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4 **Guideline on data requirements for veterinary medicinal**
5 **products for the prevention of transmission of vector-**
6 **borne diseases in dogs and cats**
7 **Draft**

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42 **Executive summary**

43 This guideline provides recommendations for the design and conduct of studies to support the efficacy
44 of veterinary medicinal products (VMPs) intended for the prevention of transmission of vector-borne
45 pathogens (VBPs) in dogs and cats, which can be transferred by blood-feeding arthropods. The
46 guideline outlines the requirements for laboratory and field studies.

47 Prevention of transmission of vector-borne disease in the context of this guideline means the reduction
48 of the risk of transmission of VBPs by killing or repellent effect against the vector prior to the
49 transmission of the VBPs. This guideline, therefore, establishes criteria for the demonstration of
50 efficacy of a VMP in order to be granted a claim for the reduction of the risk of transmission of VBPs.

51 **1. Introduction (background)**

52 Vectors are living organisms, usually arthropods such as dipterans, fleas, lice, mites and ticks, that can
53 transmit pathogenic (micro)organisms from one infected host to another non-infected host (human or
54 animal), causing disease. Vector borne diseases (VBDs) are caused by a wide range of infectious
55 agents including viruses, bacteria and parasites (protozoa and helminths).

56 Given the zoonotic potential of some VBDs, there are 'One Health' considerations for the management
57 of ectoparasitic infestations in dogs and cats for protecting public health in addition to the health and
58 well-being of companion animals (Day, 2011; Mencke, 2013).

59 Veterinary medicinal products may provide indirect protection against VBPs by repelling or killing the
60 vector.

61 Across the EU, many ectoparasiticides are authorised as VMPs for the treatment of tick and flea
62 infestations in dogs and cats. The efficacy of these VMPs was assessed in accordance with the
63 'Guideline for the testing and evaluation of the efficacy of antiparasitic substances for the treatment
64 and prevention of tick and flea infestation in dogs and cats' (EMA/CVMP/EWP/005/2000). Similarly
65 ectoparasiticides authorised as VMPs for the treatment of lice, mites and diptera have been assessed
66 against the efficacy thresholds set in the guideline 'Demonstration of efficacy of ectoparasiticides'
67 (7AE17a, 1994). However, those guidelines do not give advice on how to design studies for the
68 demonstration of efficacy in the reduction of the risk of transmission of VBPs.

69 The repellent, insecticidal and/or acaricidal efficacy of a VMP demonstrated against a vector may not
70 be directly linked to the efficacy of the VMP in reducing the risk of VBDs. That is, a VMP that has
71 achieved the required threshold for efficacy sufficient for an insecticidal and/or acaricidal claim may not
72 be effective at reducing the risk of transmission of a VBP, as the vector may still have the ability to
73 transmit the pathogen causing the VBD before it is killed.

74 **2. Scope**

75 This guideline provides guidance on how to support an indication for a VMP for the reduction of the risk
76 of transmission of canine and feline VBPs transferred by blood-feeding arthropods.

77 This guideline applies to all applications where, according to Directive 2001/82/EC, new data has to be
78 generated to support clinical efficacy. Immunological VMPs and biocides are excluded from the scope of
79 this guideline.

80 **3. Legal basis**

81 This document should be read in conjunction with Directive 2001/82/EC. Applicants should also refer to
82 relevant European and VICH guidelines, including those listed among the references at the end of this
83 document.

84 In accordance with the provisions of the European Convention for the Protection of Vertebrate Animals
85 Used for Experimental and Other Scientific Purposes and Directive 2010/63/EC on the protection of
86 animals used for scientific purposes, the 3R principles (replacement, reduction and refinement) should
87 be applied whenever possible.

88 **4. General consideration**

89 For VMPs with insecticidal/acaricidal and/or repellent properties, a claim for the reduction of the risk of
90 transmission of VBPs needs to be demonstrated by laboratory and/or clinical field studies (irrespective
91 of the method of administration).

92 The underlying principle for the reduction of the risk is the killing and/or repellent effect against the
93 vector prior to the transmission of the VBP. In principle, the following aspects should be considered:

- 94 • Unless otherwise justified, it is expected that efficacy of the VMP against the defined vector(s)
95 will have been confirmed according to the requirements of existing guidelines (Vol. 7AE17a,
96 1994 and EMEA/CVMP/EWP/005/2000-Rev. 3). This implies that the mode of action and the
97 dosing regimen against the vector (repellent and/or killing effect, duration of persistent effect)
98 are known.
- 99 • An already authorised VMP with a new claim for the reduction of the risk of vector-borne
100 disease transmission should usually have the same dosing regimen as already authorised for
101 the acaricidal/insecticidal/repellent effect of the product. In case a change in the dosing
102 regimen (increased treatment frequency or dose) is necessary for this new claim, new
103 tolerance and dose justification study/ies should be provided.
- 104 • Unless otherwise justified, the efficacy of a VMP for the reduction of the risk of transmission of
105 VBPs has to be proven by appropriate clinical studies under laboratory and field conditions.
- 106 • Efficacy should be confirmed for each VBP transmission, extrapolation from one VBP species to
107 another is not acceptable.
- 108 • Clinical studies, whether laboratory trials or field studies, should be conducted according to
109 VICH GL9 Good Clinical Practices (GCP) or GLP.

110 Appropriate statistical methods in line with the CVMP guideline on statistical principles for veterinary
111 clinical trials (EMEA/EWP/81976/2010) should be applied in all clinical studies. It should be noted that
112 lower infection rates of the vectors might require higher numbers of study animals.

113 To evaluate the suitability of a claim on reduction of the risk of transmission of VBPs, the speed of
114 kill/repellent effect of the VMP against the ectoparasites from the dog or cat in relation to the
115 transmission time of a VBP from the vector to host is considered to be supportive information.

116 **4.1. General study design**

117 The reduction of the risk of transmission of canine and feline VBPs should be demonstrated under
118 defined conditions and performed with the final formulation for marketing administered at the
119 proposed dose and dosage interval confirmed to be effective against the vector (except the situation

120 where a change in dosing regimen is considered necessary; please see above). When designing a
121 clinical study the following aspects should be taken into consideration to ensure that the study will
122 provide conclusive information:

- 123 • The mode of transmission of a given VBP by its respective vector;
- 124 • The minimum time period of feeding by the infesting vector required for the transmission of
125 the specific VBP, if available;
- 126 • The time period from the transmission of a VBP until the presence of detectable infection
127 parameters (e.g. clinical signs, serological, antigen or DNA detection) in the final host animal,
128 for confirmation of the infection/disease in the host;
- 129 • Epidemiological considerations (e.g. the prevalence rate for the VBP in the vector population,
130 zoonotic potential);
- 131 • Appropriate method(s) to determine the infection/infectivity status of the vector should be
132 applied;
- 133 • Appropriate diagnostic method(s) to determine the infection/disease in the final host animal
134 and, if necessary, confirmatory methods should be applied.

135 The evaluation of the claimed effect should be based on the absence/presence of the VBP in the final
136 host animal, the antibody response of the final host animal, and/or the molecular detection of specific
137 DNA of the VBP in the final host animal (with or without clinical signs of the disease), as appropriate
138 for each VBP. The type of evaluation conducted may vary depending on the study design and the
139 availability of appropriately validated diagnostic procedures. The applicant should justify the diagnostic
140 approach used to demonstrate transmission after challenge or natural infection. With respect to the
141 claimed VBP, it is considered important to select appropriate time points for efficacy evaluation taking
142 into account current knowledge on diagnostic options and the biology of the disease.

143 **5. Laboratory trials**

144 ***5.1. Type of study***

145 Unless otherwise justified, at least one well-designed study under laboratory conditions covering the
146 entire period of reduction of the risk of disease transmission is considered necessary for each claimed
147 VBP. Studies should be performed in a parallel group design with a treated (test) group and an
148 untreated or placebo-treated group (negative control group). The inclusion of a negative control group
149 is considered necessary in order to confirm the validity of the test system. A rescue protocol needs to
150 be defined for all animals that have become infected.

151 ***5.2. Study animals***

152 The experimental animals should be clinically healthy and proven to be free of infections with the
153 targeted VBPs (e.g. immunologically naive, proven as sero- and/or PCR-negative). Methods confirming
154 the presence/absence of VBP/VBD in the animals before and after challenge should be justified and
155 validated with a sufficient diagnostic accuracy.

156 Included animals should not have been treated with an ectoparasitic substance within a time frame
157 that might impact on the study outcome. Whenever indicated (e.g. for ticks), animals should be tested
158 for their ability to carry adequate numbers of parasites prior to the start of the study. This pre-
159 allocation infestation should be done with uninfected vectors. The origin, sex, age, body weight and

160 type of hair coat of animals should be described. The housing conditions and group allocation should
161 follow the 'Guideline for the testing and evaluation of the efficacy of antiparasitic substances for the
162 treatment and prevention of tick and flea infestation in dogs and cats' (EMA/CVMP/EWP/005/2000
163 Rev.3).

164 **5.3. Treatment**

165 The VMP should be administered to the study animals of the test group before the first infestation with
166 the vector at an appropriate time point for preventing VBP transmission, i.e. when an adequate level of
167 efficacy is expected to have been reached (repellent/insecticidal/acaricidal) after product
168 administration. The appropriate time point may vary depending on the mode of action of the active
169 substance, and the pharmaceutical form (e.g. collar, spot-on solution).

170 The chosen treatment dose should preferably be the established minimum recommended treatment
171 dose (or a dose as close as possible to the minimum recommended dose).

172 **5.4. Information on the vector**

173 Laboratory studies may be performed either with laboratory-bred vector strains artificially infected with
174 a VBP or with vectors from infested habitats with a known prevalence of the VBP. The origin and
175 number of the vector(s) chosen for these studies should be described and justified. Methods used to
176 induce infections in the vector should be described in the study reports, taking into account their
177 reproducibility and validity.

178 For characterisation of the vector, the following parameters should be considered:

- 179 • Classical taxonomic determination, including data on the geographic origin of the particular
180 batch of used vectors;
- 181 • Molecular barcoding or similar relevant data confirming the species classification.

182 **5.5. Information on the vector borne pathogen**

183 The VBP used in experiments should be properly characterised, and the following data should be
184 provided:

- 185 • The biology of the VBP in the vector and in the host;
- 186 • Classical taxonomic determination including the data of the geographic origin;
- 187 • Molecular barcoding or similar relevant data confirming the species classification; such as DNA
188 sequences deposited in GenBank or similar depository.

189 The infection rate of the batch of vectors used for each challenge should be confirmed and justified
190 depending on the VBP. The number of vectors analysed to estimate the proportion of the batch
191 infected, the method to confirm the infectivity and all results should be reported and justified.

192 **5.6. Procedure of infestation**

193 A description of the infestation method of the study animals (both treated and untreated) with the
194 vector should be provided. The information should include the number of vectors per animal and the
195 time points for the initial and repeated challenges reflecting the period of the effect claimed (e.g. short
196 term up to 4 weeks, long term effect more than 4 weeks). For a VMP with short-term effect, it is
197 recommended to perform two challenges, one at the start and one close to the end of the claimed

198 protection period, while for a VMP with long-term effect, multiple challenges are required. In such
199 case, the time points of re-challenge should be justified. The number of challenges during the course of
200 the study should be kept as low as possible for animal welfare reasons without compromising the
201 integrity of the study. The experimental model should be robust enough to reflect field conditions in
202 terms of number of challenges, duration of the challenge, number of bites (exposures) in the field,
203 infection rate of the vector, climatic conditions, etc.

204 The vector should be left on or near the host animals for a time period which is known to ensure the
205 transmission of a VBP. The time point for removal of the vectors should be indicated. During the study
206 period, a defined number of untreated control animals, which have been infested with the vector at the
207 same time as the animals in the test group, should become infected with selected pathogen(s).
208 Appropriate measures should be applied to reduce any negative impact on animal welfare (e.g.
209 appropriate exit clauses and rescue protocols; see 5.1).

210 **5.7. Evaluation of efficacy**

211 The primary efficacy parameter should be the relative blocking efficacy of the test product on VBP
212 transmission. Possible secondary criteria should also be defined. A definition of success (non-infected
213 host animals, which are free of infections with the targeted VBPs; e.g. immunologically naive, proven
214 as sero- and/or PCR-negative) and failure (infected host animals, in which the presence of VBP has
215 been confirmed) should be provided.

216 The difference in the proportions of infected animals in the treatment group and in the untreated
217 control group must be statistically significant.

218 Depending on the study design, there are several possible approaches for calculating the blocking
219 efficacy. The efficacy threshold should be $\geq 90\%$.

220 The recorded efficacy should be presented together with its confidence interval.

221 The following example is a common approach:

222 *Calculation of the blocking efficacy based on infective vector challenges*

223 This approach is independent of the number of animals in the study groups and calculates the
224 percentage of protection in comparison to the number of consecutive pre-infection challenges.

$$225 \text{ Protection (\%)} = 100 \times \frac{IcC - IcT}{IcC}$$

226 Where

- 227 • IcC = the “infection proportion” calculated as the number of infected animals in the untreated
228 control group divided by the total number of pre-infection challenges with vectors from a batch
229 infected with the pathogen in the **untreated control group**
- 230 • IcT = the “infection proportion” calculated as the number of infected animals in the treatment
231 group divided by the total number of pre-infection challenges with vectors from a batch
232 infected with the pathogen in the **treatment group**.

233 **6. Field trials**

234 Unless otherwise justified, field trials should be conducted and may constitute pivotal data where no
235 valid laboratory transmission model is available. The trials should be conducted with naturally infected
236 vectors/animals, and should be randomised, blinded and controlled.

237 The studies should be conducted in Europe in different geographical locations. The applicant should
238 provide information on the seasonal prevalence rate for the VBP in the vector population, considering
239 the seasonal occurrence of the vector parasites, ecological conditions and lifestyle/ husbandry of target
240 animals (see also 6.1).

241 **6.1. Information on study animals**

242 Only animals tested negative for both the VBP and the relevant immune response should be considered
243 for efficacy evaluation. Animals that, at study enrolment, tested positive for the VBP of concern should
244 be excluded from efficacy evaluation for that specific VBP. The detection methods used should
245 preferably be based on generally acknowledged procedures combining direct and indirect diagnostic
246 tools, e.g. pathogen detection (e.g. by PCR, culture or staining like immunofluorescence) or antibody
247 detection.

248 **6.2. Infestation and evaluation of infestation level**

249 Under field conditions, the animals will be exposed to naturally infected vectors. Before the first
250 expected exposure of the enrolled animals to the vectors, the product should be administered at an
251 appropriate time point for preventing VBP transmission, i.e. when an adequate level of efficacy is
252 expected to be reached after product administration.

253 The chosen treatment dose should preferably be the established minimum recommended treatment
254 dose (or a dose as close as possible to the minimum recommended dose) and the established dosing
255 interval.

256 When the efficacy of blocking the transmission of a VBP by a vector is intended to be demonstrated
257 under field conditions, a control group (positive or negative) should be included. In those cases where
258 laboratory data are not fully supportive of prevention of transmission, the inclusion of an untreated
259 (negative) control group in the field study is considered necessary, to obtain information on the real
260 transmission pressure. An adequate number of initially VBP-free animals in the untreated control group
261 should be diagnosed positive for a VBP infection compared to the treated animals to ensure the validity
262 of the study. A rescue protocol needs to be defined for all animals that have become infected.

263 **6.3. Evaluation of efficacy**

264 Under field conditions, it is recommended to calculate the efficacy by comparing the incidence of
265 infection of the VBD in the treated and the control group. The time period should be appropriate, and
266 the lifecycle and prevalence of the vector (e.g. a year, a season or a month) should be taken into
267 consideration.

268 A) The reduction of the risk of VBPs transmission (%) for a defined time period calculated as follows:

269 The reduction of the risk of VBPs transmission (%) =
$$\frac{\text{incidence in the control group} - \text{incidence in the treated group}}{\text{incidence in the control group}} \times 100$$

270 The incidence (%) should be calculated for each group as follows:

271
$$\frac{\text{no. of newcases of infected animals}}{\text{no. of negative animals initially enrolled} - \text{no. of lost or dead animals}} \times 100$$

272 B) To overcome problems with dogs lost to follow-up during field studies (e.g. death, withdrawal, etc.),
273 and to account for potential differences in time spans during which the animals were included in the
274 study; the incidence density rate (IDR) could be used.

275 The IDR is defined as the number of newly infected cases per population at risk during a given follow-
276 up time period calculated in animal-time. IDR is calculated by dividing the number of new cases by the
277 number of animal-time as follows:

$$278 \text{ IDR (per 100 cases per time)} = \frac{\text{number of new cases}}{\text{animal - time (years)}} \times 100$$

279 "Animal-time" is the sum of the periods of observation for each animal during which the animal is free
280 from the disease (i.e. is at risk). As soon as an animal becomes diseased, it no longer contributes to
281 this value. Usually IDR is expressed in years but the IDR can also be calculated on a monthly basis.

282 Animals that were tested only once at D+0 after which they were excluded then from the study (e.g.
283 lost, dead), do not contribute at any time to the incidence calculation.

$$284 \text{ Protection (\%)} = 100 \times \frac{\text{IDRc} - \text{IDRt}}{\text{IDRc}}$$

285 IDRc = the IDR in the control group

286 IDRt = the IDR in the treated group.

287 Concerning the methods of efficacy evaluation, the reduction of the risk of VBPs transmission % should
288 be ≥90%. The recorded efficacy should be presented together with its confidence interval.

289 The proportions of infected animals between the treatment and negative control group must differ
290 statistically significantly. For positively controlled field studies, non-inferiority or superiority to an
291 authorised veterinary medicinal product with recognised efficacy in the indication concerned should be
292 demonstrated.

293 **7. Summary of product characteristics (SPC)**

294 The SPC draft should take into account the guidance in the Notice to Applicants (Volume 6C).

295 **Section 4.2 (Indications for use)**

296 This section should list the specific name of the VBDs against which the product has demonstrated
297 sufficient efficacy (effective against VBP transmission), as confirmed in the documentation of part 4,
298 e.g.

299 *"Reduction of the risk of infection with <name of VBP> via transmission by <name of vector> for up to*
300 *<time period reflecting the proposed recommended treatment interval for efficacy against the*
301 *pathogen transmission>. The effect is indirect due to product's activity against the vector.";* or
302 *"By [repelling and/or killing] the vector [name of the vector], the product reduces the risk of*
303 *transmission of the VBP [name of the pathogen]."*

304 **Section 4.4 (Special warnings for use)**

305 A warning for the animal owner should be included that the risk of VBP transmission cannot be
306 completely excluded, e.g. by using the following example:

307 *"After treatment <an attachment of single ticks> <feeding of single insect (fleas, phlebotomus etc.)>*
308 *cannot be excluded (for locally acting products) or a transmission of infectious diseases cannot be*
309 *completely excluded since the <arthropod> <insect> has to attach to the host before achieving the*
310 *ectoparasitocidal effect (for systemically acting products)."*

311 **Section 4.5 (Special precautions for use)**

312 In context with data on the onset of effect or with the extrapolation from data on the speed of kill, the
313 following warning for owner should be included if necessary:

314 *"The protection may be insufficient the <first hours/ days> after administration of the product."*

315 **Section 5.1 (Pharmacodynamic properties)**

316 Relevant information about the clinical trial(s) might be included in this section. However, if relevant,
317 appropriate information should be mentioned in section 4.4 or 4.5, as appropriate.

318 **Definitions**

319 **Vectors:** Vectors are living organisms that can transmit disease-causing organisms; that is, they can
320 transmit VBPs from an infected host to an uninfected host.

321 **Vector-borne diseases (VBD):** Vector-borne diseases are diseases caused by pathogens that are
322 transmitted by a vector. The vector may merely be a passive carrier of the infectious agent, but many
323 kinds of infectious agents undergo a stage of biological development in the vector, known as biological
324 vectors. In this case, both the vector and the host are essential for the multiplication and life cycle of
325 the infectious agent.

326 **Vector-borne pathogens (VBP):** Vector-borne pathogens are disease-causing organisms that are
327 transmitted between their hosts by arthropod vectors. Vector-borne pathogen transmission occurs
328 when host, vector and pathogen interact in space and time within a permissive environment.

329 **Pre-infection challenge:** For each animal the number of consecutive challenges with an infected
330 vector population until the transmission of a pathogen has been diagnosed.

331 **Incidence density rate:** The number of new cases of an infection at risk in a given follow-up time
332 period calculated in animal-time.

333 **References**

334 CVMP Guideline on the demonstration of efficacy of ectoparasiticides; Vol. 7AE17a, 1994.

335 CVMP Guideline for the testing and evaluation of the efficacy of antiparasitic substances for the
336 treatment and prevention of tick and flea infestation in dogs and cats (EMA/CVMP/EWP/005/2000-Rev
337 3).

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339 CVMP Guideline on the conduct of pharmacokinetic studies in target animal species
340 (EMA/CVMP/133/99)

341 CVMP Guideline on statistical principles for veterinary clinical trials (CVMP/EWP/81976/2010)

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344 Directive 2001/82/EC of the European Parliament and of the Council as amended on the Community
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346 Directive 2010/63/EC of the European Parliament and of the Council as amended on the protection of
347 animals used for scientific purposes.

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350 veterinary medicinal view on CVBDs such as tick borreliosis, rickettsiosis and canine leishmaniosis.
351 Vet Parasitol 195, 256-271.

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