

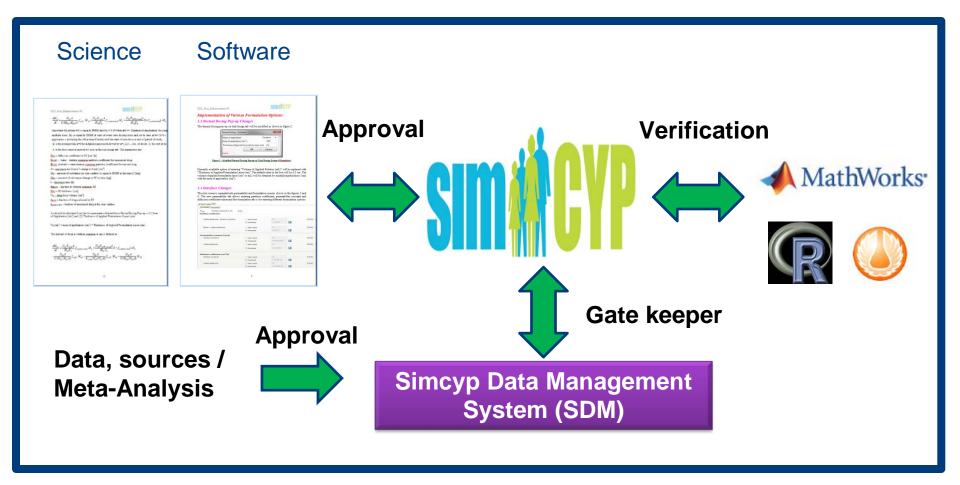
Qualification of the Simcyp platform for the intended purposes

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EMA workshop, London, UK
 21st Nov 2016

How do we qualify the Simcyp Simulator for intended purposes?

The Simcyp platform qualification is a *process* implemented within a quality assurance by design framework.

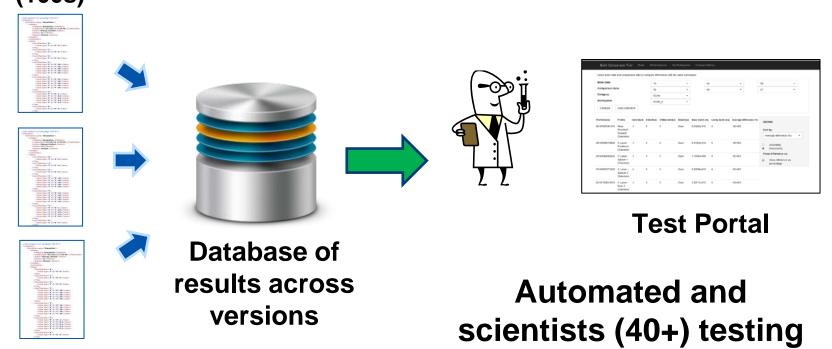




Automated/Scientists (40+) regression/verification analysis

During the course of each version development (up to 100 builds) the Simulator is continuously tested against a subset of workspaces ~50 and periodically tested against a full set of workspaces ~500 for verification and regression against previous versions and identifying deviations.

Multiple Workspaces (100s)



Updated and expanded after each version CERTARA

Transparency and independent scrutiny

Continues publication of algorithms and models including detailed equations, compound and population models, as well as, active interaction with peers and experts expedite qualification.

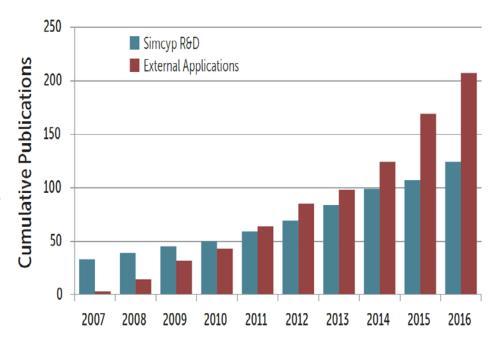
Free academic licenses

Research only: 69

Teaching only: 122

Teaching and research: 1329

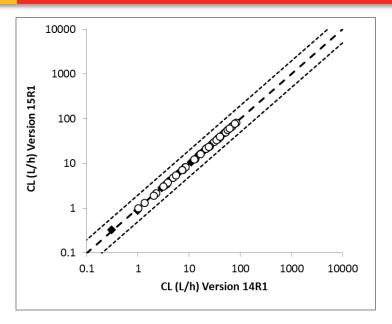
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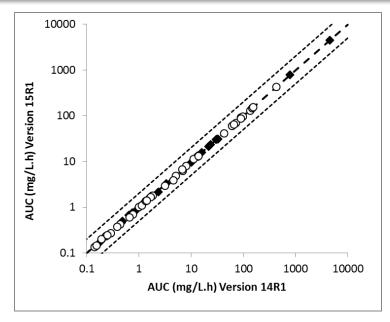


Cumulative peer-reviewed publications



Across version comparisons available to consortium/regulatory scientists





Static parameters

Dynamic parameters

The Version Comparison repository contains version-to-version comparison documentation of the compounds. The aim is to compare the overall compound file performance relative to the previous version.

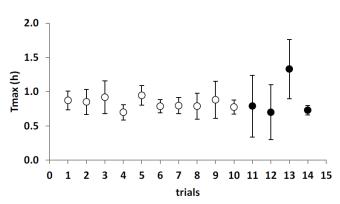
	Download	Size	Updated
1	V13R2_V14R1_VersionComparison.pdf	0.44 MB	23/07/2015
×	V14R1_V15R1_VersionComparison_32bit.pdf	0.46 MB	07/12/2015
3	V14R1_V15R1_VersionComparison_64bit.pdf	0.45 MB	07/12/2015

Qualification of compounds/populations

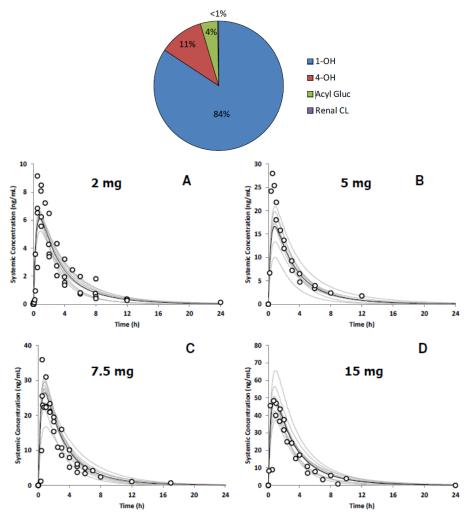
We've been posting summaries on Simcyp members area accessible to consortium/regulatory scientists.

Model Summary

Absorption 1 st order absorption		User inputs for fa and ka	
Distribution Minimal PBPK		User input for V _{ss}	
Elimination	Enzyme kinetics	Recombinant data for CYP3A4 and CYP3A5; renal clearance from clinical studies	



Key PK parameters including t_{max} , C_{max} , V_{ss} , CL, CL_{PO}

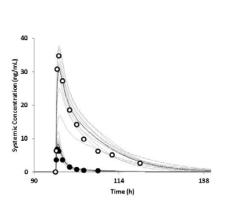


Qualification against a range of



Victim compounds: Verification of fm among other parameters

DDIs of specific CYP inhibitors



	Observed		Predicted	
	C _{max} ratio	AUC ratio	C _{max} ratio	AUC ratio
Fluconazole + Midazolam po	2.30	3 AA	1.92 (1.72 - 2.21)	3.16 (2.65 - 3.75)
Ketoconazole + Midazolam po	4. 4	15.1	3.67 (3.06 – 4.68)	12.74 (10.71 - 14.85)
Ketoconazole + Midazolam iv		4.80		4.56 (3.61 – 5.53)

	Observed		Predicted	
	C _{nax} rato	A LC ratio	C _{max} ratio	AUC ratio
Quinidine + Dextromethorphan	6.10	6.34	3.77 (2.85 – 4.38)	5.02 (3.75 – 5.63)
Quinidine + Dextromethorphan	4.38	7.31	3.90 (3.62 – 4.22)	5.38 (5.04 – 5.83)

	Observed		Predicted	
	C _{max} ratio	AUC ratio	C _{max} ratio	AUC ratio
Fluvoxamine + caffeine	7.4	13.71	1.31 (1.18 – 1.53)	11.10 (7.82 – 14.69)
Ciprofloxacin + caffeine	1.07	1.17	1.08 (1.07 – 1.11)	1.20 (1.13 – 1.27)

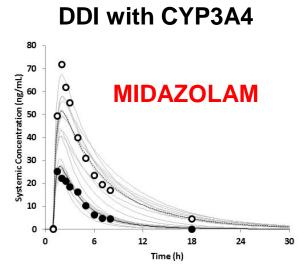
Also, looking into site of metabolism/interaction (liver vs gut) whenever possible e.g. midazolam.

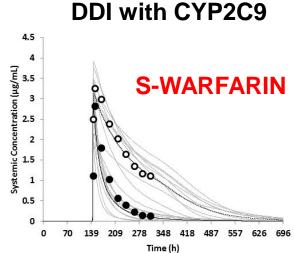


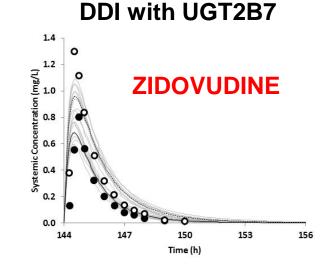
Perpetrator compound qualification

FLUCONAZOLE

Inhibitor of CYP3A4: Inhibitor of CYP2C9: Inhibitor of UGT2B7







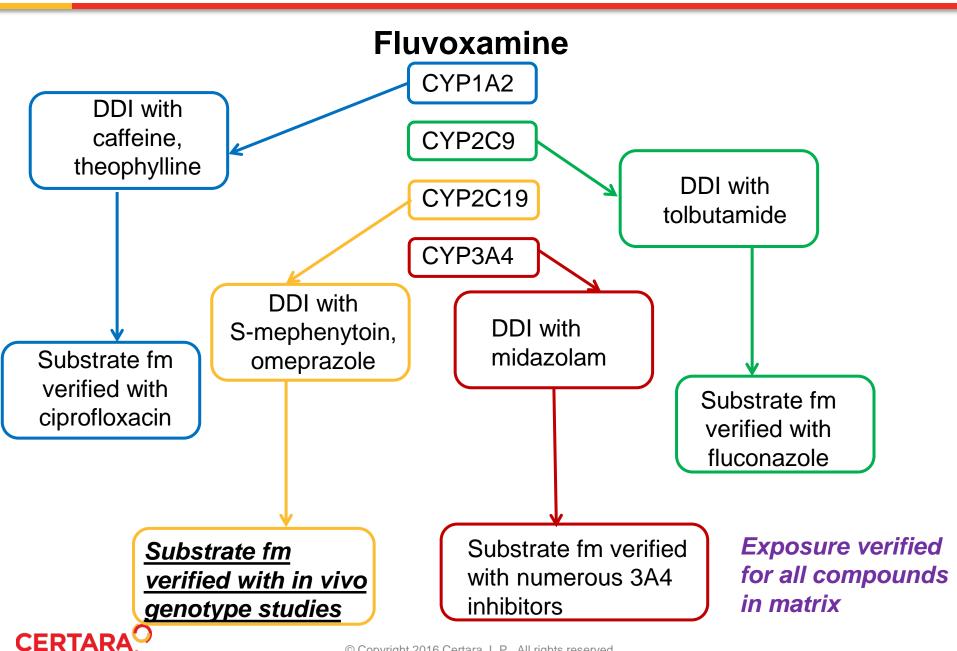
	Ratio			
	C_{max}	AUC		
Parameter				
Mean	1.36	1.70		
Trial 1	1.28	1.62		
Trial 2	1.38	1.75		
TRIAZOLAM				
111a1 S	1.41	1./0		
Trial 6	1.32	1.65		
Trial 7	1.38	1.74		
Trial 8	1.35	1.68		
Trial 9	1.40	1.81		
Trial 10	1.39	1.75		
Observed	1.47	1.63		

2C9 substrate	Fluconazole dose	Observed AUC ratio	Mean Predicted AUC ratio (trial range)			
Tolbutamide	150mg SD 1.90		1.38 (1.34 -1.40)			
Tolbutamide	Tolbutamide TOLBUTAMIDE - 1.68)					
Phenytoin	PHENYTOIN - 1.56)					
Phenytoin	200mg QD	1.88	1.41 (1.33 - 1.52)			



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Matrix approach for perpetrators qualification



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Qualification processes/datasets

The three practical qualification processes seem to be adequate. However, the qualification by "learned societies" may need clarification.

Qualification datasets for site of inhibition can be challenging while alternative solutions, e.g. accurate prediction of AUC may indicate the model reliability.

Line 236:

"Again, the qualification will only be valid for situations covered by the qualification dataset, e.g. only for the specific enzyme(s), *site of inhibition (e.g., liver, intestine)* and the type of background data (including pharmacokinetic data, the system parameters and the population used) on which the simulations were based.



In a constructive way - what changes would you propose?

"Accurate prediction" is mentioned but isn't defined?

Line 417:

"Sensitivity analysis should be performed for *all parameters* that are likely to markedly influence the outcome of the simulated pharmacokinetics and/or the model application."

The use of sensitivity analysis should be targeted/specific to parameters which are uncertain and/or specific assumptions made.

We know, for example, changing tissue blood flows and organ sizes will change the PK profile but is this a useful/necessary exercise?



In a constructive way - what changes would you propose?

Line 399:

"Consideration should be given to whether there are parameters in the model that are *correlated* and if there is uncertainty in the value of *more than one* of the parameters. In the case that an *identifiability issue is suspected* additional in vitro or clinical data may be required to increase certainty in the parameters. A description on how *any identifiability issues* have been handled should be given."

In a mechanistic PBPK model, almost all parameters are (and have to be) correlated. Further, given the structure of population based PBPK models, there are many cases where identifiability can be an issue. Identifiability issues only in certain cases need investigation.



Thank you for your attention!

