



1 27 May 2016
2 EMA/CVMP/ERA/689041/2015
3 Committee for Medicinal Products for Veterinary Use

4 **Guideline on the plant testing strategy for veterinary**
5 **medicinal products**

6 Draft

Draft agreed by Environmental Risk Assessment Working Party (ERAWP)	March 2016
Adopted by CVMP for release for consultation	19 May 2016
Start of public consultation	27 May 2016
End of consultation (deadline for comments)	30 November 2016

7 Comments should be provided using this [template](#). The completed comments form should be sent to vet-guidelines@ema.europa.eu

8



9 **Guideline on the plant testing strategy for veterinary**
10 **medicinal products**

11 **Table of contents**

12 **1. Introduction 3**
13 1.1. Background 3
14 1.2. General considerations 3
15 **2. Plant testing in Tier A and B 4**
16 2.1. Tier A 4
17 2.2. Tier B 4
18 **3. Higher Tier Assessment 4**
19 3.1. Species Sensitivity Distributions (SSD) 4
20 3.2. Testing of Transformation Products 6
21 3.3. Plant test using manure-mediated exposure 6
22 3.4. Chronic toxicity in higher plants 6
23 **4. Interested parties 7**
24 **5. References 7**
25 **Annex I 8**

26

27 **1. Introduction**

28 **1.1. Background**

29 Plant toxicity tests are used in the terrestrial environmental risk assessment of veterinary medicinal
30 products (VMPs) as described in the VICH guideline on environmental impact assessment for
31 veterinary medicinal products Phase II (CVMP/VICH/790/2003) (CVMP/VICH, 2005).

32 The OECD Test Guideline (TG) 208 for plant testing (OECD, 2006) has been updated since the
33 publication of the VICH Phase II guideline. In the updated OECD 208 guideline, guidance on how many
34 plant species are needed for testing of veterinary pharmaceuticals is no longer provided.

35 If a risk for plants is still identified in Tier B of the environmental risk assessment, there are three
36 options for further risk assessment:

- 37 • A statistical extrapolation technique, the so-called species sensitivity distribution (SSD) (EMA,
38 2011)
- 39 • Testing of metabolites/transformation products as described in the OECD TG 208
- 40 • An extended plant test for substances which form non-extractable residues and/or transformation
41 products in manure.

42 **1.2. General considerations**

43 Guidance on how to perform Tier A and Tier B plant testing, including an explanation of the SSD
44 approach for higher tier assessment has already been provided in the reflection paper on testing
45 strategy and risk assessment for plants (EMA/CVMP/ERA/147844/2011) (EMA, 2011). The current
46 guideline replaces this reflection paper and provides additional options for a higher tier assessment.
47 The SSD approach presented in the reflection paper has now been complemented with two additional
48 options for higher tier testing: testing of metabolites or transformation products, and a plant toxicity
49 test using manure mediated exposure. Chronic plant tests are also considered, but currently not
50 recommended.

51 The extended toxicity test on plants for active substances in VMPs is suitable for those substances
52 which form a high amount of non-extractable residues or transformation products. For example,
53 studies on the determination of the fate of VMPs in manure have shown that some antibiotics with high
54 plant toxicity may form a high amount of non-extractable residues. However, it is not known whether
55 these fractions of non-extractable residues are bioavailable in the manure, since non-extractability also
56 depends on the extraction method. Besides this, the manure matrix consists of a high amount of
57 organic matter and undergoes decomposition after spreading onto soil. During this process, non-
58 extractable residues might be released and become bioavailable again. Consequently, the risk of VMPs
59 that bind strongly to manure is unknown, and adapted ecotoxicological tests may need to be
60 considered for the refinement of the risk assessment following Tier B.

61 In some cases, the active ingredient may be metabolized in the animal or transformed to
62 transformation products in manure or soil. In case these major metabolites or transformation products
63 ($\geq 10\%$ of the applied amount) are identified and available for testing, it may also be an option to test
64 those metabolites/transformation products in a standard plant test according to OECD TG 208.

65 **2. Plant testing in Tier A and B**

66 **2.1. Tier A**

67 Six plant species from six different families should be tested in Tier A. It is highly recommended to use
68 species belonging to six different families of four dicotyledonous and two monocotyledonous species,
69 which represent the types of plants grown on agricultural land which would receive a manure
70 application. This is to better reflect the variety in the plant kingdom. Acceptable plant species for use
71 in the test are presented in annex 2 of the OECD TG 208. The lowest EC₅₀ value for the most sensitive
72 endpoint is used in combination with an assessment factor of 100 to derive the predicted no effect
73 concentration (PNEC). The PNEC is compared to the predicted environmental concentration in soil
74 (PEC_{soil_initial}) (CVMP/VICH 2005). If the PEC/PNEC ratio (risk quotient (RQ)) using the PEC_{soil_initial} is
75 higher than 1, the PEC_{soil_initial} can be refined as explained in the CVMP guideline (EMA, 2008). If the
76 resultant RQ calculated with the PEC_{soil_refined} is below 1, the assessment can stop. If the RQ is ≥ 1 it is
77 necessary to proceed to Tier B.

78 Studies with three plant species that were performed before the reflection paper came into force in
79 2012 (EMA, 2011) could still be accepted at Tier A, provided that the PEC/PNEC is < 0.1 .

80 **2.2. Tier B**

81 From the same plants species tested in Tier A, the lowest NOEC or EC₁₀ value is used in combination
82 with an assessment factor of 10. If the resultant RQ is below 1 the assessment can stop. If the RQ is \geq
83 1, it is necessary to proceed to the higher tier assessment.

84 It should be noted that NOEC values often depend on the experimental design, variation within the
85 treatments and the power of the statistical test. Thus, the design of the test (including number of
86 replicates) should be optimized in order to obtain reliable and statistically significant results.
87 Experience has shown that statistically derived NOEC values obtained from plant studies sometimes
88 are associated with effects significantly above 10%. In such cases it is recommended to use EC₁₀
89 values. It should be noted that EC₁₀ values can only be derived if a reliable dose-response relationship
90 is generated and the EC₁₀ is within the range of the tested concentrations (including the controls).

91 No further refinement options for PEC_{soil} are available in Tier B, therefore the PNEC is compared to the
92 refined PEC_{soil} as determined at the end of Tier A.

93 **3. Higher Tier Assessment**

94 **3.1. Species Sensitivity Distributions (SSD)**

95 The species sensitivity distribution (SSD), a statistical extrapolation technique, can be used to derive a
96 PNEC if in Tier B a potential risk for plants is still identified. Using the SSD method, the concentration
97 at which 95% of the species are theoretically protected (HC₅) can be estimated. More information
98 about the SSD method can be found in Posthuma *et al.* 2001.

99 To better reflect the variety of plant species and to improve the statistical power of the SSD, two
100 additional species – preferably from two additional plant families - should be tested in combination
101 with the six species/families tested in Tier B. Only one data point for each species should be included
102 in the SSD.

103 In other legal frameworks such as the REACH regulation, the HC₅ of the SSD is used as the basis for
104 deriving a PNEC in combination with an additional assessment factor ranging typically between 1 and

105 5. However, no specific and generic criteria for selecting the assessment factor is outlined in any of the
106 legal frameworks, as it should be determined on a case-by-case basis. To move away from case-by-
107 case decisions on the magnitude of this assessment factor, the CVMP recommends using the lower
108 confidence level of the HC₅ (LL HC₅) directly as the PNEC.

109 An improved dataset in the SSD assessment, i.e. increased number of tested species covering the
110 same endpoint (e.g. growth), will result in a narrower difference between the median (HC₅) and the
111 lower confidence level (HC₅ LL) of the HC₅, and consequently in an enhanced confidence in the
112 assessment.

113 All data used in the SSD assessment have to meet the general requirements on quality as applicable
114 already in the lower tier risk assessment of VMPs. The additional tests should be performed and
115 reported according to the OECD TG 208, including a report on the fulfillment of the validity criteria.

116 In order to use the SSD, the following additional criteria have to be fulfilled in addition to the general
117 quality criteria:

- 118 • A minimum of eight plant species from at least six different families have to be tested.
- 119 • A minimum number of three monocotyledonous and five dicotyledonous plant species should be
120 included.
- 121 • When reliable EC₁₀ values are available it is highly recommended using these for the SSD. When
122 this is not the case, it can be acceptable to use a combination of NOEC and EC₁₀ values. Only
123 definitive EC₁₀ or NOEC values (excluding ">" and "<" values) can be used in the SSD calculation
124 to ensure the SSD is statistically correctly fitted. In case no reliable EC₁₀ value or NOEC can be
125 calculated because significant effects are found at the lowest test concentration, these species
126 should then be retested. If no significant effects are observed at the highest test concentration
127 (resulting in a 'higher than'- value), the LL HC₅ can be derived with the remaining NOEC and/or
128 EC₁₀ values, provided the SSD contains a minimum of 6 values, and that at least 8 species have
129 been tested.
- 130 • The NOECs or EC₁₀s should all reflect the same, most sensitive, endpoint. If a plant species has
131 been tested more than once, the geometric mean of the NOEC and/or EC₁₀ values of the same
132 endpoint should be used in the SSD assessment. It is not possible to mix NOECs and EC₁₀ values
133 determined in standard tests with those determined in tests with manure.
- 134 • The HC₅ and LLHC₅ are calculated based on a log-normal distribution. The data should be tested
135 by "Goodness of Fit" methods to confirm the likelihood of the data coming from a normal
136 distribution. The Anderson-Darling test on normal distribution is recommended for datasets with
137 less than 20 numbers. If the Anderson-Darling statistics is above the 5% critical value, normality
138 must be rejected and data cannot be used for the SSD.

139 If it is known that plants are sensitive to the substance under evaluation, the stepwise approach of Tier
140 A and Tier B could be waived, and eight or more plants species could be tested in the first instance and
141 the data used in the SSD method, provided the criteria as mentioned above are met.

142 Different software programmes are available to calculate the HC₅ and HC₅ LL and to assess whether
143 the data follow a normal distribution, e.g. the ETX 2.1 program developed by RIVM (2015) and the
144 SSD Generator developed by EPA CADDIS (2005). The choice of software program is optional.

145 The PNEC determined with the SSD is compared to the PEC_{soil} as refined at the end of Tier A to
146 determine the risk quotient for plants.

147 **3.2. Testing of Transformation Products**

148 If the active ingredient is metabolised in the target animal or transformed in manure to relevant
149 transformation products ($\geq 10\%$), the standard OECD 208 test may also be performed with the
150 relevant metabolites and transformation products. The criteria for Tier A and Tier B tests as described
151 above apply.

152 The results of the OECD TG 208 study feed into the risk assessment, where PEC is calculated for the
153 parent and all metabolites or transformation products $\geq 10\%$. To assess the overall risk of the mixture
154 of parent and metabolites/transformation products, the resulting risk quotients have to be summed up.

155 **3.3. Plant test using manure-mediated exposure**

156 The aim of the extended plant test is to assess the effects of VMPs on terrestrial plants considering a
157 more realistic exposure scenario by applying pig or cattle manure spiked with the substance of concern
158 into the soil, by doing an extended OECD TG 208. All requirements and recommendations of the OECD
159 TG 208 still apply to this extended approach. As in Tier A, six plant species from six different families
160 should be tested.

161 Veterinary medicinal products administered to the target animal orally or by injection enter the
162 environment via manure. The modified exposure scenario used in this approach takes into account the
163 degradation of the parent compound into transformation products and/or formation of non-extractable
164 residues. More information on non-extractable residues is available in the CVMP reflection paper on
165 poorly extractable and/or non-radiolabelled substances (EMA/CVMP/ERA/689041/2015) (EMA, 2016).
166 For manure, it is assumed that chemicals are potentially released when manure is mixed into soil or
167 undergoes decomposition.

168 In this extended OECD TG 208, manure is spiked with the test substance and incubated under
169 anaerobic conditions. The scenario of spiking manure is intended to simulate the fate and behaviour of
170 VMPs in manure which is usually stored in tanks before spreading onto agricultural soil. The relevant
171 type of manure should be used for this test; e.g., cattle manure should be used if the product is
172 intended for use in cattle and pig manure should be used if the product is intended for use in pigs. The
173 test design has been successfully verified with pig and cattle manure (Simon et al. 2015). The
174 technique for manure storage and acclimation generally follow the EMA guideline on determining the
175 fate of VMPs in manure (EMA/CVMP/ERA/430327/2009) (EMA, 2011).

176 To determine the PNEC of the extended plant test, the same assessment factors apply as in Tier A or
177 B. The PNEC is compared to the $PEC_{\text{soil refined}}$ determined at the end of Tier A. It is not possible to
178 further refine the PEC for degradation in manure because this process is already taken into account in
179 the determination of the PNEC.

180 The details of the test design, evaluation and reporting are given in Annex I.

181 **3.4. Chronic toxicity in higher plants**

182 The International Organisation for Standardisation (ISO) has developed a chronic toxicity test for
183 higher plants ISO 22030:2005 (ISO, 2005) mainly for the testing of contaminated soils. In this test,
184 not only emergence and growth, but also reproduction parameters such as number of flowers or seed
185 pods are measured. The European Food Safety Authority (EFSA) evaluated the study (EFSA, 2014) NS
186 concluded that its usefulness for testing herbicide effects on non-target terrestrial plants is very
187 limited, as only two crop species with a very short life cycle are recommended for the ISO tests, and
188 the artificial soil recommended for the ISO tests is a very poor soil in which plants do not grow well (10

189 % sphagnum peat, 20 % kaolin clay, 69 % sand). Furthermore, experience has shown that the test
190 may be difficult to perform and the variability in the measured reproductive endpoints is often very
191 high. Therefore, the test is currently not recommended for higher tier testing of VMPs.

192 4. Interested parties

193 Pharmaceutical industry, EU national competent authorities, consultants, contract laboratories.

194 5. References

195 CVMP/VICH, 2005. Guideline on Environmental impact assessment for veterinary medicinal products -
196 Phase II, VICH GL 38 (CVMP/VICH/790/03-FINAL).

197 CVMP, 2008. Revised guideline on Environmental Impact Assessment for Veterinary Medicinal Products
198 in support of the VICH guidelines GL6 and GL38 (EMA/CVMP/ERA/418282/2005-Rev.1).

199 Environmental Protection Agency (EPA), 2005. The SSD Generator/EPA CADDIS. Available online at: http://www.epa.gov/caddis/da_software_ssdmacro.html
200

201 European Food Safety Authority (EFSA), 2014. Scientific Opinion addressing the state of the science on
202 risk assessment of plant protection products for non-target terrestrial plants. EFSA Journal 12(7):3800

203 European Medicines Agency (EMA), 2011. Guideline on determining the fate of veterinary medicinal
204 products in manure (EMA/CVMP/ERA/430327/2009).

205 European Medicines Agency (EMA), 2012. Reflection paper on testing strategy and risk assessment for
206 plants (EMA/CVMP/ERA/147844/2011).

207 European Medicines Agency (EMA), 2016. Reflection paper on poorly extractable residues and/or non-
208 radiolabelled substances. (EMA/CVMP/ERA/689041/2015).

209 ISO, 2005. Soil quality – Biological methods – chronic toxicity in higher plants (ISO
210 22030:2005). <https://www.iso.org/obp/ui/#iso:std:iso:22030:ed-1:v1:en>

211 OECD, 2006. Test No. 208: Terrestrial Plant Test: Seedling Emergence and Seedling Growth Test,
212 OECD Guidelines for the Testing of Chemicals, Section 2, OECD Publishing, Paris. [http://www.oecd-
213 ilibrary.org/environment/test-no-208-terrestrial-plant-test-seedling-emergence-and-seedling-growth-
214 test_9789264070066-en](http://www.oecd-ilibrary.org/environment/test-no-208-terrestrial-plant-test-seedling-emergence-and-seedling-growth-test_9789264070066-en)

215 Posthuma L., Suter II G.W, Trass T.P. (Eds.), 2001. Species Sensitivity Distributions in Ecotoxicology.
216 2001. Lewis Publishers, Boca Raton, 616 pp.

217 RIVM, 2015. ETX 2.1 programme, available online at:
218 http://www.rivm.nl/Documenten_en_publicaties/Professioneel_Praktisch/Software/ETX_2_1.

219 Simon M. Herrchen M. , Nadin Graf N., Förster B., Römbke J., 2015. Concept development for an
220 extended plant test in the environmental risk assessment of veterinary medicinal products. German
221 Federal Environment Agency (UBA). UBA Texte 15/2015. ISSN 1862-
222 4804. [https://www.umweltbundesamt.de/sites/default/files/medien/378/publikationen/texte_15_2015
223 _ebert_tierarzneimittel.pdf](https://www.umweltbundesamt.de/sites/default/files/medien/378/publikationen/texte_15_2015_ebert_tierarzneimittel.pdf)

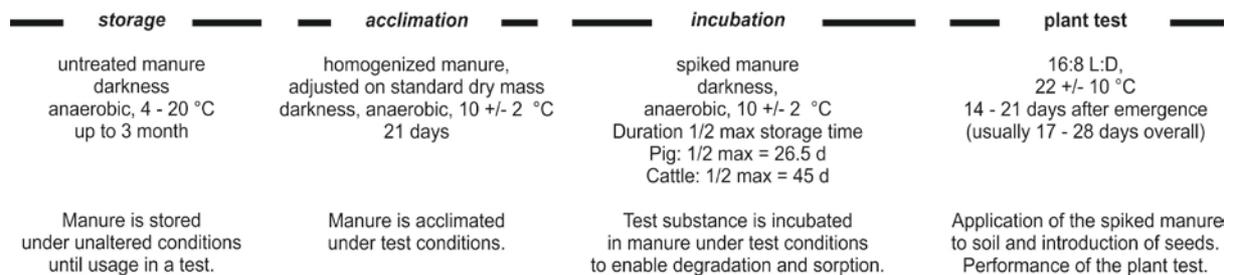
224

225 Annex I

226 Standard operating procedure on test design, performance, evaluation and reporting for the 227 extended plant test

228 I.1 Definitions

- 229 • Manure in this guideline means liquid manure from a tank (mixture of urine and faeces).
- 230 • Manure storage or pre-storage tank is the basin where the manure is stored at the farm.
- 231 • Storage means storing of manure after sampling under unaltered conditions (unprocessed, at
232 anaerobic conditions, 4 – 20 °C, in the dark), comparable with those of storage or pre-storage
233 tanks at farms until use.
- 234 • Acclimation means storing of manure after homogenisation and adjustment to standardised dry
235 matter content, at conditions to acclimate microorganisms before incubation. An acclimation period
236 of 21 days is recommended to ensure comparable conditions between experiments (Hennecke et
237 al. 2015).
- 238 • Half-maximum storage duration is the half of the mean maximum storage time of manure in
239 storage tanks at farms according to table 6 of EMEA/CVMP/ERA/418282/2005-Rev.1 (EMA, 2008).
- 240 • Incubation means storage of manure after acclimation and application of the test substance, at
241 conditions mimicking abundance of manure in storage tanks at farms under standardised conditions.



243 **Figure 1. Schedule and definition of main phases in the extended test design**

244 I.2 Manure

- 245 • The manure applied should originate from animals reared under well controlled conditions. The use
246 of manure contaminated with VMPS, biocides and other material that might impair plant growth or
247 survival should be avoided. The type of animal feed, the feeding regime and the veterinary history
248 of the animals from which the manure will be collected should be recorded and reported.
- 249 • The manure used should reflect the target animals for the intended use of the product. E.g., pig
250 manure when the product is intended to be used for pigs and cattle manure if it is intended to be
251 used for cattle.
- 252 • Manure should be sampled from manure storage or pre-storage tanks which are above or below
253 ground. Prior to collection the liquid manure should be thoroughly mixed in the respective manure
254 tank. Pig manure should be stirred immediately before sampling as separation into liquid and solid
255 phase easily occurs. Duration of mixing depends on the kind of storage tank. However, it should be
256 ensured that the sample of liquid manure is a representative mixture of liquid and solid phase. The
257 sampling site, procedure and the type and size of manure tank (above/below ground,
258 covered/open) should be recorded.

- 259 • Prior to further processing, the manure should be stored preferably at acclimation and incubation
260 temperature for a maximum of three months (EMA, 2011) and under anaerobic conditions.
- 261 • For acclimation, the dry matter content of the manure has to be adjusted to standardised values.
262 The recommended dry matter content in pig manure is 5% ± 1%, in cattle manure 10% ± 1%
263 (EMA, 2011; Weinfurtner, 2010). Manure should be processed using a mixer (e.g. a food processor
264 or similar apparatus) in order to obtain a homogenised phase and to reduce the variability of the
265 test result. All operations should be carried out under anaerobic conditions; exposure to oxygen
266 has to be kept to an absolute minimum if it cannot be avoided. The period of anaerobic acclimation
267 should be 21 days at 10 ± 2 °C in the dark.
- 268 • Key parameters of the manure as mentioned in the EMA guideline on determining the fate of VMPs
269 in manure (EMA, 2011) and listed in table 1 should be measured and reported.

270 **Table 1: Schedule for manure key parameter measurements**

Parameter	stage of test procedure	
	Start of storage	during acclimation
pH	X	X
Microbial activity		X
Organic carbon content [C _{org} mg/kg]		X
Total nitrogen content [N _{total} ; mg/kg]		X
Ammonium content [NH ₄ -N; mg/kg]		X
Phosphate content [mg/kg]		X
Copper content (for pig manure only) [mg/kg] optional	X	
Redox potential [mV]	X	X*
Dry matter content [%]		X*
Temperature [°C]	X	X

271 * Should be measured at the start and end of acclimation.

- 272 • Anaerobic conditions in manure should be ensured and demonstrated by measuring and reporting
273 the redox potential at the end of the acclimation and incubation period where -100 mV should
274 never be exceeded. Typical redox potentials measured in pig and cattle manure have been found
275 to range from -230 mV to -400 mV (Weinfurtner, 2010).

276 I. 3 Application of the test substance

- 277 • Untreated manure by itself can also impair seedling emergence (Simon *et al.* 2015). Therefore, it
278 is advised to check in a pre-test without test substance whether the intended manure
279 concentration in soil has adverse effects on the test plants.
- 280 • Based on nitrogen content, the maximum amount of manure must not exceed 227 mg N_{total} /kg
281 dry soil (170 kg N_{total} /ha per year assuming an incorporation depth of 5 cm and a soil density of
282 1.5 g/cm³). An amount of 20 g fresh manure per kg dry soil, corresponding to approximately 45 –
283 55 kg N_{total} /ha, was shown to be a suitable amount regarding seedling tolerance (Simon *et al.*
284 2015).
- 285 • The quantity of test substance required to obtain the theoretical test concentrations in soil,
286 assuming no transformation during incubation, is mixed with a portion of manure (dry mass

287 content of the manure: 5 ± 1 % for pig manure, 10 ± 1 % for cattle manure) e.g. in glass
288 beakers. Example: If 20 g fresh manure should be applied to 1 kg dry soil and a theoretical test
289 concentration in soil assuming no transformation during incubation should be 100 mg/kg, 100 mg
290 test substance have to be applied to 20 g fresh manure.

291 • Water-soluble substances or those suspended in water can be added directly to the manure and
292 mixed e.g. with a pipette tip. The volume of water added should be the same for each test
293 concentration and should not result in a difference to the desired dry mass content of the manure.
294 The water additionally provided by the stock solution has to be taken into account when adjusting
295 the manure for acclimation (i.e. the manure should thus be adjusted to an appropriate higher dry
296 mass content for acclimation).

297 • Substances of poor solubility in water should be dissolved in a suitable volatile solvent and mixed
298 either directly with the manure or via quartz sand. For direct application, the solvent concentration
299 should not be greater than 0.1 ml/l manure and should be the same concentration in all test
300 vessels. The solvent should be removed from the manure e.g. by using low-pressure followed by
301 pressure compensation using oxygen free air or nitrogen. If the test substance is applied in a
302 solvent, a respective solvent control has to be included. For direct application this should be a
303 solvent control containing manure and solvent, for application via spiked quartz sand (as little as
304 possible), a solvent control containing manure and evaporated spiked quartz sand. The quartz
305 sand added is not considered for dry mass content of the manure. However, every effort should be
306 made to keep the solvent concentration to a minimum.

307 • Solid, insoluble test substances can be applied either directly to manure or via quartz sand. For the
308 latter, the test substance and finely ground industrial quartz sand (as little as possible) is mixed in
309 a suitable mixing device. Hereafter, the mixture is added to the manure and mixed thoroughly. The
310 quartz sand added is not considered for dry mass content of the manure.

311 • It should be kept in mind that all spiking and mixing operations should be carried out in a way that
312 the manure has minimal contact with oxygen.

313 • To reflect representative influences of storage on manure, the spiked manure is incubated under
314 anaerobic conditions in the dark for a period representing the half-maximum storage duration of
315 the respective manure type (26.5 days for pig manure, 45 days for cattle manure) (EMA, 2011).
316 To reflect a realistic case scenario, incubation temperature should be 10 ± 2 °C.

317 • It is recommended to mix the spiked manure with soil in a two-step approach to ensure a
318 homogenous distribution. The spiked manure is added to a sub-portion of test soil and mixed
319 thoroughly. Subsequently, the pre-mixture is added to the rest of test soil and mixed thoroughly.

320 **I. 4 Verification of test substance concentration**

321 • The concentrations/rates of application into the fresh manure must be confirmed by an appropriate
322 chemical analysis, comparable to the requirements of the standard OECD TG 208.

323 • It is strongly recommended to measure the test substance concentration in the incubated manure
324 prior to the start of the plant test at the time of incorporation of manure into soil. As a minimum,
325 samples of manure with the highest concentration and one lower concentration should be
326 considered for analysis. These determinations of test substance concentration provide information
327 about the degradation/adsorption of the test substance in the manure. Depending on the question
328 to be addressed, determination of transformation products and non-extractable residues might be
329 required. As mentioned in the reflection paper on poorly extractable substances (EMA, 2016), the
330 best available extraction technique should be used. This means that determination of the
331 extractable fraction may have to be pursued by various extraction methods with increasing

332 strength. The evaluation of the feasibility of various extraction techniques should be reported in
333 the final study report.

334 **I.5 Plant Test**

335 • In general, the extended test approach follows the standard test in accordance with the OECD TG
336 208 (OECD, 2006) with all respective requirements and recommendations. Additionally six plant
337 species from six different families should be tested as well as in Tier A. However, any modifications
338 are listed below.

339 • The planting of seeds has to be done for all replicates on the same day when the test
340 substance/manure mixture is incorporated into soil to prevent aerobic transformation of the test
341 substance before contact with the seeds.

342 • Control groups with non-spiked manure only are established to assure that effects observed are
343 associated with or attributed only to the test substance exposure. The manure controls or
344 solvent/manure controls are used for evaluation of the effects caused by the test substance. The
345 number of replicates and seeds depend on the chosen test design.

346 • A standard control without manure has to be established to detect possible adverse effects on
347 seedling emergence or growth caused by manure by comparing with the non-spiked manure
348 control. The standard control should consist of at least four replicates with 20 seeds at least in
349 total, independent from the chosen test design. The standard control should not be used for test
350 substance effect evaluation.

351 • The start of the 14 – 21 day growth period is defined by 50% emergence in the manure control
352 and not in the standard control.

353 • Endpoints: The purpose of this approach is to achieve NOEC and/or EC_x values.

354 • For establishment of the number and spacing of concentrations, the following should be
355 considered:

356 – Prior knowledge of the toxicity of the test substance to plants, e.g. derived in a standard test
357 according to OECD TG 208, could help selecting appropriate test concentrations. However, it is
358 strongly recommended to perform a range finding test following the extended test design as
359 the magnitude of effects caused by the test substance together with manure is often not
360 predictable.

361 – A combined approach allowing for the determination of both NOEC and EC_x is highly
362 recommended. Eight treatment concentrations in a geometric series should be used with four
363 replicates each, together with eight manure control replicates. The concentrations should be
364 spaced by a factor not exceeding 2.5.

365 – For determination of the NOEC, at least five concentrations in a geometric series should be
366 tested. Eight replicates for each test concentration plus eight manure control replicates are
367 recommended. The concentrations should be spaced by a factor not exceeding three.

368 • Effect concentrations should be related to soil dry mass and calculated on basis of either the
369 measured concentrations in the applied stock solution (in case of water soluble substances) or the
370 applied weights (in case of insoluble test substances).

371

372

373 **I.6 Validity of the test**

- 374 • All requirements as stated in the OECD 208 TG apply to this extended approach as well. Especially
375 the performance criteria in terms of seedling emergence rate (70%) and post-emergence survival
376 rate (90%) have to be fulfilled in all controls.

377 **I.7 Test Report**

- 378 • All requirements as stated in the OECD 208 TG (test substance, test species, test conditions,
379 results) apply as well to this extended approach. However, additional issues regarding the manure
380 and its preparation, acclimation, incubation and application are listed below and should be
381 reported, too.
- 382 • Type of manure (pig or cattle)
- 383 • Name and location of the farm the manure originates from
- 384 • Feed type, feeding regime and the veterinary history of the animals from which the manure
385 originates (if data are available)
- 386 • Type of manure tank from which the manure originates (e.g. above/below ground, open/covered,
387 size) (if data are available). Sampling procedure; how was the manure mixed before sampling?
- 388 • Key parameters of the manure at the respective time: temperature, pH, redox potential, dry
389 matter content, Corg, N, P, etc.
- 390 • Techniques and conditions (duration, temperature) for manure storage, preparation, acclimation,
391 and incubation (e.g. cooling and/or incubation chamber, mixing device for manure
392 homogenisation).
- 393 • Details on preparation of the spiked manure and verification of the test concentrations.

394 **I.8 References**

- 395 CVMP, 2008. Revised guideline on Environmental Impact Assessment for Veterinary Medicinal Products
396 in support of the VICH guidelines GL6 and GL38 (EMA/CVMP/ERA/418282/2005-Rev.1)
- 397 European Medicines Agency (EMA), 2011. Guideline on determining the fate of veterinary medicinal
398 products in manure (EMA/CVMP/ERA/430327/2009).
- 399 European Medicines Agency (EMA), 2016. Reflection paper on poorly extractable residues and/or non-
400 radiolabelled substances (EMA/CVMP/ERA/689041/2015)
- 401 Hennecke, D., Atorf, C., Bickert, C., Herrchen, M., Hommen, U., Klein, M., Weinfurtner, K., Heusner,
402 E., Knacker, T., Junker, T., Römbke, J., Merrittig-Bruns, U. (2015). Development of a test protocol to
403 study the transformation of veterinary pharmaceuticals and biocides in liquid manure. German Federal
404 Environment Agency (UBA). UBA Texte 78/2015. ISSN: 1862-
405 4804 [https://www.umweltbundesamt.de/sites/default/files/medien/378/publikationen/texte_78_2015_](https://www.umweltbundesamt.de/sites/default/files/medien/378/publikationen/texte_78_2015_development_of_a_test_protocol_to_study_the_transformation.pdf)
406 [development_of_a_test_protocol_to_study_the_transformation.pdf](https://www.umweltbundesamt.de/sites/default/files/medien/378/publikationen/texte_78_2015_development_of_a_test_protocol_to_study_the_transformation.pdf)
- 407 OECD, 2006. Test No. 208: Terrestrial Plant Test: Seedling Emergence and Seedling Growth Test,
408 OECD Guidelines for the Testing of Chemicals, Section 2, OECD Publishing, Paris. [http://www.oecd-](http://www.oecd-ilibrary.org/environment/test-no-208-terrestrial-plant-test-seedling-emergence-and-seedling-growth-test_9789264070066-en)
409 [ilibrary.org/environment/test-no-208-terrestrial-plant-test-seedling-emergence-and-seedling-growth-](http://www.oecd-ilibrary.org/environment/test-no-208-terrestrial-plant-test-seedling-emergence-and-seedling-growth-test_9789264070066-en)
410 [test_9789264070066-en](http://www.oecd-ilibrary.org/environment/test-no-208-terrestrial-plant-test-seedling-emergence-and-seedling-growth-test_9789264070066-en)
- 411 Simon M. Herrchen M. , Nadin Graf N., Förster B., Römbke J., 2015. Concept development for an
412 extended plant test in the environmental risk assessment of veterinary medicinal products. German
413 Federal Environment Agency (UBA). UBA Text 15/2015. ISSN 1862-

- 414 4804. https://www.umweltbundesamt.de/sites/default/files/medien/378/publikationen/texte_15_2015
415 [_ebert_tierarzneimittel.pdf](https://www.umweltbundesamt.de/sites/default/files/medien/378/publikationen/texte_15_2015_ebert_tierarzneimittel.pdf)
- 416 Weinfurtner K. 2010. Matrix parameters and storage conditions of manure. German Environment
417 Agency (UBA). UBA Text 02/2011. ISSN: 1862-
418 4804 <http://www.umweltbundesamt.de/sites/default/files/medien/461/publikationen/4054.pdf>