



1 13 December 2012
2 EMEA/CHMP/BMWP/32775/2005_Rev. 1
3 Committee for Medicinal products for Human (CHMP)

4 **Guideline on non-clinical and clinical development of**
5 **similar biological medicinal products containing**
6 **recombinant human insulin and insulin analogues**

7
8 Draft 1

Draft agreed by Biosimilar Medicinal Products Working Party (BMWP)	March 2005
Adopted by CHMP for release for consultation	May 2005
End of consultation (deadline for comments)	October 2005
Draft agreed by BMWP	January 2006
Adopted by CHMP	22 February 2006
Draft revision agreed by BMWP	November 2012
Adopted by CHMP for release for consultation	13 December 2012
Start of public consultation	14 December 2012
End of consultation (deadline for comments)	30 June 2013

9
10 This guideline replaces 'Guidance on similar medicinal products containing recombinant human soluble
11 insulin' (EMEA/CHMP/BMWP/32775/2005).

Comments should be provided using this [template](#). The completed comments form should be sent to BMWP.secretariat@ema.europa.eu

12
13



KEYWORDS

recombinant human insulin, insulin analogues, similar biological medicinal products, biosimilar, comparability, non-clinical studies, clinical studies, insulin clamp, hyperinsulinaemic euglycaemic clamp, hyperinsulinaemic isoglycaemic clamp

14

15

16 Guideline on non-clinical and clinical development of
17 similar biological medicinal products containing
18 recombinant human insulin and insulin analogues
19

20 **Table of contents**

21	Executive summary	4
22	1. Introduction	4
23	2. Scope.....	4
24	3. Legal basis and relevant guidelines	5
25	4. Non-clinical studies	5
26	5. Clinical studies	6
27	6. Pharmacovigilance plan.....	10
28	7. Extrapolation of indication	10
29		

30 **Executive summary**

31 This guideline lays down the non-clinical and clinical requirements for recombinant insulin containing
32 products, including human insulin and insulin analogues, claiming to be similar to another one already
33 marketed.

34 The non-clinical section addresses the pharmaco-toxicological assessment. The clinical section
35 addresses the requirements for pharmacokinetic, pharmacodynamic and safety studies as well as the
36 risk management plan.

37 **1. Introduction**

38 The Marketing Authorisation (MA) application dossier of a new recombinant insulin claimed to be
39 similar to a reference medicinal product already authorised shall provide the demonstration of
40 comparability of the product applied for to this reference medicinal product.

41 Human insulin is a non-glycosylated, disulphide-bonded heterodimer of 51 aminoacids. Insulin
42 analogues differ from human insulin by the substitution of aminoacids or other chemical changes such
43 as addition of a fatty acid chain within the molecule. Insulin preparations differ mainly by their
44 kinetic/pharmacodynamic profiles. They are usually classified as rapid-, short-, intermediate-, and
45 long-acting preparations, and are used alone or as free mixtures or premixed preparations of
46 rapid/short-acting insulin and intermediate/long-acting insulin in various proportions.

47 There is extensive experience with the production of insulin for therapeutic use from animal sources, in
48 the form of semisynthetic insulin, and through different recombinant techniques. Physico-chemical and
49 biological methods are available to characterise the primary, secondary and tertiary structures of the
50 recombinant insulin molecule, as well as its receptor affinity and biological activity *in vitro* and *in vivo*.
51 Current quality guidelines on comparability provide information on the characterisation and analysis of
52 similar biological medicinal product and its comparator. For recombinant insulins, attention should be
53 given to product related substances/impurities and process related impurities, and in particular to
54 desamido forms and other forms that may derive from the expression vector or arise from the
55 conversion steps removing the C-peptide and regenerating the three-dimensional structure.

56 Currently available insulins are administered subcutaneously or intravenously. The effects of insulin are
57 mediated predominantly via stimulation of the insulin receptor but insulin is also a weak natural ligand
58 of the insulin-like growth factor-1 (IGF-1) receptor. The same receptors are known to be involved in
59 the mechanism of action relevant for the currently approved therapeutic indications of insulins.

60 Antibodies to insulin occur frequently, mainly as cross-reacting antibodies. These have been rarely
61 described to have major consequences for efficacy or safety. The potential for development of
62 product/impurity-specific antibodies needs to be evaluated. Possible patient-related risk factors of
63 immune response are unknown.

64 **2. Scope**

65 The guideline on similar biological medicinal products containing biotechnology-derived proteins as
66 active substance: non-clinical and clinical issues (EMA/CHMP/BMWP/42832/2005) lays down the
67 general requirements for demonstration of the similar nature of two biological products in terms of
68 safety and efficacy.

69 This product-class specific guideline presents the current view of the CHMP on the non-clinical and
70 clinical requirements for demonstration of comparability of two recombinant insulin-containing
71 medicinal products. This guideline should be read in conjunction with the requirements laid down in the
72 EU Pharmaceutical legislation and with relevant CHMP guidelines (see section 3 Legal Basis and
73 relevant guidelines).

74 **3. Legal basis and relevant guidelines**

- 75 • Directive 2001/83/EC, as amended, in particular in Directive 2001/83/EC Art 10(4) and Part II of
76 the Annex I of Directive 2001/83/EC, as amended.
- 77 • Guideline on similar biological medicinal products (CHMP/437/04)
- 78 • Guideline on similar biological medicinal products containing biotechnology-derived proteins as
79 active substance: non-clinical and clinical issues (EMA/CHMP/BMWP/42832/2005).
- 80 • Guideline on similar biological medicinal products containing biotechnology-derived proteins as
81 active substance: Quality issues (EMA/CHMP/BWP/49348/2005) and
82 EMA/CHMP/BWP/247713/2012)
- 83 • ICH guideline S 6 (R1) Preclinical safety evaluation of biotechnology-derived pharmaceuticals
84 (EMA/CHMP/ICH/731268/1998)
- 85 • Guideline on the clinical investigation of the pharmacokinetics of therapeutic proteins
86 (EMA/CHMP/ 89249/2004)
- 87 • Guideline on the investigation of bioequivalence (CPMP/EWP/QWP/1401/98)
- 88 • Guideline on Immunogenicity Assessment of Biotechnology-derived Therapeutic Proteins
89 (EMA/CHMP/BMWP/14327/2006)
- 90 • Guideline on good pharmacovigilance practices (EMA/500020/2012)
- 91 • Guideline on good pharmacovigilance practices, Module V – Risk management systems
92 (EMA/838713/2011)

93 **4. Non-clinical studies**

94 Before initiating clinical development, non-clinical studies should be performed. These studies should
95 be comparative in nature and should be designed to detect differences in the response to the similar
96 biological medicinal product and the reference medicinal product and should not just assess the
97 response *per se*. The approach taken will need to be fully justified in the non-clinical overview.

98 **Pharmacodynamic studies**

99 *In vitro* studies

100 In order to assess any differences in properties between the similar biological medicinal product and
101 the reference medicinal product, comparative studies such as *in vitro* bioassays for affinity, insulin- and
102 IGF-1-receptor binding assays, as well as tests for intrinsic activity should be performed. Partly, such
103 data may already be available from bioassays that were used to measure potency in the evaluation of
104 physico-chemical characteristics. It is important that assays used for comparability testing are
105 demonstrated to have appropriate sensitivity to detect minute differences and that experiments are

106 based on a sufficient number of dilutions per curve to characterise the whole concentration-response
107 relationship.

108 *In vivo* studies

109 Comparative study(ies) of pharmacodynamic effects would not be anticipated to be sensitive enough to
110 detect differences not identified by *in vitro* assays, and are normally not required as part of the
111 comparability exercise.

112 **Toxicological studies**

113 Generally, separate repeated dose toxicity studies are not required. In specific cases, e.g. when novel
114 or less well studied excipients are introduced, the need for additional toxicology studies should be
115 considered

116 Studies regarding safety pharmacology and reproduction toxicology are not required for non-clinical
117 testing of a biosimilar containing insulin or insulin analogues. Studies on local tolerance are not
118 required unless excipients are introduced for which there is no or little experience with the intended
119 route of administration. If other *in vivo* studies are performed, local tolerance may be evaluated as
120 part of these studies.

121 **5. Clinical studies**

122 **Pharmacology studies**

123 Demonstration of similar pharmacokinetic and pharmacodynamic profiles is considered the mainstay of
124 proof of similar efficacy of the biosimilar and the reference insulin. For this purpose, cross-over,
125 preferably double-blind insulin clamp studies using single subcutaneous doses of the test and reference
126 agents and performed at an interval of a few days to a few weeks are considered suitable. The time-
127 concentration and time-action profiles may be studied separately or, preferably, simultaneously (in the
128 same clamp study). Separate pharmacology studies for intravenous use, if applicable, are not required.

129 Study population

130 The study population should be homogenous and insulin-sensitive to best detect potential product-
131 related differences and may consist of normal-weight healthy volunteers or patients with type 1
132 diabetes.

133 Besides their better availability, healthy volunteers have the advantage of relatively consistent fasting
134 blood glucose levels but the disadvantage of presence of endogenous insulin which cannot be
135 distinguished from exogenously administered insulin by the available assays, except for some insulin
136 analogues. Methods for suppressing endogenous insulin or adjusting measured insulin serum
137 concentrations for estimated endogenous insulin should be considered (see below).

138 Patients with type 1 diabetes recruited into clamp studies should have their serum C-peptide
139 concentration measured to ensure absence of relevant remaining endogenous insulin secretion. It is
140 important to establish stable and comparable baseline blood glucose and insulin levels for some time
141 (e.g. one hour) prior to the study intervention in order to achieve comparable baseline conditions in all
142 experiments, which is usually more difficult in patients with type 1 diabetes compared to healthy
143 subjects.

144 Insulin sensitivity in women may vary during the menstrual cycle and it is unclear whether this may
145 affect study results. Thus, inclusion of only men in the studies would be justified.

146 Insulin clamp studies

147 There is general agreement that the euglycaemic or isoglycaemic hyperinsulinaemic clamp technique is
148 the best available method for the measurement of insulin action. In these clamp experiments, the
149 plasma insulin concentration is raised (e.g. by subcutaneous injection of insulin) and the blood-glucose
150 level maintained ("clamped") at a pre-defined level by means of a variable infusion of glucose.
151 Measurements of plasma insulin concentrations and glucose infusion rate (GIR) allow an estimation of
152 the time-concentration and time-action profile and, if investigated in the same clamp study, of the
153 dose-response relationship of an insulin preparation. For the purpose of comparing the
154 pharmacokinetic and pharmacodynamic profiles of a biosimilar and its reference insulin, these clamp
155 experiments will need to be conducted by experienced investigators under highly standardised
156 conditions.

157 Different clamp methods and feedback algorithms for maintaining blood glucose levels exist. Clamp
158 studies can be performed manually or using an automated procedure, e.g. the Biostator. With a
159 Biostator the blood glucose concentration is measured continuously (every minute), and the glucose
160 infusion rates are calculated in a computerised manner by means of a negative feedback algorithm.
161 The major disadvantage of the Biostator appears to be its age (successor models are under
162 development) and difficulties to maintain the system. Manual clamps, on the other hand, are
163 associated with higher blood loss when blood glucose measurements are performed with standard
164 laboratory methods (typical measurement intervals 5 to 10 min) and have a considerable demand for
165 manpower. Manual clamps are also more prone to bias by the examiner compared to automatic
166 clamps. A double-blind design is therefore strongly recommended or, if this is not possible, other
167 means to effectively reduce potential investigator-related bias. Both techniques require substantial
168 experience. However, both methods have been reported to provide similar and reproducible results as
169 long as there are no rapid changes in glucose requirements, which may not be recognised in time
170 depending on the length of intervals between the blood glucose measurements during the manual
171 clamp.

172 Test conditions for a comparative clamp study need to be strictly standardised. Study subjects should
173 undergo the clamp experiments after an overnight fast (usually 10 to 12 hours, only water allowed)
174 and remain fasting throughout the tests to avoid a confounding effect on study results. In patients with
175 diabetes, carry-over effects from the participants' last pre-study insulin injection should be prevented
176 and intravenous insulin infusion started at least 4-6 hours prior to study insulin administration to attain
177 steady-state baseline glucose levels. Ideally, the clamp glucose target should be reached at least one
178 hour before study insulin administration without any glucose infusion during this last hour.
179 Standardisation of clamp technique and factors influencing insulin sensitivity such as time of day,
180 physical activity and food intake/diet, avoidance of alcohol, caffeinated drinks, smoking or medication
181 other than the study medication and absence of intercurrent illness/infection or mental stress are
182 important. Standardisation of habits may be relevant up to several days preceding the day of
183 examination. In the test facility, the subjects should be allowed to adapt to the experimental situation
184 (e.g. for 2 hours prior to the test) to establish a comparable metabolic situation and should stay in bed
185 throughout the experiment in a quiet and pleasant environment. This highlights that even small details
186 are very important. There is, however, evidence that, despite such standardisation, the first of the two
187 clamps may be associated with a somewhat decreased insulin sensitivity, possibly due to an
188 unavoidable increase in the test-related stress level of study subjects with the first clamp.

189 When healthy volunteers are used for the clamp studies, their endogenous insulin production can be
190 suppressed, although usually not entirely, by a priming dose of rapid- or short-acting insulin, followed
191 by a basal rate (e.g., 0.10 to 0.15 mU/min/kg). Alternatively, somatostatin has been used for maximal
192 suppression of endogenous insulin, glucagon and growth hormone during the test period but it should

193 be noted that somatostatin reduces insulin clearance by about 20%, thus prolonging the duration of
194 insulin action artificially. Setting the target glucose level below the patient's fasting glucose also helps
195 suppress endogenous insulin production. Serum C-peptide should be measured in parallel to insulin
196 concentrations to estimate the extent and consistency of suppression of endogenous insulin throughout
197 the experiment. In the absence of insulin suppression, C-peptide correction methods have also been
198 proposed but their value is unclear. Regardless which method is used, it should be justified and
199 consistent throughout the clamp studies to ensure comparable test conditions.

200 The subcutaneously administered dose of the test and reference insulin should reflect commonly used
201 therapeutic doses. For rapid-/short-acting insulins doses of 0.2 to 0.3 U/kg bodyweight and for
202 intermediate-/long-acting insulins doses of 0.3 to 0.4 U/ kg bodyweight are frequently used. The mid-
203 physiological range of hyperinsulinaemia (60-70 μ U/ml), which represents the typical insulin
204 concentration after a standard meal, has been shown to correspond to the steepest part of the dose-
205 response curve of insulin and can thus be expected to be most sensitive to detect potential differences
206 in the time-action profiles of two insulins. All injections should preferably be performed by the same
207 experienced investigator in order to ensure a reproducible subcutaneous injection. The site of injection,
208 known to potentially influence the rate of absorption of insulin, should also be the same to decrease
209 variability.

210 In healthy subjects the blood glucose concentrations are usually clamped 5 mg/dL below the subjects
211 fasting glucose or at 80-100 mg/dL (4.4-5.6 mmol/L). In patients with type 1 diabetes blood glucose
212 concentrations may also be clamped in the euglycaemic range or at typical/target fasting blood glucose
213 levels (isoglycaemic clamp), which may exceed the normal range for healthy subjects. Glucose levels
214 below approximately 60 mg/dL should be avoided because they result in the stimulation of
215 counterregulatory hormones (epinephrine, glucagon, cortisol, growth hormone) to increase blood
216 glucose concentrations and lead to a rapid and pronounced worsening of insulin sensitivity, thus
217 influencing the estimated time-action profile of the investigated insulin preparation.

218 The duration of the clamp studies needs to take into account the known duration of action of the
219 investigated insulin preparation and its dose-dependency. The duration of action in glucose clamp
220 studies may be defined as the time from insulin injection to GIR returning to baseline or, in patients
221 with diabetes, of blood glucose values exceeding a predefined threshold, e.g. 150 mg/dL (8.3 mmol/l).
222 Clamp durations of 8 to 10 hours for rapid- and short-acting insulins and of 24 hours and more for
223 long-acting insulins have been reported for healthy volunteers or patients with type 1 diabetes when
224 using therapeutic doses. A rationale for the selection of the clamp duration should be provided in any
225 case.

226 Endpoints/statistical analyses

227 Pharmacokinetics

228 Comprehensive comparative data should be provided on the time-concentration profiles of the
229 biosimilar and the reference insulin with AUC and C_{max} as the primary and T_{max} , early and late $T_{50\%}$,
230 and $T_{1/2}$ as secondary pharmacokinetic endpoints. Alternatively to early and late $T_{50\%}$, other measures
231 (e.g. $AUC_{0-T_{max}}$) may be used, as appropriate. For the primary endpoints AUC and C_{max} , the 90%
232 confidence interval of the ratio test/reference should lie within 80% to 125%, the conventional
233 acceptance range for bioequivalence, unless otherwise justified. For the other parameters descriptive
234 statistics would be appropriate.

235 Pharmacodynamics

236 The glucose-infusion rate (GIR) over time describes the time-action profile of an insulin preparation.
237 GIR_{AUC} and GIR_{max} should be measured as primary and T_{GIRmax} , and early and late $T_{GIR50\%}$ as secondary

238 pharmacodynamic endpoints. Alternatively to early and late $T_{GIR50\%}$, other measures (e.g. $GIR-AUC_{0-T_{max}}$)
239 T_{max}) may be used as appropriate. Calculation of 95% confidence intervals will be required for PD
240 parameters. Equivalence margins should be pre-defined and justified.

241 It is not easy to control the blood glucose concentrations during the clamp study. Depending on the
242 measurement intervals and feedback algorithm, and due to the inherent measurement delay between
243 sampling and resetting the glucose infusion and the subsequent delay of change in blood glucose levels
244 in response to GIR changes, blood glucose values usually do not correspond to the exact target value
245 but vary around it. In response to that, variations (“noise”) in GIR occur. The Applicant should provide
246 an estimate of the quality of the performance of the clamp study, e.g. by calculating the coefficient of
247 variation of the blood glucose concentrations. The mean intra-individual coefficient of variation of well
248 executed euglycaemic hyperinsulinaemic clamps should usually not exceed 10% for glucose infusion
249 rate. The noise of the GIR measurements can be reduced by fitting a mathematical model. The
250 algorithm for GIR adjustment should be predefined and the appropriateness of the applied smoothing
251 method demonstrated.

252 Specifics of long-acting insulin preparations

253 Long-acting insulin preparations are intended to produce a time-concentration profile which, as far as
254 possible, approximates physiological basal insulin secretion. For long-acting insulins with a very flat
255 pharmacokinetic profile, determination of C_{max} and T_{max} (for insulin and GIR) may be difficult to assess
256 and may even become meaningless. For long-acting insulins with a slow decline in insulin action,
257 together with the unavoidable variations of the GIR, it may be difficult to determine the duration of
258 action, particularly in healthy subjects with interfering endogenous insulin. Therefore, patients with
259 type 1 diabetes are more suitable to determine the time-action profile of long-acting insulins. Insulin
260 sensitivity may increase over time in long-term clamp studies, which may affect GIR. However, when
261 strict standardisation of the test conditions (as described above) is implemented, a similar increase in
262 insulin sensitivity over time in the same individual would be expected in both treatment phases of the
263 cross-over study and should thus not impair the comparison of the biosimilar with the reference
264 insulin.

265 Despite these limitations and the increased intra-subject variability of long-acting compared to short-
266 acting insulins, the hyperinsulinaemic euglycaemic clamp has been successfully used for the
267 comparison of the pharmacokinetic and pharmacodynamic profiles of currently approved long-acting
268 insulin preparations. It should be noted that clamp studies for long-acting insulins may need to be of
269 substantial duration (e.g. for insulin glargine in the clinically relevant dosage range, the duration of
270 action is close to 24 hours in patients with type 1 diabetes).

271 Taken together, hyperinsulinaemic euglycaemic/isoglycaemic insulin clamps, with some limitations,
272 may be appropriate to compare the time-concentration and time-action profiles of long-acting
273 biosimilar and reference insulins but will usually require a relatively large sample size and a long
274 duration for the purpose of demonstrating similarity

275 **Clinical efficacy**

276 There is no anticipated need for specific efficacy studies since endpoints used in such studies, usually
277 HbA1c, are not considered sensitive enough for the purpose of showing biosimilarity of two insulins.

278 **Clinical safety**

279 Convincing demonstration of similar physicochemical characteristics, pharmacokinetic and
280 pharmacodynamic profiles of the biosimilar and the reference insulin will already provide reasonable
281 reassurance that adverse drug reactions which are related to exaggerated pharmacological effects

282 (e.g. hypoglycaemia) can be expected at similar frequencies. Therefore, the main focus of the safety
283 study is the evaluation of immunogenicity, although similarity in the adverse event profile, e.g. with
284 regard to hypoglycaemia and local tolerability, of the biosimilar and the reference product should also
285 be confirmed.

286 Immunogenicity studies should always include a reasonable number of patients with type 1 diabetes. If
287 a mixed population is included, stratification for type of diabetes and pre-existing anti-insulin
288 antibodies is necessary. The study duration should be at least 12 months, including a comparative
289 phase of at least 6 months. The primary outcome measure should be the incidence and titres of
290 antibodies to the test and reference medicinal products but there is no need to power the study to
291 formally demonstrate non-inferiority regarding immunogenicity. The potential impact of antibodies, if
292 detected, on glycaemic control, insulin requirements and safety, especially local and systemic
293 hypersensitivity reactions, should be investigated, and the necessity for further characterisation, e.g.
294 with regard to their neutralising potential, considered.

295 **6. Pharmacovigilance plan**

296 Within the authorisation procedure the applicant should present a risk management plan in accordance
297 with current EU legislation and pharmacovigilance guidelines. The RMP of the biosimilar should take
298 into account identified and potential risks associated with the use of the reference product and, if
299 applicable, safety in indications licensed for the reference product that are claimed based on
300 extrapolation. In addition, it should be discussed in detail how these safety concerns will be addressed
301 in post-marketing follow-up.

302 **7. Extrapolation of indication**

303 Demonstration of similar pharmacokinetic and pharmacodynamic profiles of the biosimilar and the
304 reference product and absence of safety issues such as excessive immunogenicity with subcutaneous
305 use will allow extrapolation of efficacy and safety data to intravenous use, if applicable, and to other
306 indications and patient populations licensed for the reference product.