INDUSTRY PERSPECTIVE ON SYNTHETIC OLIGONUCLEOTIDES

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European Pharma Oligonucleotide Consortium (EPOC)

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Agenda

- The European Pharma Oligonucleotide Consortium
- Published papers
- Drug substance starting materials
- Drug substance impurities
- Terminal sterilization
- Future papers



Mission Statement

Through innovative collaboration, identify areas for harmonization of oligonucleotide* CMC development and propose solutions to influence the external regulatory environment; expediting patient access to revolutionary medicines

* Currently defined as antisense, siRNA, aptamers and sgRNA; includes small molecule conjugates. Currently excludes mRNA and antibody/protein conjugates

Publications

EPOC

- 1. Technical Considerations for Use of Oligonucleotide Solution API | Nucleic Acid Therapeutics (liebertpub.com)
- 2. Perspectives on the Designation of Oligonucleotide Starting Materials | Nucleic Acid Therapeutics (liebertpub.com)
- 3. <u>Strategies for Identity Testing of Therapeutic Oligonucleotide Drug Substances and Drug Products | Nucleic Acid</u> <u>Therapeutics (liebertpub.com)</u>
- 4. <u>Determination of Purge Factors for Use in Oligonucleotide Control Strategies | Organic Process Research &</u> <u>Development (acs.org)</u>
- 5. Solution Oligonucleotide APIs: Regulatory Considerations | SpringerLink
- 6. Terminal sterilization; in the press
- 7. Nitrosamine Risk Assessments in Oligonucleotides | Organic Process Research & Development (acs.org)

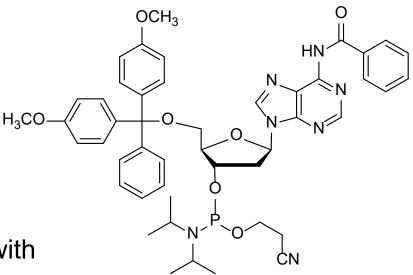
Non-EPOC

- 1. <u>Quality Aspects of Oligonucleotide Drug Development: Specifications for Active Pharmaceutical Ingredients |</u> <u>SpringerLink</u>
- 2. Impurities in Oligonucleotide Drug Substances and Drug Products | Nucleic Acid Therapeutics (liebertpub.com)

Drug Substance Starting Materials

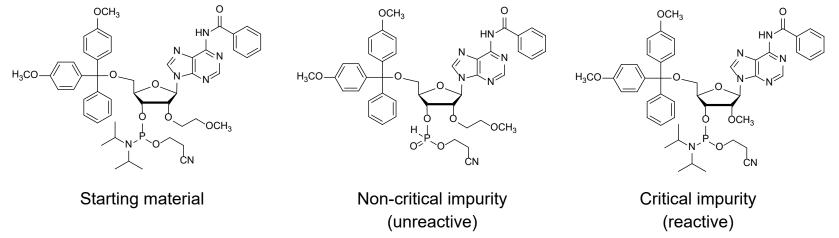
<u>Perspectives on the Designation of Oligonucleotide Starting Materials | Nucleic Acid</u> <u>Therapeutics (liebertpub.com)</u>

- Aim: To create harmonized starting material justification packages (to avoid inconsistencies among different sponsor companies and different regulatory agencies)
- Focus is phosphoramidite starting materials for solid phase synthesis
- Try to apply as much of Q11 as possible
 - Starting materials have defined and stable structures with characteristic chemical and physical properties
 - Emphasize requirement to provide complete proof of structure and characterization, e.g., ¹H, ¹³C, ³¹P NMR (and 2D NMR), HPLC-MS



Drug Substance Starting Materials

- Impurity characterization and fates of impurities
- Critical and non-critical impurities, e.g.,



- Major impurities are unreactive. Reasonable to assume unreactive impurities flow though column and are non-critical
 - In most cases, non-reactivity should be obvious should be no requirement to conduct formal fate and purging studies
- Reactive impurities lead to product related impurities
 - Propinquity arguments very weak for solid phase synthesis
 - Corresponding product related impurities typically possess similar physical properties to parent oligonucleotide limited opportunities to reduce by purification (and sometimes difficult to detect and quantify in drug substance)
 - Potential for multiplicative effects

Drug Substance Starting Materials

Specification

- Should focus on testing critical quality attributes (CQA)
- CQA
 - Identity determines drug substance identity
 - Reactive (nucleoside) phosphoramidite impurities
 - Likely need to be controlled to low levels (due to multiplicity issues)
 - Normally determined by validated HPLC, UHPLC or LC-MS techniques (³¹P NMR typically doesn't add much).
- <u>Non</u> CQA (although may be controlled for economic reasons)
 - Non-reactive nucleotide (nucleoside) impurities subject to extensive purging during solid phase synthesis
 - Solvents subject to extensive purging during solid phase synthesis
 - Water solutions typically dried before use, starting materials used in excess, coupling failure typically results in yield reduction only
 - Stereochemistry at P-atom* racemization during activation, and therefore starting material P-stereochemistry does not impact drug substance stereochemistry
 - Purity (assay) starting materials used in excess, coupling failure typically results in yield reduction only

* Excepting P-stereopure starting materials used to make P-streopure drug substance

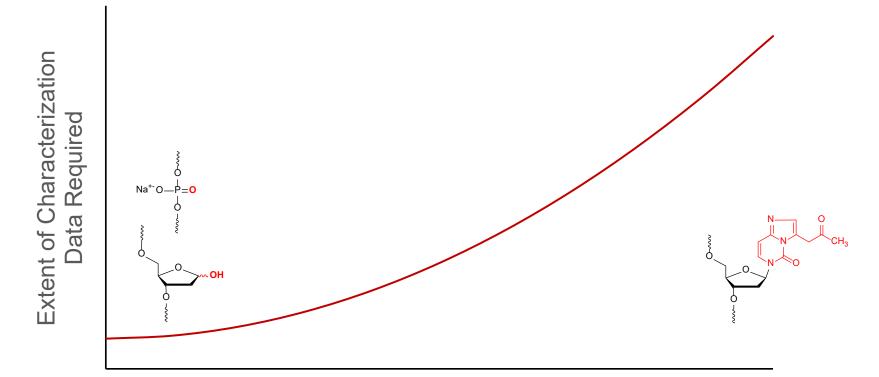
Drug Substance and Drug Product Impurities

Impurities in Oligonucleotide Drug Substances and Drug Products | Nucleic Acid Therapeutics (liebertpub.com)

- Focus is product-related impurities
 - Oligonucleotide impurities derived from impurities in starting materials and reagents
 - Oligonucleotide impurities formed due to side- or incomplete reactions of the manufacturing process
 - Oligonucleotide impurities arising by degradation
- Chemistry considerations impurity characterization and reporting (grouping, reporting threshold, specification limits)
- Safety considerations classes of oligonucleotide impurities, qualification requirements (identification and qualification thresholds)

Impurity Characterization

• Propose a spectrum of empirical data requirements



Impurities previously characterized independently by several groups for several oligonucleotides Impurities characterized for the first time

Impurity Reporting

- Specifications should include tests of <u>specified impurities</u> (frequently observed impurities that are diagnostic of the success or otherwise of a particular step or control strategy in the manufacturing process)
- Example
 - Single-stranded oligonucleotides typically contain low levels of n-1 impurity
 - Levels of n-1 largely dependent on successful completion of the detritylation step of solid-phase oligonucleotide synthesis
 - Therefore, monitoring n-1 in the drug substance may confirm that the detritylation step performed as expected
- Complete separation of impurities from likely not be practical for most (perhaps all?) oligonucleotides - will be necessary to <u>report impurities as groups</u>, or classes
 - Several means of assembling impurities into groups
 - Structural class: e.g., n-1, n+1, abasic oligonucleotides, CNET etc.
 - Benefit is that all members of a particular group are subject to the same mechanism of formation
 - Consistent with the idea that reported results reflect the success of a particular synthetic step or control strategy

Impurity Reporting: Reporting Threshold

- Reporting Threshold: Level above which there is an expectation to report (and sum) impurities
- For most small molecule drugs, <u>reporting threshold = 0.05%</u>
 - Important consideration is the LOQ of the analytical method
 - Impurities of small molecule drugs are typically single components that are readily separated chromatographically as sharp, well-defined peaks detected by UV absorbance
 - LOQ values on the order of 0.02% or less are readily achieved; therefore, 0.05% reporting threshold is technically feasible
- Product-related impurities of oligonucleotides often difficult to separate chromatographically (in some instances can only be detected using mass spectrometry)
 - When chromatographic separation is practical, composite nature of impurities contributes to chromatographic peak broadening
 - Peak broadening reduces signal-to-noise and impacts LOQ
 - Mass spectrometry detection typically has higher LOQ than UV detection
 - However, oligonucleotides typically contain a greater number of impurities than most small molecule drugs if reporting threshold is too high, risk severely under-reporting total impurities

<u>Reasonable compromise reporting threshold = 0.20%</u>

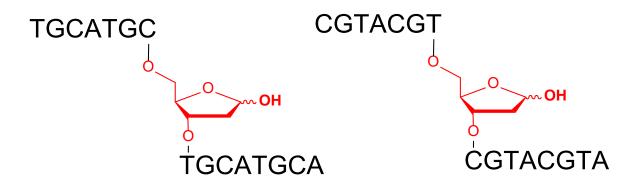
Safety Considerations

Impurity Class	Examples	Safety Assessment?
Class I Impurities that are also <u>major metabolites</u> (structure and sequence are the same as parent)	 Impurities that lack nucleotides from the 3' or 5'- end of the parent oligonucleotide Impurities formed by incomplete conjugation of (parent) conjugated oligonucleotides Parent single-stranded impurity of double- stranded oligonucleotides 	No; impurities that are also major metabolites are generally considered qualified
Class II Impurities containing <u>only structural elements</u> found in naturally occurring nucleic acids	 Phosphate diester impurity of phosphorothioate diester oligonucleotides 	No; endogenous presence of the structural element rules out inherent safety concerns associated with the impurity
Class III Impurities that are <u>sequence variants</u> of the parent (impurity contains only structural elements found in the parent)	• n-1 • n+1	No; safety concerns limited to potential off- target antisense effect or generation of inflammatory motif: therefore, appropriate to consider safety of each <u>individual</u> impurity, which are too low to generate a pharmacologic effect
Class IV Impurities that contain <u>structural elements not</u> <u>found in the parent</u> or in naturally occurring nucleic acids	 e.g., CNET, abasic impurities Unidentified impurities 	Yes; potentially altered reactivity and metabolism, lead to potentially toxic metabolites

Safety Considerations

• Likely that the safety of the impurity is independent of the sequence context

Example: Potentially reactive abasic impurity



Sequence surrounding modified linkage, sugar or base unlikely to impact the physico-chemical properties of the lesion significantly

- Concept aligned with database structural alert approaches e.g., ToxAlerts, DEREK, NOXNET
- Animal studies should not need to be repeated to qualify the same impurity classes in other oligonucleotides of the same or similar chemical structure

Unspecified Impurities and the Identification Threshold

- Drug substance (and product) specification should contain limit on unspecified impurities; standard
 practice to set the limit on unspecified impurities <u>equal to the Identification Threshold</u>
- Driven by <u>Quality</u> expectations
 - Formation of new and unidentified impurities above identification threshold be avoided
 - In a state of control, manufacturing processes, control strategies, and analytical methods should be capable of avoiding new impurities above the identification threshold
 - Set to 0.10% for most small molecule drug substances
- Same Quality expectation should apply to oligonucleotides
- But if Identification threshold is driven largely by what we should be capable of doing (when everything goes to plan)...
 - Oligonucleotides typically produced by solid-phase synthesis, where products of one reaction are fed into the next without purification; little opportunity for impurity removal
 - Minute quantities of reactive impurities in starting materials may lead to significant quantities of drug substance impurities
 - Complex impurity profiles; often difficult to distinguish impurities from the parent and each other
- Places practical limitations on ability to control unspecified impurities

Identification threshold of 1.0% is about the best we can do

Reasons that 1.0% of a new oligonucleotide impurity doesn't pose a safety concern

- Physico-chemical properties likely to be very similar to those of the parent (water soluble polyanion of similar molecular weight and protein binding properties)
- Large difference in MW between oligonucleotide and small molecule impurities. Same quantities of each represent very different molar amounts
- Most oligonucleotides are dosed at much less than 100 mg/day
 - Therefore, 1% of daily dose is usually much less than the 1 mg amount discussed in Graham et al [Calculating qualified non-mutagenic impurity levels: Harmonization of approaches Regulatory Toxicology and Pharmacology 126 (2021) 105023 https://doi.org/10.1016/j.yrtph.2021.105023]
- Because all impurities tested to date are well-tolerated, future impurities likely to be well-tolerated too



Is it Safe?

Terminal Sterilization (TS)

- Objective: Fill gaps between regulatory expectations and application of TS of oligonucleotide drug products, with special attention to the EMA guidance (<u>Sterilization of the medicinal product</u>, <u>active substance</u>, <u>excipient and primary container - Scientific guideline | European Medicines</u> <u>Agency (europa.eu)</u>
- Most prescriptive guideline published
- Provides guidance on the selection of TS methods and technical requirements
- Efforts should be made to enable terminal sterilization
 - Selection of pH, excipients, container closure system, optimization of sterilization method/process
- Change in impurities above qualification thresholds does not preclude the use of TS

TS is preferred - sterile filtration and aseptic processing should be justified

Key Concepts

- General acceptance that proteins and other biologics are too labile to withstand TS
- Oligonucleotide drug products traditionally manufactured by sterile filtration and aseptic processing (like biologics)
- Theoretical risk assessment
 - Double stranded oligonucleotides with T_m values below 110°C, aptamers and oligonucleotides conjugated to proteins likely excludable from the outset; some single stranded oligonucleotides are more thermally stabile (hard to exclude without further work)
- Formulation development
 - Oligonucleotides most stable around pH 8 (low pH \rightarrow depurination; high pH \rightarrow deamination)
 - Phosphate buffer pKa = 7.2; buffering range = 6.2 to 8.2; negligible temperature coefficient; compatible with oligonucleotides
 - More buffering capacity required as [oligonucleotide][†]; 10-25 mM generally sufficient

Key Concepts

- Sterilization cycle development
 - EMA guideline
 - Indicators used for validation have a decimal decay value of \geq 1.5 min at 121°C (D121 \geq 1.5 min)
 - Initial bioburden concentration ≤ 100 CFU/100 mL (up to 100 CFU/filled unit)
 - Therefore, a cycle lethality of F₀ ≥ 12 min is required to achieve an SAL of ≤ 10⁻⁶; target F₀ = 12 min during feasibility studies (and not the minimum F₀ = 8 min referenced)
 - Degradation occurs during temperature ramping (when temperature is < 110°C and there is no additional accumulation of cycle lethality); goal is to mimic the anticipated capability of the commercial equipment
 - Autoclave cycles commonly controlled by setting the hold temperature and time
 - Should include tolerances expected in commercial-scale equipment. Temperature band of 2°C is typical, therefore, use temperature at the top of the band (123°C)
 - No sense examining lower temperatures for longer times
 - Decreasing sterilization temperature by 10°C results in 10-times increase in time (to achieve same lethality)
 - Rate constant for deamination (first-order reaction mechanism, Ea = 25 kcal/mol, decreases by a factor of 2.3 going 121 to 110°C
 - Higher temperatures require much higher level of control of ramping and support pressure (and are likely not feasible in commercial scale equipment)

TS Conclusions

- For aqueous products, assess moist heat sterilization methods per the decision tree; irradiation or chemical methods not recommended
- Feasibility studies should be carried out using the most heat-stable formulation(s) available and container closure of representative materials of construction. Generate enough data to rule confidently on the feasibility of TS
- TS not recommended when any of the following occur:
 - Level of an impurity increases beyond its toxicological qualification limit (consistent with EMA guideline)
 - Significant changes occur to other CQA, e.g., appearance, visible and subvisible particulates, pH
 - Total degradation is sufficient to risk failure of lower assay release limit while factoring analytical and manufacturing variability (and overages are not recommended)
 - Chemical or physical changes to the container negatively impact its suitability or functionality. For example,
 - CCIT is adversely impacted (such that maintenance of sterility over the shelf life is compromised)
 - Unacceptable levels of leachables
 - Glass delamination

Acknowledgements

EPOC Steering Committee members

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