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4 **Guideline for the demonstration of efficacy for veterinary**
5 **medicinal products containing antimicrobial substances**
6 **Draft**

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8 This guideline will replace the CVMP guideline for the demonstration of efficacy for veterinary medicinal
9 products containing antimicrobial substances ([EMA/CVMP/627/2001](#))

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11 Comments should be provided using this [template](#). The completed comments form should be sent
to vet-guidelines@ema.europa.eu



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45 **Executive summary**

46 This guideline provides recommendations for the design and conduct of pre-clinical and clinical studies
47 to support clinical efficacy for an antimicrobial¹ veterinary medicinal product. Appropriate methods to
48 identify and describe the pharmacology of the active substance in relation to the target bacteria are
49 presented and important aspects to consider for justifying the use of a certain active substance for a
50 particular indication are outlined. Advice regarding study design, selection of comparator and efficacy
51 endpoints is given for the purpose of gaining conclusive study results for the intended claim which
52 could be treatment, treatment and metaphylaxis, or prevention. Alternative study designs may be
53 applied if justified.

54 **1. Introduction (background)**

55 The objective of this guideline is to specify the data required to demonstrate the therapeutic efficacy of
56 a veterinary medicinal product (VMP) containing an antimicrobial agent for (a) given indication(s) using
57 an appropriate therapeutic regimen. Thus, the following sections provide guidance on the essential
58 topics which the applicant should cover in order to demonstrate efficacy i.e. pharmacodynamics
59 (including resistance mechanisms), pharmacokinetics and clinical trials. In the context of this guideline
60 an antimicrobial is defined as a substance primarily acting against bacteria.

61 **2. Scope**

62 This guideline applies to antimicrobial substances used in veterinary medicines for all routes of
63 administration and to all pharmaceutical forms. For antimicrobials intended for intramammary
64 administration the pharmacology section (except the PK/PD section, 5.7) of this guideline applies
65 whereas regarding clinical issues the CVMP guideline for the conduct of efficacy studies for
66 intramammary products for use in cattle (EMA/CVMP/344/99) would only have to be considered.
67 Cross-reference is also made to the CVMP guideline on the conduct of pharmacokinetic studies in
68 target animal species (EMA/CVMP/133/99).

69 For fixed combinations please see also the CVMP guideline on pharmaceutical fixed combination
70 products (EMA/CVMP/83804/2005).

71 This guideline applies to all applications where according to Directive 2001/82/EC, as amended new
72 data has to be generated to support clinical efficacy.

73 The guideline does not apply to products containing an antimicrobial agent if the indication is not for
74 combating a bacterial infection. However, for such products safety issues like development of
75 resistance needs to be addressed as outlined in this and other relevant guidelines.

76 **3. Legal basis**

77 This guideline replaces the current CVMP guideline for the demonstration of efficacy for veterinary
78 medicinal products containing antimicrobial substances ([EMA/CVMP/627/2001](#)), and should be read in
79 conjunction with Directive 2001/82/EC. Directive 2010/63/EC regarding the protection of animals used

¹ Antimicrobial agent: A naturally occurring, semi-synthetic or synthetic substance that exhibits antimicrobial activity (kill or inhibit the growth of micro-organisms) at concentrations attainable *in vivo*. Antiparasitics and substances classed as disinfectants or antiseptics are excluded from this definition (OIE Terrestrial Animal Health Code definition). In the context of this guideline the focus is on compounds acting against bacteria.

80 for scientific purposes also applies. Applicants should also refer to other relevant European and VICH
81 guidelines, including those listed in the reference list of this document.

82 **4. General considerations**

83 Antimicrobials are powerful and important tools to combat bacterial infections in animals. However, all
84 use will inevitably select for antimicrobial resistance. Thus it is vital that all unnecessary or inadequate
85 use is avoided, in order to prolong the time period during which the compound will remain effective. In
86 addition, potential risks to public health need to be considered such as outlined in separate guideline
87 (VICH GL27).

88 The following is to be specifically addressed in order to justify the need/s and selection of an
89 antimicrobial in relation to the indication investigated in the clinical development program:

- 90 • The indication should be justified. Use of antimicrobials for treatment of mild and transient
91 infections that will resolve independent of treatment will be questioned. In case of multi-factorial
92 diseases, efforts should be made to describe the expected contribution from the antimicrobial
93 treatment, e.g. through reference to published information, and studies should be designed
94 considering where and when there is a place for an antimicrobial in the treatment strategy.
- 95 • The target population for therapy should be well defined and possible to identify under field
96 conditions. The study population in field trials should reflect the intended target population for
97 therapy to the best possible extent.
- 98 • Official guidance on preferred choices of antimicrobials to be used and those to be reserved for
99 certain conditions such as CVMP recommendations² (when available) should be considered, with
100 an intention to obtain the best achievable alignment between the study population and the target
101 population for treatment. Any deviation from official guidance recommendations should be
102 justified.
- 103 • The dose and the dosing interval of the antimicrobial product can be justified by considering the
104 pharmacodynamic/pharmacokinetic (PK/PD) relationship, if established, as well as the severity of
105 the disease, whereas the number of administrations should be in line with the nature of the
106 disease. To avoid unnecessary exposure to antimicrobials (and thus unnecessary selection
107 pressure for resistant bacteria), the duration of exposure should not be longer than necessary to
108 accomplish the desired outcome.

109 **5. Pharmacology**

110 The pharmacokinetic and pharmacodynamic properties of the active moiety should be adequately
111 documented.

112 For the conduct of pharmacokinetic studies please see the CVMP guideline on conduct of
113 pharmacokinetic studies in target animal species (EMA/CVMP/133/99). Studies on pharmacodynamics
114 should be performed according to validated and internationally accepted methods, and according to
115 Good Laboratory Practice (GLP), when applicable. Measures should be in place to ensure any negative
116 impact on animal welfare is kept to a minimum. Data requirements are detailed below.

2

http://www.ema.europa.eu/ema/index.jsp?curl=pages/regulation/general/general_content_000384.jsp&mid=WC0b01ac058002dd37#Antimicrobials

117 **5.1. Antimicrobial class**

118 The antimicrobial class should be stated.

119 **5.2. Mode and mechanism of action**

120 The mode and mechanism of action of the antimicrobial substance on the target bacteria should be
121 reported.

122 The spectrum of the antimicrobial activity of the substance should be defined. Naturally resistant
123 bacterial species relevant to the intended use of the veterinary medicinal product should be reported.

124 **5.3. Minimum Inhibitory Concentration (MIC)**

125 The minimum inhibitory concentration (MIC) is the lowest concentration (expressed in µg/ml or mg/l)
126 of an antimicrobial substance which, under defined *in vitro* conditions, prevents the visible growth of
127 bacteria. MIC values should be determined using accepted standardised methodology, such as those
128 described in CLSI³ documents. Dilution methods, when available, should be used and the methods
129 clearly described. However, it is recognised that to date standardised methodologies are not available
130 for all organisms. MIC data should be provided for all target bacteria. A scientifically justified number
131 of clinical isolates of each target bacteria, representative of the EU area, should be collected, to allow
132 detection of isolates with MICs deviating from the normal distribution of isolates without any acquired
133 resistance (wild type). For rare pathogens a lower number of isolates could be justified than for
134 commonly encountered pathogens. The isolates of the target bacteria to be tested should have been
135 collected within five years prior to the submission of the application. Isolates should be
136 epidemiologically unrelated (not coming from the same episode of disease in the same herd or same
137 animals) and constitute a representative sample from within the EU. For bacteria isolated from food-
138 producing animals, selection of livestock farms should include units of different types. In these cases,
139 the tested isolates should preferably come from the animal subgroup(s) or production type(s) that
140 reflect the target population for the indication. The origin of the isolates investigated (animal species,
141 condition, farm type, geographic area) and dates of collection should be stated.

142 The susceptibility for antimicrobials varies not only between different bacterial species but also
143 between strains and over time. The complete MIC distribution data for all isolates tested of each
144 bacterial species should be reported in tables and if relevant, divided by subgroups. In case the MIC
145 distribution indicates the presence of subtypes of bacteria with reduced susceptibility (bi or multimodal
146 distribution), these should be compared with already available (historical) data to allow conclusions to
147 be drawn on mechanisms for acquired resistance. Based on these conclusions and supportive clinical
148 data, the subpopulation with reduced susceptibility may be included in the intended population to be
149 treated. It is acknowledged that for historical data information of the full distribution may not be
150 available. In such cases all available data such as MIC₅₀ and MIC₉₀ should be provided.

151 The data on MIC distribution should be interpreted using adequate interpretation criteria. The
152 epidemiological cut-off value⁴ should be determined, if feasible, to define the population without any
153 acquired resistance. The epidemiological cut-off value may be proposed as the clinical breakpoint but
154 should always be supported by PK/PD and clinical data. In case a population with reduced susceptibility
155 is identified, the applicant can suggest a clinical breakpoint (i.e. a MIC value under which the selected

³ CLSI - Clinical and Laboratory Standards Institute

⁴ For definitions of epidemiological cut-off values and clinical breakpoints, please refer to EUCAST.

<http://www.srga.org/Eucastwt/eucastdefinitions.htm>

156 dose is shown efficient) deviating from the epidemiological cut-off value. Any clinical breakpoint must
157 be supported by microbiological, clinical and available PK/PD data. In case reference is made to a
158 clinical breakpoint established by an external institute or published in literature it should be
159 demonstrated that this value is relevant for the product under study.

160 It is recommended to include also major metabolites contributing significantly to the antimicrobial
161 activity in the *in vitro* susceptibility testing.

162 **5.4. Minimum Bactericidal Concentration (MBC) and kinetics of bacterial** 163 **killing**

164 MBC is the lowest concentration of an antimicrobial substance (expressed in µg/ml or mg/l) which,
165 under defined *in vitro* conditions, reduces bacterial counts by 99.9%.

166 Data on the kinetics of bacterial killing should be provided to characterize the action of the
167 antimicrobial against the target bacteria and to demonstrate whether its antimicrobial activity is
168 bacteriostatic or bactericidal and whether it is time-dependent (i.e. dependent upon the period of time,
169 during which the concentration of the antimicrobial substance exceeds the MIC, but for which
170 concentrations of several magnitudes of the MIC do not increase efficacy), concentration dependent
171 (i.e. efficacy increases when administered at doses which confer concentrations several times the MIC)
172 or co-dependent (i.e. which depends both upon concentrations above the MIC and the period of time
173 during which the concentration of the antimicrobial substance exceeds the MIC). This can be bacterium
174 or condition specific and data should be provided for different target pathogens when appropriate.
175 Where available, publications providing information on the pharmacodynamic activity of the
176 antimicrobial can be used as supportive information. Kinetics of bacterial killing should be performed
177 according to validated and internationally accepted methods. The clinical relevance of claimed
178 bactericidal activity against certain target bacteria should be discussed.

179 **5.5. Resistance**

180 Isolates with MICs deviating from the normal distribution for a certain antimicrobial class should be
181 tested for co-resistance and cross-resistance.

182 Where possible, information on the resistance mechanism(s), the molecular genetic basis of resistance
183 and on the rate of transfer of resistance determinants should be provided and discussed. This
184 information may come from literature or proprietary studies and may derive from related antimicrobial
185 substances in the absence of data on the specific substance.

186 Cross-reference can be made to the information supplied in accordance with the VICH GL 27: Guidance
187 on the pre-approval information for registration of new veterinary medicinal products for food
188 producing animals with respect to antimicrobial resistance.

189 **5.6. Additional in-vitro studies**

190 Additional *in vitro* studies should, whenever relevant, include an investigation of possible synergy or
191 antagonism and may include, for example, investigation of post-antibiotic effects and, for certain
192 antibacterial agents, an estimate of the rate of selection of resistant mutants and how concentrations
193 above the MIC may affect or prevent selection of mutants. The methods for additional studies should
194 be well described and the clinical relevance of the obtained results should be justified.

195 Some environmental factors (e.g. pH, O₂, inhibitors, cation concentration) may influence the
196 antimicrobial activity at certain sites of infection and in biological fluids. When available and if relevant

197 to the proposed indications for use of the antimicrobial substance, these data should be reported. The
198 clinical relevance of the environmental factors should be discussed.

199 **5.7. The pharmacokinetic/pharmacodynamic (PK/PD) relationship**

200 To be effective, the dose of an antimicrobial agent must be selected considering the susceptibility of
201 the target bacteria. Therefore, for all compounds with systemic activity, the MIC data collected should
202 be compared with the concentration of the compound at the relevant biophase following administration
203 at the assumed therapeutic dose as recorded in the pharmacokinetic studies.

204 Based on *in vitro* susceptibility data, and target animal PK data, an analysis for the PK/PD relationship
205 may be used to support dose regimen selection and interpretation criteria for a clinical breakpoint. In
206 circumstances in which it is not feasible to generate extensive clinical efficacy data (e.g. in rare types
207 of infections or against rare types of pathogens, including multidrug resistant pathogens that are rarely
208 encountered) PK/PD analyses may also provide important supportive information on the potential
209 efficacy of the test antibacterial agent.

210 The overall assessment of the PK/PD relationship should be sufficiently comprehensive to assess with a
211 reasonable confidence whether or not the test antibacterial agent, when used at an adequate dose
212 regimen, would show clinical efficacy against claimed target pathogens that appear to be susceptible *in*
213 *vitro*.

214 It is acknowledged that the PK/PD analyses will be based on PK data obtained from healthy or
215 experimentally infected animals. Nevertheless, the sponsor is encouraged to collect PK data from
216 naturally diseased animals using population kinetic models. Knowledge of kinetic variability
217 considerably increases the value of the PK/PD analysis.

218 In some cases where the PK/PD relationship is well established using validated models, it may be
219 possible to omit dose-determination studies and to evaluate in a clinical trial the efficacy of one or a
220 very few regimens. However, to be acceptable the choice of the PK/PD parameter considered as best
221 predictive of efficacy must be prospectively justified. In addition, the use of PK/PD to predict the
222 optimal duration of treatment is not well established at present and sponsors should consider whether
223 preliminary regimen-finding studies are needed to identify a suitable duration of treatment for any one
224 indication.

225 Currently the most commonly used parameters to express the PK/PD relationship are C_{max}/MIC
226 (maximum concentration in serum or plasma/MIC), %T > MIC (fraction of time during which the
227 concentration exceeds the MIC) and AUC/MIC by convention referred to as AUIC (area under the
228 inhibitory curve). Use of free (unbound) fraction is normally required for calculation of PK/PD
229 parameters. Further characterisation of PK/PD parameters should be specified according to the
230 antimicrobial and microorganism under investigation.

231 PK data from other matrices than plasma might be used provided there are validated models available.

232 **6. Clinical studies**

233 **6.1. General Principles**

234 It is recommended to conduct preclinical and clinical studies according to Good Laboratory Practice
235 (GLP) and/or Good Clinical Practice (GCP). In case GLP and/or GCP is not applied, traceability and
236 integrity of data should be adequately guaranteed by other means. For clinical field trials, GCP status is
237 required.

238 All studies should be controlled studies and the choice of control should be justified. For treatment
239 claims, one clinical study using a negative control group should be provided preferably as a minimum,
240 unless the superiority of the product is proven otherwise. The “3R-principles” (replacement, reduction,
241 refinement) should always be applied whenever possible.

242 Clinical trials should cover each proposed indication and bacterial species in each target animal species
243 claimed. The number of clinical trials will depend on the type of veterinary medicinal product and
244 nature of the disease. Several controlled trials are generally required dependent on the size and quality
245 of studies conducted.

246 Appropriate statistical methods should be used (see CVMP guideline on statistical principles for
247 veterinary clinical trials, CVMP/EWP/81976/2010).

248 The product formulation used should be the one proposed for authorisation. Any deviation should be
249 justified. If the formulation used in the field trials differs from the final formulation, the relative
250 bioavailability should be documented.

251 The method of determining the disease and clinical condition of the animals should be appropriate and
252 fully described. Whenever possible, established methods for diagnosis should be applied.

253 Principally there are three different kinds of claims: treatment, metaphylaxis and prevention (for
254 definition: see glossary). A metaphylaxis claim can only be accepted in conjunction with a treatment
255 claim.

256 **6.2. Dose-determination studies**

257 Detailed information about an adequate therapeutic scheme for each bacterial species and claim should
258 be collected from experimental studies performed under controlled conditions. Dose determination
259 studies encompass dose level, dosing interval and number of administrations. They are important to
260 ensure efficacy of the product without unnecessary exposure to the compound.

261 Dose determination studies should always include a negative control. Appropriate measures should be
262 applied to reduce any negative impact on animal welfare. Group sizes of negative controls should be
263 the minimum required to produce meaningful data. If acute clinical signs of disease are expected,
264 monitoring should be focused around the peak of expected effect.

265 Consideration should be given to study designs that incorporate more than one of the parameters
266 mentioned above (dose level, dosing interval, number of administrations) so as to reduce the number
267 of negative control groups used overall in dose-determination studies. If it is not feasible to perform
268 studies to explore different dosing strategies other data could be used as support. Regarding the dose
269 level and the dosing interval, the PK and PD characteristics of the product should be considered to
270 support the necessary exposure and consequently achieve a satisfactory balance between efficacy and
271 risk for selecting for resistance. The recommended treatment duration could be justified on basis of the
272 time course of disease progress. In addition, when available, data from published clinical studies
273 comparing different dosing regimens for a similar product or of the same substance class with a similar
274 activity may be used to support the need for any certain duration of exposure to the active substance.

275 Where possible, experimentally induced infections should be used in the dose-determination studies.
276 The origin and *in vitro* susceptibility of the strains used in the study should be presented (see section
277 5.4). A strain representative of the wild type population (e.g. fully susceptible towards the substance
278 under study) could be used. However, if a claim is made for bacteria where reduced susceptibility
279 relative to the wild type is common, this should be taken into account when selecting the test strain. In
280 case several primary pathogens will be included in the sought indication (e.g. bovine respiratory

281 disease) dose determination should be based on the least susceptible species as evident from relevant
282 data. If this is not possible due to lack of an established experimental model, conclusive information
283 regarding the treatment effect for the least susceptible species needs to be presented from dose
284 confirmation and/or clinical field studies. The validity of the experimental models used should be
285 justified with regard to their capability to establish infection and cause a clinical disease similar to that
286 in naturally infected animals. For treatment claims, the drug administration should normally not be
287 initiated before the clinical signs relating to the bacterial infection are observed. However, initiation of
288 treatment before clinical signs appear may be acceptable in case of per-acute disease, when a
289 validated model is available that justifies this procedure.

290 If no experimental model is available and study conditions are well controlled, naturally infected
291 animals can be used.

292 When appropriate, it is recommended to include PK data in dose finding studies to allow the recorded
293 effects to be related not only to dose but more specifically to time-concentration curves during
294 treatments.

295 Usually three levels of dosage of the veterinary medicinal product should be tested, preferably using
296 the final formulation. The aim is to demonstrate that the chosen dose provides sufficient efficacy
297 without over-exposure.

298 Efficacy evaluation should be based on clinical and bacteriological response as determined by
299 appropriate clinical and bacteriological assessment. Mortality should be assessed and *post mortem*
300 data should be added wherever meaningful. The endpoints (primary and secondary) and timing of the
301 response assessment used should be justified in relation to the disease and substance under study.
302 Observations should be collected repeatedly before, during and after treatment, as appropriate. The
303 time of response assessment should be selected so as to show the effect of treatment in a relevant
304 matter as compared to the negative control, thereby taking into account the effect of the treatment
305 and the natural course of the disease.

306 Statistical comparisons between different treatment groups and the negative control group should be
307 provided if possible, although it is acknowledged that dose determination studies are often not
308 designed to generate statistical support and thus conclusions are often based on descriptive
309 information.

310 From the results of dose determination studies, the applicant could decide upon an appropriate dosage
311 regimen which should be pursued in confirmation studies and subsequent field trials.

312 **Locally acting products**

313 The dosing regimen should be substantiated also for locally active products. For formulations applied
314 directly to the infection site and which do not undergo significant dilution, a justification can be
315 sufficient, taking into account the product strength, the formulation and *in vitro* susceptibility data for
316 the target bacteria. In other cases, such as e.g. locally active products for the gastro-intestinal tract
317 clinical dose finding studies should be performed as detailed in the previous section. Notably, the
318 extent of systemic absorptions is one aspect limiting the upper dose for a locally active compound. For
319 details on intramammary products, please see the CVMP guideline for the conduct of efficacy studies
320 for intramammary products for use in cattle (EMA/CVMP/344/99-FINAL-Rev.1). Regarding
321 systemically administered products intended to combat a localized infection (e.g. metritis) the dose
322 should be established according to recommendations given in the previous section.

323 **6.3. Dose-confirmation studies**

324 The aim of dose-confirmation studies is to confirm the efficacy of the selected dosage regimen in
325 individual animals (treatment claims) or groups of animals (including metaphylaxis claims) under
326 controlled clinical conditions. These studies can be performed using experimental models of infections
327 but well controlled studies using naturally infected animals are preferred.

328 A study should preferably include a negative control group. Appropriate measures should be applied to
329 reduce any negative impact on animal welfare. In treatment claims where the use of a negative control
330 is not possible an appropriate positive control may be acceptable provided internal validity and
331 sensitivity of the study is ensured (CVMP/EWP/81976/2010).

332 Efficacy criteria used to assess the outcome of disease and/or infection are similar to those for dose-
333 determination studies. The primary endpoint(s) should preferably be the same as the one(s) intended
334 for use in the field trials.

335 Dose confirmation studies may allow for the assessment of relapse among animals that were
336 considered successfully treated at the time of primary efficacy assessment. A high relapse rate could
337 indicate that the treatment was not sufficiently effective to combat the infection. The objective is to
338 distinguish between relapse and re-infection; therefore, if it is clinically feasible then the study design
339 should accommodate this. It is acknowledged, however, that relapse rate assessment may be based on
340 clinical signs and that microbiological analysis is not performed on all animals with the consequence
341 that relapse and re-infection cases cannot always be fully separated. An appropriate assessment time
342 point for relapse rate assessment would be after concentrations of the active substance have
343 decreased below therapeutic concentrations in plasma or in the target tissue and the risk for including
344 re-infected animals is still low. The time selected should be justified in this respect. (For definition of
345 relapse and re-infection, see glossary).

346 It can be acceptable to waive dose-confirmation studies provided all of the following criteria are
347 fulfilled:

- 348 • the conditions of the dose determination studies are representative of the field conditions in terms
349 of the type of infection and the animals involved,
- 350 • the susceptibility pattern for any challenge strain used for dose determination is relevant for the
351 field situation,
- 352 • a clear dose-effect relationship is documented as supported by adequate dose determination data,
- 353 • the dose determination data allows for the selection of one appropriate dose level,
- 354 • the dosing interval and the number of administrations is adequately justified.

355 At least one dose confirmation study must be presented if the dose finding is based on *in vitro* PD data
356 only.

357 For group/flock medication via water or feed, the variability between animals in feed/water intake
358 should be explored as far as possible, with the purpose of ensuring that the dose selected will provide
359 therapeutic exposure levels in all animals. Population PK/PD models (such as Monte Carlo simulations)
360 based on data from field trials could be used to bring support for a post-hoc analysis of the selected
361 dose.

362 **6.4. Field trials**

363 **6.4.1. Study design and population**

364 Field trials should be multicentric, randomized, blinded and controlled, and conducted in naturally
365 infected animals. For a given indication, the study population should be well defined, and
366 representative of the intended target population for therapy. This includes considerations regarding
367 housing conditions, production types and geographical location. Furthermore, the sample size should
368 be determined according to appropriate statistical principles (CVMP guideline on statistical principles
369 for veterinary clinical trials, CVMP/EWP/81976/2010). Blinding of the study needs to be ensured
370 through appropriate study design measures such as dummy treatment, if necessary.

371 **6.4.2. Control**

372 **Negative control**

373 Including a placebo or an untreated control group may be of value in situations where a high self-cure
374 rate could be suspected since the risk for erroneous conclusions is present in these situations.
375 Negatively controlled studies can also be useful when there are no approved products for the indication
376 in question to serve as control, and in infections with bacteria resistant to previously authorised
377 substances. A negatively controlled study is normally necessary to support a prevention claim and in
378 some situations also to gain support for a metaphylactic claim (see separate sections).

379 The welfare of animals in the study must be given the highest priority, e.g. through the establishment
380 of appropriate exit clauses and rescue protocols.

381 **Positive control**

382 A positive control should be a product authorised under Directive 2001/82/EU, as amended, for the
383 same indication. The applicant should pay attention to that the chosen control product needs to be
384 sufficiently effective for the target indication at the time the study is conducted.

385 Susceptibility of the target pathogens might differ between regions and over time. Products for which
386 recent susceptibility data suggest that posology may be inadequate for the infection under study, or
387 products where posology differs between member states should be avoided. A comparator should
388 always be used according to the label instructions.

389 Since it is of vital importance that the positive control is appropriate it is recommended that advice is
390 sought from the authorities if applicants are not sure if their proposed control product would be
391 suitable.

392 When a study is performed to explore non-inferiority of the test product, appropriateness of the study
393 design should be ensured and the non-inferiority limit should be pre-specified and justified from a
394 clinical relevance perspective, according to the statistical principles outlined in the relevant CVMP
395 guideline (CVMP/EWP/81976/2010). It should further be ensured that the current study design is
396 appropriate in the sense that it can be reliably expected that a recognized level of efficacy will be
397 demonstrated for the control treatment.

398 In case the aim is to demonstrate superiority to an authorised product it has to be ensured that the
399 positive control is a relevant treatment alternative for the current indication at the time of investigation
400 (see above). This would include the presentation of susceptibility data for the control to ensure that
401 any difference is not dependent on resistance development.

402 A superiority trial including an existing control product is a valuable means to support efficacy where
403 the target population corresponds to clinical conditions of particular severity and where there is
404 reasons to suspect that approved products would not be sufficiently effective.

405 **6.4.3. Inclusion criteria**

406 Clinical trials should incorporate strictly defined clinical and microbiological inclusion criteria as
407 appropriate for the claimed indication. Susceptibility of the isolated bacteria to the test product (and to
408 the control product, where applicable) should be tested *in vitro*.

409 When the aim is to confirm efficacy against one or several specified bacteria, isolation of the target
410 pathogen(s) from the animals or a representative proportion of them is required through
411 microbiological sampling performed at the time of inclusion.

412 If individual bacteriological testing of all included animals is not feasible (e.g. herd treatment), the
413 sample size should be large enough to allow confirmation of the etiological diagnosis with sufficient
414 level of certainty. For those animals which are included on basis of clinical signs of disease only, the
415 causal relationship to the target bacterium should be made evident through appropriate clinical
416 diagnostic criteria.

417 The microbiological sampling technique used on all or a proportion of the study animals should be
418 justified and valid in the sense that it accurately reflects the infectious status of the animal (see also
419 section 6.4.4).

420 The inclusion criteria should be selected to ensure that the study population reflects the intended
421 target population in the best possible way. Any deviation should be justified in consideration of possible
422 differences in clinical outcome between the two populations.

423 For products which according to official guidance should be reserved for certain situations only (i.e. for
424 cases of treatment failure or expected failure of other substances, due to resistance or to less
425 favourable activity characteristics) the inclusion criteria have to be considered with particular care. An
426 appropriate study population for such products could for example be animals from herds with known
427 history of resistance among isolates of the target pathogens towards substances that would normally
428 have been the first treatment choice. Occurrence of resistance should in this case be confirmed
429 through *in vitro* susceptibility tests of isolates from a relevant proportion of the animals. This study
430 would have to include an effective positive control, implying the bacterium(a) under study is (are) fully
431 susceptible to the chosen control. If full correspondence between the study population and the
432 intended target population is not feasible, sufficient information regarding the efficacy could be
433 obtained accordingly (alternative approaches may also be relevant).

434 The effect of treatment is evaluated in two studies:

435 One includes a study population which does not fully correspond to the target population for treatment
436 (e.g. the animals are not treatment failure cases but the product is used as first treatment option) This
437 study is dimensioned to allow for a statistical confirmation of the results and the clinical relevance of
438 the observed effect needs to be justified. Further to this, the effect of treatment is evaluated in a
439 second study, using a smaller group of animals which fully corresponds to the target population (e.g.
440 animals who have not responded sufficiently to previous treatment). The number of animals to be
441 included in that study depends on the indication, the species, the expected efficacy level and the
442 between-animal variation in treatment response. The objective is to obtain a reasonably reliable
443 estimation of the expected treatment effect in the target population. It is acceptable that the second
444 study is not dimensioned to statistically confirm the outcome of the treatment. The results of these

445 two studies will however need to be in general agreement and any deviation will have to be justified
446 with regard to its potential clinical significance.

447 Further information to support and justify the treatment can be obtained from *in vitro* data that
448 demonstrate sufficient susceptibility of the target pathogens towards the antimicrobial substance under
449 study and common and wide-spread resistance in Europe to other substances that would have been
450 first priority for treatment.

451 In diseases characterized by mixed infections (e.g. metritis), inclusion may be based mainly on clinical
452 signs. However, to support the clinical diagnosis samples should be collected from the animals or a
453 relevant proportion of the included animals to clarify which bacteria are involved in the disease
454 process, and the *in vitro* susceptibility pattern should be tested for the most commonly occurring
455 pathogens.

456 **6.4.4. Exclusion criteria**

457 Animals where the effect assessment could be biased from any previous or concomitant treatment
458 should not be included in the study. Appropriate and justified time intervals between previous
459 treatment and study inclusion should be applied. Any other relevant exclusion criteria, dependent on
460 the infection to be treated, can be established. These criteria will help defining the target population in
461 any future marketing authorisation.

462 **6.4.5. Concomitant diseases**

463 Information on concomitant viral, fungal or parasitic infections should be provided, where appropriate
464 so that the impact on the study results of these potential confounding factors can be evaluated.

465 **6.4.6. Endpoints and timing of efficacy assessment**

466 Response to therapy should be mainly based on clinical response criteria and where relevant on
467 microbiological criteria for the specific disease under study. The time points and methods to assess the
468 effects of treatment in field cases should be explained and justified.

469 The choice of the clinical endpoint is critical and determines the study design. The primary endpoint,
470 should be the parameter capable of providing the most relevant and convincing evidence for effect
471 from a clinical perspective, directly related to the primary objective of the trial.

472 Clinical cure rate following appropriate diagnostic procedures is in most situations the preferred
473 primary endpoint. However, depending on the epidemiology and pathogenesis of the disease,
474 microbiological cure rate may also be highly relevant and sometimes necessary as a primary or co-
475 primary endpoint. Support from relevant secondary endpoints will often be necessary to justify a claim.

476 When efficacy assessment on an individual level is not applicable, such as in claims for chicken and
477 fish, treatment success is to be evaluated on group/herd level through relevant efficacy endpoints such
478 as a change in mortality rate. Post mortem examinations including bacteriological sampling are
479 necessary to explore treatment effect in these situations.

480 Post-treatment follow-up should be performed to assess the risk for relapse after the effects of
481 treatment are expected to have ceased i.e. after sub-therapeutic concentrations have been reached in
482 plasma or target tissue. Clinical failures identified at time of primary effect assessment and at time of
483 post-treatment follow-up should be addressed in detail. High relapse rate may call into question the
484 overall efficacy of the product for treatment (and metaphylaxis, if relevant), if return of clinical signs

485 cannot be attributed to re-infection (see related comment section 6.3). The timing of the follow-up
486 measurement should be considered carefully (see section 6.3). Bacteriological sampling and
487 susceptibility tests from clinical failures and relapses should be performed, if feasible.

488 **6.4.7. Special considerations for metaphylaxis claims**

489 Outbreaks of infections may occur in a herd/unit due to the introduction and quick spread of a certain
490 microbe that causes clinical disease in a large proportion of the stock within a short time span. A
491 similar situation can occur when the introduction of an external factor (e.g. a virus infection) causes
492 clinical disease due to an opportunistic bacterial infection which is harboured within the herd. For
493 highly contagious and/or severe diseases simultaneous treatment of clinically diseased animals and
494 metaphylaxis of clinically healthy animals that are likely to be in the incubation phase due to close
495 contact with diseased animals or exposed to the same external factor may be justified from an
496 epidemiological point of view. The objective would be to control disease spread and/or prevent further
497 development of clinical signs in the group.

498 A metaphylaxis claim is only accepted in conjunction with a treatment claim and never as a separate
499 indication. The need for metaphylaxis should always be discussed and the threshold for the initiation of
500 metaphylactic treatment (e.g. the proportion of clinically diseased animals at a certain time point
501 within a group and the severity of clinical signs) should be justified on epidemiological and clinical
502 grounds. The justification may refer to published literature studies.

503 Some formulations (e.g. products to be mixed into drinking water) allow only a claim for both
504 (treatment and metaphylaxis) as all animals will be treated independent of their individual clinical
505 status, whereas formulations intended for individual treatment, like injectables may be approved either
506 for only treatment or for treatment and metaphylaxis.

507 If the study formulation is to be used for group/flock administration only (such as oral powders for
508 drinking water), standard principles for study design will be applicable (see above) using relevant
509 efficacy endpoints to document treatment success. A metaphylaxis claim will be accepted in addition to
510 a treatment claim, if sufficient efficacy on group level is demonstrated, and if the need for
511 metaphylaxis can be justified for the disease.

512 A metaphylaxis claim can also be approved for formulations intended for individual treatment (e.g.
513 injectables).

514 Literature data that document the disease characteristics, epidemiology and the clinical effects of
515 metaphylaxis may be used to support a metaphylaxis claim in this case. When literature data clearly
516 show that effective metaphylaxis can be obtained in target disease outbreaks by use of a product
517 which is comparable to the product under investigation (in terms of active substance, pharmaceutical
518 form and duration of activity), it would be sufficient to confirm efficacy of treatment in a clinical trial
519 which only includes clinically affected animals (i.e. only the treatment claim would have to be
520 confirmed). No data from in-contact animals would have to be presented in this situation, since it will
521 be assumed that the metaphylactic efficacy, will not be less than the efficacy level in clinically affected
522 animals.

523 When insufficient literature information is available to support a metaphylaxis claim for a product
524 intended for individual treatment of group housed animals, new clinical data should be provided. When
525 designing such clinical studies the following should be specifically considered:

- 526 • The threshold for the initiation of metaphylactic treatment (e.g. the proportion of clinically
527 diseased animals at a certain time point within a group) should be justified and reflected in the
528 inclusion criteria for the clinical trial protocol.
- 529 • The primary endpoint should be the clinical health status of the animals as measured by
530 appropriate parameters.
- 531 • The duration of treatment should be carefully justified, taking into account factors such as
532 duration of shedding of infectious organisms, development of immunity and the need to limit
533 development of antimicrobial resistance.
- 534 • Non antimicrobial supportive treatment should be allowed in the treatment and the placebo group.
- 535 • The follow-up period should be sufficient to conclude on the efficacy for prevention of clinical
536 disease in unaffected but treated animals.
- 537 • The study design and selected herds and houses used where any such studies are performed
538 should assure that management or housing do not add unacceptable bias to the study results.
- 539 • All studies should include a negative control. A rescue protocol should be included in consideration
540 of animal welfare.
- 541 • The effect of metaphylaxis and treatment may be documented in the same study. If so, efficacy
542 must be recorded on individual level and treatment outcome should be presented separately for
543 the two groups (diseased animals and animals with no clinical signs but at risk of developing
544 clinical disease). In case the treatment effect is evaluated through comparison with an authorized
545 product, a negative group needs to be included to evaluate efficacy regarding the metaphylaxis.

546 **6.4.8. Special considerations for preventive claims**

547 Preventive claims refer to individual administration of a VMP to healthy animals to prevent infection.
548 Such claims should only be considered in those situations where the risk for infection is very high and
549 the consequences are severe. Prevention claims are not expected to be common and will be carefully
550 scrutinized to ensure that the intended use complies with prudent use principles. The need for
551 prevention must be fully justified for each target species and indication.

552 To support a preventive claim a study including a negative control is normally needed and animal
553 welfare should be accounted for through the acceptance of adequate supportive treatment in both test
554 and control group and implementation of rescue protocols. Alternative study designs may exceptionally
555 be accepted provided the efficacy of the preventive treatment can be determined with sufficient
556 certainty. The criteria used to assess the outcome of disease and/or infection should be fully described.

557 The timing of the preventive treatment in relation to the expected time of exposure to infectious
558 agents should be justified in consideration of the duration of the effect of the product under study.

559 Study animals should be kept in well managed conditions to ensure that bias is not introduced through
560 poor management.

561 **7. Summary of product characteristics (SPC)**

562 The SPC should be drafted taking into account the guidance in the Notice to Applicants (Volume 6C)
563 and the revised CVMP guideline on the SPC for veterinary medicinal products containing antimicrobial
564 substances (EMA/CVMP/SAGAM/383441/2005). Recommendations presented by CVMP for different
565 classes of antimicrobials should be considered.

566

567 Glossary

- 568 • Metaphylaxis: Treatment of all clinically healthy (but presumably infected) animals kept in the
569 same group as animals with clinical signs of a contagious disease. Metaphylaxis is always
570 combined with the treatment of the diseased individuals and consequently a metaphylaxis claim
571 will only be accepted in conjunction with a treatment claim.
- 572 • Prevention: administration of a VMP to healthy animals to prevent infection if the risk for infection
573 is very high and the consequences severe.
- 574 • Re-infection: The re-occurrence of an infection in an animal which according to any relevant
575 microbiological investigation performed after previous antimicrobial treatment was free from
576 infection. The second period of infection could be due to a different bacterial species or strain. In
577 the context of this guideline it is assumed that the infection occurs in conjunction with clinical
578 signs typical for the disease under study.
- 579 • Relapse: The confirmation of an infection in an animal which after antimicrobial treatment was
580 clinically but not bacteriologically cured from the same infection. In the context of this guideline it
581 is assumed that the infection occurs in conjunction with clinical signs typical for the disease under
582 study.
- 583 • Treatment: A treatment claim refers to the administration of a VMP after the onset of clinical signs
584 of disease and only clinically affected individuals are to be treated.

585 References

- 586 Directive 2001/82/EC
- 587 CVMP guideline for the conduct of efficacy studies for intramammary products for use in cattle
588 (EMA/CVMP/344/99)
- 589 CVMP note for guidance on fixed combination products (EMA/CVMP/83804/05)
- 590 CVMP guideline on statistical principles for veterinary clinical trials (CVMP/EWP/81976/2010)
- 591 CVMP guideline on the SPC for veterinary medicinal products containing antimicrobial substances
592 (EMA/CVMP/SAGAM/383441/2005)
- 593 CVMP strategy on antimicrobials 2011-2015 (EMA/CVMP/287420/2010)
- 594 VICH guideline 27 (GL 27): Guidance on the pre-approval information for registration of new veterinary
595 medicinal products for food producing animals with respect to antimicrobial resistance
- 596 VICH guideline 9 (GL 9): Good Clinical Practice (CVMP/VICH/595/1998)
- 597 Good Laboratory Practice (GLP) (see Council Directive 88/320/EEC as amended)
- 598 CVMP guideline on the conduct of pharmacokinetic studies in target animal species
599 (EMA/CVMP/133/99)