milk or dry udder secretion, as well as in udder
103 tissue. In this respect parameters like composition, particle size distribution, viscosity and dissolution
104 in milk should be discussed with regard to the claimed indication.

105 **5. Dose determination**

106 The aim of dose determination studies is selection of an optimal dose and dosing regimen, taking the
107 target pathogen species into account and minimising the risk for development of resistance.

108 When selecting the appropriate dose the following aspects should be considered:

- 109 • quantity/activity of the active substance(s) and volume of the product, administered to a single
110 quarter,
- 111 • number of administrations per day (dosing interval),
- 112 • number of administrations needed to achieve complete cure (duration of treatment).

113 The rationale for the dosing regimen should be provided.

114 Dose determination studies should be performed in line with the principles of good clinical practice
115 (GCP, see VICH GL 9 - Good clinical practice, CVMP/VICH/595/98-final). Where possible, the final
116 formulation of the test product should be used.

117 For defining the target dose usually three dose levels need to be tested. The inclusion of a negative
118 control group is mandatory, which consequently requires the implementation of an adequate rescue
119 protocol for animal welfare reasons. With regard to clinical and subclinical mastitis during lactation,
120 investigation of different treatment durations is recommended in order to identify an optimal dosing
121 strategy. Dosing intervals should be aligned with usual milking intervals.

122 Dose determination studies could be performed either in naturally or experimentally infected cows. In
123 the absence of experimental models for dry cow therapy dose determination should be conducted
124 under field conditions.

125 **5.1. Experimental studies in lactating cows**

126 In lactating cows, dose determination should be studied under controlled conditions in experimentally
127 infected animals using suitable and, preferably, well documented models. In this respect the “3R-
128 principles” (replacement, reduction, refinement) should be considered.

129 The experimental infection should be performed with an udder pathogen which is relevant for the
130 claimed indication, and which can induce a disease pattern of clinical and/or subclinical mastitis similar
131 to natural infection. Information with regard to origin and *in vitro* susceptibility of the challenge strain
132 of the target pathogen to the proposed substance(s) should be provided. The choice of the challenge
133 strain should be justified. The design of an experimental study (e.g. time point for initiation of
134 treatment, sampling procedure, observation period, efficacy criteria etc.) should mimic field conditions.
135 If an experimental infection study is not feasible, dose determination may also be performed in
136 naturally infected animals.

137 **6. Dose confirmation**

138 Confirmation of the selected dosing regimen should be performed with the final formulation, preferably
139 in naturally infected animals. The evaluation can be performed under field conditions or under well-
140 controlled clinical conditions (e.g. laboratory conditions). It may also be appropriate to use dose
141 confirmation studies to investigate different treatment durations if this cannot be explored in dose
142 determination studies.

143 Preferably the study should include a negative control group (see also 7.1); this may require
144 appropriate measures with regard to animal welfare. Where study conditions do not allow inclusion of a
145 negative control group (e.g. in clinical mastitis cases with low spontaneous cure rates) it may be
146 acceptable to use a suitable positive control. In such a case the internal validity of the study needs to
147 be ensured.

148 Dose confirmation studies may be waived in circumstances where dose-finding data are available that
149 provide convincing support that the selected dosing regimen is appropriate for the treatment of
150 naturally occurring infections.

151 **7. Field studies**

152 **7.1. General considerations**

153 Field studies should be carried out to confirm the efficacy (and safety) of the test product at the
154 selected dosage regimen under practical conditions. The final formulation of the test product should be
155 used.

156 The studies should be multicentric and representative for European conditions, taking into account
157 differences in animal husbandry systems, geographical location and climate, and they should be
158 performed in line with GCP. Appropriate statistical methods should be applied (see CVMP guideline on
159 statistical principles for veterinary clinical trials, CVMP/EWP/81976/2010).

160 **7.2. Study design and population**

161 Field studies should be blinded (whenever feasible), controlled and animals should be allocated
162 randomly to test and control groups. The details of the blinding method used should be provided.

163 The number of cows selected from a single herd should not exceed 20% of the total number of cases
164 included in the complete study (to avoid that treatment outcome evaluation is dominated by the
165 results in one single herd).

166 The study should be designed so as to ensure that blinding is not jeopardised in circumstances where
167 the withdrawal periods differ between test and control treatment. The **positive control** should be an
168 intramammary product with the same indications as the test product and should be authorised in
169 accordance with Council Directive 2001/82/EC as amended. The applicant should justify the choice of
170 the positive control in relation to the indication and the target population for treatment. Products for
171 which recent susceptibility data suggest that posology may be inadequate for the infection under
172 study, or products where posology differs between Member States should be avoided. In the absence
173 of a suitable positive control the applicant should seek scientific advice from the authorities.

174 A **negative control** is considered mandatory for demonstration of efficacy for preventive treatments
175 at drying off, implying that an untreated group of cows with non-infected animals/quarters needs to be
176 included. Comparison with a negative control is also considered necessary for infections with a high
177 spontaneous cure rate (e.g. subclinical infections, *E. coli* infections), since a non-inferiority study
178 design is unlikely to yield conclusive information for this situation. Appropriate measures with regard to
179 animal welfare should be taken into account.

180 The choice of the type of control should be justified by the applicant.

181 **7.3. Pathogens**

182 A claim for efficacy should be demonstrated for each target pathogen separately. The choice of the
183 claimed pathogens should be justified with regard to the intended use of the product (either during
184 lactation or at drying off), and with regard to the spectrum of activity of the substance under study. In
185 general, the clinical study should be sufficiently powered to demonstrate a statistically significant effect
186 for each claimed bacteria species separately. For pathogens less common in the field, it may be
187 difficult to recruit sufficient cases. In such a situation, a lower number of cases may be justifiable
188 provided the overall data base can support conclusions on efficacy.

189 **7.4. Bacteriological diagnostic procedures**

190 Milk sampling and microbiological investigations should be carried out in accordance with standard (or
191 accepted) methods, for example, those recommended by the National Mastitis Council or by other
192 adequate references.

193 For recruitment, bacteriological examinations of milk samples should be performed from all udder
194 quarters of any cow in order to meet the inclusion criteria. After treatment, bacteriological
195 examinations of milk samples should be performed from all included quarters.

196 For mastitis pathogens isolated from pre-treatment milk samples, *in vitro* antimicrobial susceptibility to
197 the antimicrobial(s) used should be determined according to recognized procedures. For animals which
198 are classed as clinical failures, susceptibility testing should be performed as well.

199 **7.5. Relevant parameters for efficacy evaluation**

200 **Bacteriological status**

201 Bacteriological status is the primary parameter for evaluating success of treatment and should be
202 evaluated for each included udder quarter. Only cases of clinical and subclinical mastitis in which
203 relevant pathogens are isolated in the pre-treatment sample should be used in calculating cure rates.

204 **Clinical status**

205 In clinical mastitis cases the clinical cure is the co-primary parameter. The clinical cure should be
206 evaluated for each infected quarter and based on the return to normal of the parameters concerning
207 the cow's general condition, the appearance of the milk and the local clinical signs of the udder.

208 **Somatic cell counts (SCC)**

209 In clinical and subclinical mastitis trials, individual quarter milk SCC is determined from one pre-
210 treatment sample and from the second post-treatment sample. The same applies to cases in which
211 prevention of new infections at drying off is studied. Mean SCCs are calculated from the results for
212 each treatment group and – in case of clinical and subclinical mastitis - separately for bacteriologically
213 cured and not cured quarters. The mean SCC results for each treatment group may be used as a
214 secondary endpoint.

215 **7.6. Herd and cow information**

216 Study cows should be selected from herds with proper cow identification and health records. The
217 history of the herd and cows should be recorded after the inclusion of a cow in the trial and before the
218 commencement of the treatment.

219 Farm:

- 220 • Name and address of herd owner;
- 221 • Location of the herd;
- 222 • Number of dairy cows;
- 223 • Methods of herd management, milking, and dry cow management;
- 224 • Teat disinfection procedures if practised;
- 225 • Bulk milk SCC in the herd over preceding months.

226

- 227 Cows:
- 228 • Name or identification number;
 - 229 • Breed;
 - 230 • Number of lactations;
 - 231 • Date of calving;
 - 232 • Estimated or measured milk yield at time of treatment;
 - 233 • Cow milk SCC during preceding months;
 - 234 • History of previous mastitis treatments;
 - 235 • In clinical mastitis: carefully recorded clinical signs at the time of treatment;
 - 236 • In dry cow treatment: the milk yields of cows at drying off and the method of drying off.

237 **7.7. Inclusion criteria**

238 With regard to inclusion criteria, please refer to the chapters which address special considerations for
239 the respective indications.

240 **7.8. Exclusion criteria**

241 The following cows are to be excluded from the trial:

- 242 • Cows with concurrent disease;
- 243 • Cows given systemic or intramammary anti-infectious or anti-inflammatory treatments within a
244 30-day period before the trial;
- 245 • Cows vaccinated with products inducing an immune-mediated response against mastitis
246 pathogens.
- 247 • Cows with visible teat damage;
- 248 • In clinical mastitis: cows with severe systemic clinical signs;
- 249 • In clinical mastitis: cows with mastitis in two or more udder quarters;
- 250 • In clinical and subclinical mastitis: cows with a daily milk yield less than 5 litres of milk prior to
251 onset of clinical signs.

252 **7.9. Special considerations for clinical mastitis in lactating cows**

253 **Treatment unit**

254 In clinical mastitis the treatment and the statistical unit is the individual udder quarter.

255 **Inclusion criteria**

256 In clinical mastitis trials, all lactating cows with clinical mastitis limited to 1 quarter which can be
257 treated with intramammary treatment only are eligible. The pre-treatment milk sample should be
258 bacteriologically positive regarding the target pathogen(s) as claimed.

259 **Pre-treatment sampling**

260 Before treatment one milk sample should be taken for bacteriological analysis and determination of
261 quarter milk SCC and the cow should be clinically examined (general condition, appearance of milk,
262 udder consistency).

263 **Treatment**

264 In any included cow only the single affected quarter will be treated. A cow developing clinical mastitis
265 in additional quarters during the experimental period should be withdrawn from the study post
266 inclusion (please, refer to section 7.12). With regard to controls please refer to section 7.2. In addition
267 clinical examination should be made when considered necessary

268 **Post-treatment sampling**

269 After treatment two milk samples should be taken for bacteriological analysis. These samples should
270 be taken between day 14 and day 28 from the last treatment, at least 7 days apart. Clinical
271 examination should be performed at the first bacteriological post treatment sampling. If clinical cure
272 has not been achieved by this sampling time point, the case should be excluded from further sampling
273 (see below for assessment of success/failure). Quarter milk SCC should be determined from the second
274 post-treatment sample.

275 **Assessment of success/failure**

276 Cases of success and failure which are to be included in the final data analysis:

277 A case is regarded a treatment success if there is clinical cure at the first post-treatment sampling
278 (normal appearance of the milk, normal condition of the udder, normal general condition) as well as
279 bacteriological cure in both post-treatment milk samples (absence of the udder pathogen species which
280 was present at the time of inclusion).

281 Quarters with new infections in the originally infected, treated quarter (i.e. detection of an udder
282 pathogen which is different from that isolated at inclusion) in one or both post-treatment milk samples
283 can be classified as a bacteriological cure for the original pathogen. The number and type of new
284 infections in each treatment group should be included in the final study report and needs further
285 clarification. A high frequency of these occurrences is not acceptable.

286 A case is regarded a failure

- 287 • If the criteria for clinical cure are not met in the clinical examination at the first post-treatment
288 sampling (the cow should then have been excluded from further sampling).
- 289 • If the original pathogen detected at the time of inclusion is present in either or both post-treatment
290 samples.
- 291 • If additional antimicrobial treatment associated with the mastitis is necessary during the
292 experimental period.

293 **7.10. Special considerations for subclinical mastitis in lactating cows**

294 **Treatment unit**

295 In subclinical mastitis during lactation, the treatment unit is the cow, but the statistical unit is the
296 individual quarter.

297 **Inclusion criteria**

298 In subclinical mastitis trials, all lactating cows with the presence of the same target pathogen(s) in two
299 pre-treatment milk samples in conjunction with elevated quarter somatic cell count (SCC) > 200 000
300 cells/ml in one pre-treatment milk sample are eligible for a study. More than one quarter may qualify
301 for inclusion.

302 **Pre-treatment sampling**

303 Before treatment two quarter milk samples should be taken one to three days apart for bacteriological
304 analysis; if a pathogen can only be isolated from one out of these two samples, diagnosis should be
305 confirmed with a third sample. Quarter milk SCC should be determined from one of the pre-treatment
306 samples.

307 **Treatment**

308 Only confirmed positive quarters are eligible for treatment. With regard to controls, please refer to
309 section 7.2.

310 **Post-treatment sampling**

311 After treatment two milk samples should be taken for bacteriological analysis. These samples should
312 be taken between day 14 and day 28 from the cessation of the treatment from each included quarter,
313 and should be separated by a period of at least 7 days. Quarter milk SCC should be determined from
314 the second post-treatment sample.

315 **Assessment of success/failure**

316 Cases of success and failure which are to be included in the final data analysis:

317 A case is regarded a treatment success if the original pathogen is not detected in either of the post-
318 treatment milk samples, supported by a decrease in the somatic cell count.

319 With regard to new infections the same evaluation as defined for clinical mastitis cases will apply.

320 A case is regarded a failure

321 • If the original pathogen detected at the time of inclusion is present in either or both post-treatment
322 samples.

323 • If additional antimicrobial treatment associated with the subclinical mastitis is necessary during the
324 experimental period.

325 ***7.11. Special considerations for subclinical mastitis at drying off and***
326 ***prevention of new infections during the dry period***

327 Both treatment of subclinical infections at drying off and prevention of new infections during the dry
328 period can be studied in the same animal; however, treatment and prevention should not be studied in
329 the same quarter.

330 **Treatment unit**

331 For dry cow treatment, the treatment unit is the cow but the statistical unit is the individual quarter.

332 **Inclusion criteria**

333 For dry cow treatment, lactating cows which are approaching the end of lactation and ready for drying-
334 off are eligible for the trial.

335 Cows with subclinically infected quarters (presence of the same target pathogen(s) in two pre-
336 treatment milk samples, SCC > 200 000 cells/ml in one of these samples) are eligible for studying
337 treatment effect on subclinical infections.

338 For assessment of prevention of new infections during the dry period, only non-infected healthy
339 quarters are eligible at drying-off. Two pre-treatment milk samples should be bacteriologically negative
340 and SCC values, examined in one of these samples, should be < 200 000 cells/ml.

341 Only animals with dry periods of sufficient length (approximately 35 days or more) should be included.

342 **Pre-treatment sampling**

343 Within one week prior to drying-off, two pre-treatment quarter milk samples should be taken one to
344 three days apart from all quarters for bacteriological analysis. For subclinical mastitis cases in which a
345 pathogen can only be isolated from one out of two milk samples, a third sample may be necessary for
346 confirmation of diagnosis (see also inclusion criteria).

347 The same sampling strategy applies with respect to prevention of new infections during the dry period.
348 In cases where only one out of two pre-treatment milk samples is free of pathogens, a third sample is
349 needed to confirm the diagnosis. Quarter milk SCC should be determined from one of the pre-
350 treatment samples.

351 **Treatment**

352 At drying-off, all four quarters of animals should be treated. This may include treatment of infected
353 and non-infected quarters of one cow. With regard to controls please refer to section 7.2.

354 **Post-treatment sampling**

355 After calving two post-treatment milk samples should be taken for bacteriological analysis. The first
356 milk sample should be taken before the first regular milking after calving, and the second post-
357 treatment sample 4 to 7 days later. Quarter milk SCC should be determined from the second post-
358 treatment sample.

359 In addition the cow should be clinically examined after calving at appropriate times and intervals, for
360 any pathological changes of the udder or of the appearance of the milk.

361 **Assessment of success/failure**

362 Cases of success and failure which are to be included in the final data analysis:

363 Subclinical mastitis

364 A case is regarded a treatment success if the original pathogen is not detected in either of the two
365 post-treatment milk samples.

366 With regard to new infections the same evaluation as defined for clinical mastitis cases will apply.

367 A case is regarded a failure

- 368 • If the original pathogen detected at the time of inclusion is present in one or both post-treatment
369 samples.

- 370 • If additional antimicrobial treatment associated with the subclinical mastitis is necessary during the
371 experimental period.

372 Prevention of new infections

373 A case is regarded a prevention success if no udder pathogens can be detected in either of the post-
374 treatment milk samples after calving.

375 A case is regarded a prevention failure

- 376 • If any target udder pathogens can be detected in either or both post-treatment milk
377 samples(corresponding to a new infection).
- 378 • If additional antimicrobial treatment is necessary during the experimental period.

379 **7.12. Withdrawals**

380 Animals/quarters which are to be excluded from the final data analysis should be recorded as follows:

- 381 • Cases which are not interpretable due to lack or loss of information (e.g. quarters with no
382 pathogens in the pre-treatment samples, contaminated pre-treatment milk samples) shall be listed
383 in the final report, and their distribution in each group shall be analysed.
- 384 • Data from cows with clinical mastitis in which additional quarters had to be treated during the
385 experimental period shall be excluded from the final analysis and listed separately for each
386 treatment group. The reasons and the potential impact of the withdrawals on the study results
387 should be discussed.
- 388 • Cows treated with antibiotics due to intercurrent diseases during the experimental period should
389 be excluded from the trial and indicated in the final report.
- 390 • Any other cases in which the exclusion from final data analysis is justified should be indicated as
391 well.

392 **7.13. Presentation of data - reporting**

393 The data to be presented are described in the annex to Directive 2001/82/EC, as amended. This
394 includes that a record from each individual case should be presented in the final report. The data on
395 the bacteriological results and the bacteriological response for each organism for each treated quarter
396 should be summarized and tabulated separately for each bacterial species and treatment group. *In*
397 *vitro* susceptibility results should be enclosed in the final report.

398 The data should be expressed as number of quarters and number of cows cured clinically,
399 bacteriologically, and based on individual quarter milk SCC (subclinical mastitis only), see table 1.

400 For subclinical mastitis studies, it is preferable to present combined cure rates based on individual
401 quarter data (bacteriological cure + quarter milk SCC < 200 000 cells/ml).

402 *Table 1.* An example for data presentation for each treatment group in clinical mastitis (further details
403 are given in the text).

		Post-treatment cure in clinical mastitis					
		Clinical cure		Bacteriological cure		Bacteriological + clinical cure	
Treatment groups	No of quarters/cows	n	%	n	%	n	%

Test product				
Positive control				
and/or				
Negative control				

404 Table 2. An example for data presentation for each treatment group in sub clinical mastitis (further
405 details are given in the text).

		Post-treatment cure in subclinical mastitis			
		Bacteriological cure		Bacteriological + SCC	
Treatment groups	No of quarters/cows	n	%	n	%
Test product					
Positive control					
and/or					
Negative control					

406 Cases of clinical mastitis occurring during the dry period and during the post-calving investigational
407 period should be recorded.

408 8. Generic products – data requirements

409 The overarching principle is that generics of intramammary products should be therapeutically
410 equivalent to an originator, the reference product being a product with a complete documentation for
411 marketing authorisation. However, the Guideline “Conduct of bioequivalence studies for veterinary
412 medicinal products (CVMP/016/2000/Rev. 2)” is not applicable for locally acting products such as
413 intramammary products. In consequence, Art. 13 (3) of the Directive 2001/82/EC as amended applies,
414 i.e. data demonstrating the efficacy should be provided. In such cases comparable efficacy between
415 test and reference product should be demonstrated by an appropriate clinical trial, e.g. by a non-
416 inferiority field study. Differences in product formulation may influence penetration and distribution of
417 the active substance in the mastitic udder. Taking into account the different locations of mastitis
418 pathogens, it may therefore not be possible to predict that efficacy of a generic product will be non-
419 inferior for all target pathogens based only on efficacy for the pathogen that is the least susceptible *in*
420 *vitro*. Therefore, in order to gain all the claims for the reference product, the study should be
421 conducted using the target pathogen that is justified as the most difficult to treat *in vivo*. The
422 parameters for evaluation of efficacy in field trials apply. It is recognised that large numbers of cases
423 will be required to satisfy statistical requirements. If adequate safety parameters are also recorded in
424 the clinical trial, it may be possible to waive a dedicated local tolerance study.

425 However, specific efficacy (and safety) studies may be waived if the formulation of the generic product
426 is demonstrated to be qualitatively and quantitatively identical to the reference product, i.e. identical
427 pharmaceutical form, active substance(s) and excipient(s) and physicochemical properties (e.g.,
428 dissolution profile, crystalline form, and particle size distribution).

429

430 **Definitions**

431 **Clinical mastitis**

432 Clinical mastitis is defined as mastitis with clinical signs in one or more quarters (swelling, heat, pain,
433 redness) and changes in the appearance of milk (clots or flakes, watery appearance, discoloration),
434 with or without general signs (fever, loss of appetite).

435 **Mastitis**

436 Inflammation of one or more quarters of the mammary gland, almost always caused by infecting
437 microorganism.

438 **New infection**

439 Isolation of a pathogen from a mammary gland that has not previously been isolated from that
440 mammary gland or has not been isolated for some predetermined period of time.

441 **Subclinical mastitis**

442 Mastitis without clinical signs, but with an elevated milk somatic cell count in the quarter and isolation
443 of an udder pathogen from the milk.

444 **References**

445 VICH GL9: Guideline on good clinical practices (CVMP/VICH/595/1998)

446 VICH GL43: Guideline on target animal safety for veterinary pharmaceutical products
447 (CVMP/VICH/393388/2006), section 3.4. Mammary Gland Safety Studies

448 Local tolerance of intramammary preparations in cows (7AE21a Volume 7)

449 CVMP guideline: Demonstration of efficacy for veterinary medicinal products containing antimicrobial
450 substances (EMA/CVMP/627/2001) – *currently under revision*

451 CVMP guideline on fixed combination products (EMA/CVMP/83804/2005)

452 CVMP guideline on statistical principles for veterinary clinical trials for veterinary medicinal products
453 (pharmaceuticals) (CVMP/EWP/81976/2010)

454 CVMP guideline on Conduct of pharmacokinetic studies in target animal species
455 (EMA/CVMP/133/1999)