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6 Draft qualification opinion

7 In-vitro hollow fiber system model of tuberculosis (HFS-TB)

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Comments should be provided using this [template](#). The completed comments form should be sent to qualification@ema.europa.eu

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Keywords	Tuberculosis, regulatory, qualification, in-vitro, pharmacokinetics, pharmacodynamics, modelling and simulation
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1 Last day of relevant Committee meeting.
2 Date of publication on the EMA public website.
3 Last day of the month concerned.



17 **Background information as submitted by the applicant**

18 *Background of the CPTR initiative*

19 The CPTR initiative is a broad collaboration of pharmaceutical companies, government regulatory and
20 multilateral agencies, academia, civil society advocates and non-government organizations that aim to
21 accelerate the development of new, safe and highly effective TB treatment regimens with shorter
22 therapy durations than the current standard of care. CPTR was formed through the collaboration and
23 support of the Bill & Melinda Gates foundation, the Global Alliance for TB drug development and the
24 Critical Path institute. The CPTR PCS-WG strives to identify, develop consensus around and build the
25 evidence base to support potential new drug development tools (DDTs) for TB medical product
26 development.

27 *The hollow fiber system of TB (HFS-TB)*

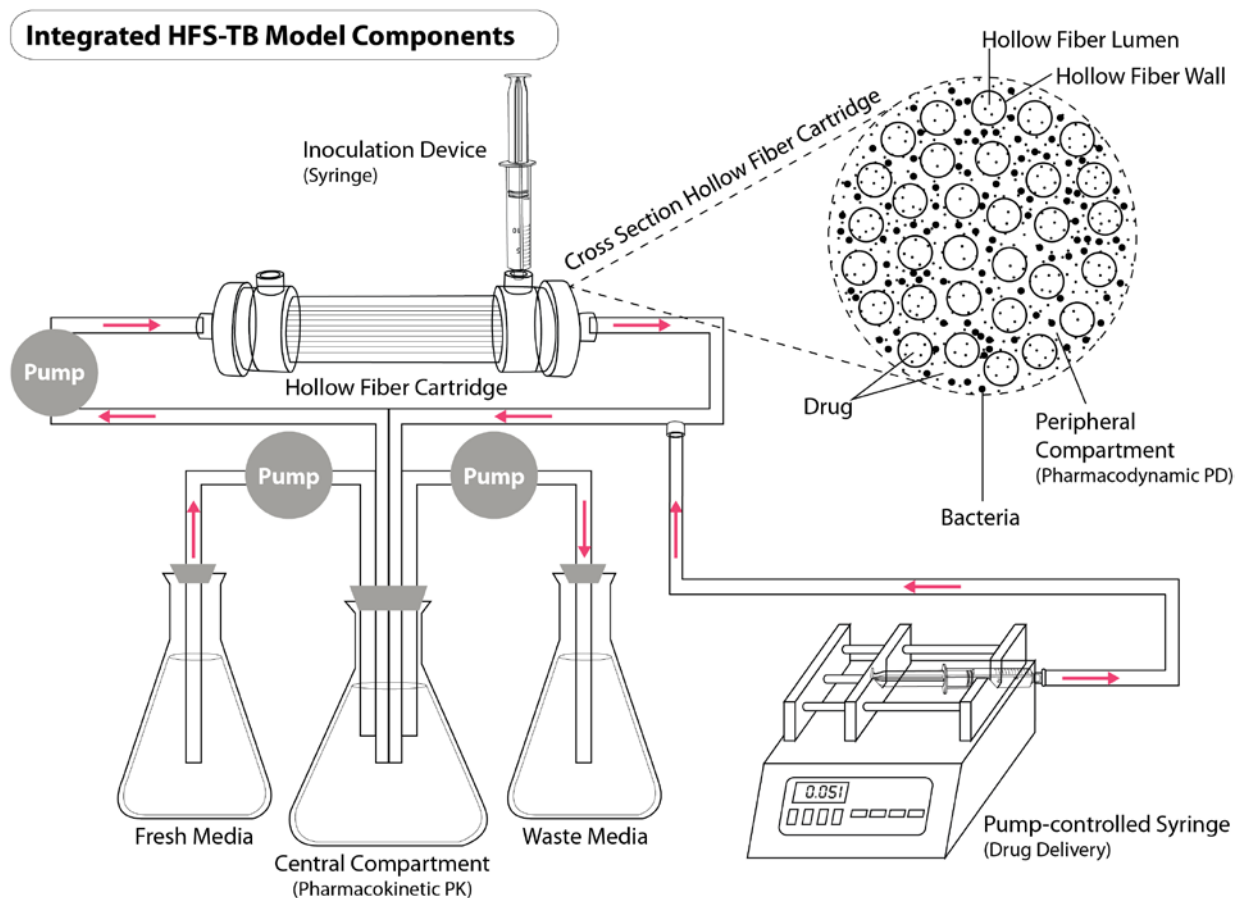
28 Since the time of both Robert Koch, who discovered mycobacterium tuberculosis (Mtb), and his
29 protégé Paul Ehrlich, who laid the foundation for modern chemotherapy and antibiotic drug
30 development, animal models have been used extensively for TB drug development. However, these
31 animal models have sometimes failed to predict clinical efficacy. In addition, the advent of
32 antimicrobial PK/PD science required a tractable, patient-relevant model of pharmacokinetics. This
33 model would allow repetitive sampling of bacteria during drug therapy, as is the standard in clinical
34 trials, and would easily delineate differential effects of drug therapy on the various metabolic sub-
35 populations of Mtb.

36 Approximately 10 years ago, the HFS-TB was developed by Gumbo et al. and first presented at the
37 Interscience conference on antimicrobial agents and chemotherapy (ICAAC). Since that time, the
38 model has been refined to become more sophisticated in its utility and application, which has
39 expanded to include the ability to influence effective dose selection in clinical settings. The applicability
40 of this tool and the utility of data produced by it are described below. The HFS-TB was developed not
41 just to fill the gaps in knowledge associated with existing preclinical efficacy models, but also to offer
42 more quantitative measurements and outputs than those provided by animal models. This is
43 important, as quantitative PK/PD assessment is crucial to the development and execution of an
44 integrated drug development process.

45 The HFS-TB was specifically designed to: (1) mimic the pharmacokinetic concentration-time profiles of
46 antibiotics observed in TB patients in both plasma and at the site of infection, (2) mimic the metabolic
47 and physiologic behavior of Mtb strains encountered in infected patients given that Mtb can exist in one
48 of three metabolic states which impact the efficacy of drugs against the bacterium, (3) quantify the
49 sensitivity and resistance of these Mtb strains to various doses and combinations of antibiotic agents
50 over time and (4) perform PK/PD studies that will inform dose selection in clinical trials. The HFS-TB
51 enables a quantitative understanding of the relationship between dynamic drug concentrations, as well
52 as dynamic populations of drug-susceptible and drug-resistant Mtb, over time. HFS-TB assembly
53 specifications can be found in the Amendment. The HFS-TB (figure 1) consists of a “pharmacodynamic”
54 compartment (also referred to as the peripheral compartment) and a pharmacokinetic (PK)
55 compartment, which consists of a central compartment that allows drug to equilibrate with contents of
56 the peripheral compartment via diffusion process across the hollow fibers. The peripheral
57 (“pharmacodynamic”) compartment also houses either extracellular or intracellular Mtb, which can be
58 maintained for several months.

59

60 Figure 1: diagram of hollow fiber system for TB (HFS-TB)



61
62 Hollow fibers are semi-permeable capillary tubes, whose pore sizes can be varied depending on the
63 type of study being performed. A fiber pore diameter of 42kDa has been commonly utilized in the TB
64 model because it allows easy and rapid equilibration of small molecules across the hollow fibers, while
65 preventing bacilli from distributing between compartments, or entering the hollow fiber lumina. The
66 lumina of the hollow fibers is part of a continuous media flow path that includes tubing and the central
67 reservoir. Drugs are administered via a computer-controlled syringe pump, with drug entering the
68 central compartment via tubing in the flow path. Drug infusion rates are designed to mimic a desired
69 peak concentration and time to peak concentration, as encountered in patients. Fresh media is
70 introduced to the central compartment via tubing in the flow path and used media removed via the
71 central reservoir to create a dilution system that allows the concentration of the drug to decline over
72 time with the same half-life as observed in humans. The system allows the investigator to mimic
73 varied concentration-time profiles and thus drug combination regimens. The actual drug or drug
74 combination levels achieved are directly quantified by direct sampling of the central compartment via a
75 stopcock mechanism in the flow path tubing at pre-specified times for measurement of achieved drug
76 concentrations. The timing of sampling within a given study is determined by the PK profile of the
77 drug(s) of interest. The actual drug concentrations observed are then utilized in mathematical *in silico*
78 models, dose-finding studies and for extrapolation to the equivalent clinical doses in patients.

79 Bacilli (which are too large to cross the pores) are inoculated into the peripheral compartment via a
80 syringe device (figure 1). The inoculated bacteria could be at low pH when semi-dormant bacilli are
81 needed for evaluation, under anaerobic conditions to generate the non-replicating persistor state, at
82 ambient air and normal pH for log-phase growth bacteria, or within macrophages or neutrophils to
83 represent the intracellular state. The growth media circulating in the system is selected to maintain a

84 particular pH, or support human-derived cell lines such as macrophages. The system is incubated at
85 37°C with oxygen and CO₂ content pre-specified according to the metabolic status of Mtb under study.
86 The peripheral compartment is then sampled at pre-specified times, typically day 0, day 3, day 7 and
87 then every seven days thereafter, similar to the sampling schedule typically used for determination of
88 total bacterial burden in sputum in liquid cultures.

89 There are multiple outputs from the HFS-TB, including: (1) total bacterial colony forming unit (CFU)
90 count; (2) drug-resistant Mtb CFU count; (3) drug concentration which can be modeled using
91 compartmental PK analysis methods; (4) macrophage count (in some studies) and number of bacteria
92 per macrophage; (5) RNA expression and (6) whole genome sequencing which can be performed on
93 sampled material. All of these PD measures can be directly correlated with PK measurements taken at
94 the same time point within a study. This is a significant advantage to in vivo model study methods.
95 The last three types of PD measures allow a systems pharmacology- based approach that can be
96 utilized in drug development.

97 *Proposed use of HFS-TB in drug development*

98 The HFS-TB can: (1) mimic the concentration-time profiles of antibiotics observed in TB patients, (2)
99 mimic the metabolic and physiologic behavior of Mtb populations commonly encountered in infected
100 patients with pulmonary TB and the intracellular Mtb characteristic of disseminated TB and (3) quantify
101 the sensitivity and resistance of these Mtb populations to various doses and combinations of antibiotic
102 agents over time. When these outcomes are correctly achieved, the results can then be used in Monte
103 Carlo simulations to identify (i) optimal doses of drugs, (ii) drug combinations which are most likely to
104 achieve desired microbial outcomes, (iii) expected response rates from a drug or combination regimen,
105 (iv) expected rates of and time to resistance emergence in patients and (v) susceptibility breakpoints
106 based on a minimum inhibitory concentration (MIC) above which therapy by a specific drug will fail.

107 The HFS-TB is proposed for use in optimization of drug regimens and dose selection to maximize the
108 bactericidal and sterilizing effect rates and minimize the emergence of resistance. When used early in
109 the drug development cycle as a complementary and additional tool to existing methodologies,
110 information regarding optimal dose selection, dosing schedules and potential combination therapies
111 can be obtained. Additionally, the HFS-TB can be used in a post-approval setting to optimize currently
112 used drug regimens (for both dose and dosing schedule) for drug-susceptible and drug-resistant TB.
113 Therefore, the results obtained by the HFS-TB are expected to support trial design for Phase I, II, III
114 and IV clinical trials.

115 Phase I dose ranging study design can be optimized by data from HFS-TB experiments especially when
116 the PKs in humans can be predicted by data from preclinical studies. The HFS-TB is used to identify
117 optimal PK/PD exposures associated with maximal and fastest bactericidal and sterilizing effect rates
118 and resistance suppression. Monte Carlo simulations would then be used to identify optimal clinical
119 dose. In appropriate circumstances the need for dose ranging study designs, which potentially expose
120 some patients to suboptimal doses, might be avoided. In later-phase development, HFS-TB provides
121 information to help identify the optimal doses and combinations for Phase II and III studies, the
122 proportion of patients expected to have maximal response and the expected rates of acquired drug
123 resistance.

124 *Advantages of the HFS-TB*

125 Animal models quickly became the preferred model in TB drug development following elegant work by
126 Paul Ehrlich using various classes of chemotherapeutic agents at the turn of the last century. In the
127 last 70 years, mice and guinea pigs have been used for anti-TB drug development with considerable
128 success, but also with some notable failures. In the 1950s, the studies of Steenken and Wolinsky in
129 guinea pigs demonstrated no effect of pyrazinamide, which led to its reduced use. Fortunately, the

130 studies of Yeager et al. in patients and Grumbach in mice, demonstrated pyrazinamide efficacy and it
131 is now known to be essential for short course chemotherapy as demonstrated in clinical studies in East
132 Africa by the British medical research council.

133 The utility of these in vivo models for new anti-TB regimens has been questioned, most recently by
134 Mitchison and colleagues (renowned subject matter experts for in vivo model systems of Mtb). The
135 predictive accuracy of these in vivo models for reducing duration of therapy in moxifloxacin containing
136 regimens (as a substitution for isoniazid) has been contested, as the preclinical results did not track
137 with clinical findings for the regimen. A more recent example of inconsistency between animal model
138 data and clinical predictability is a mouse study that demonstrated that daily dosing of rifapentine
139 would lead to cure of TB in three months or less in the standard regimen, which led to the tuberculosis
140 trials consortium study 29 (TBTC 29) of 531 patients. In TBTC 29, daily rifapentine in patients was no
141 better than daily rifampin. Moreover, the animal models have limited use in assessment of acquired
142 drug resistance in combination regimens due to comparatively low bacterial burdens than achieved in
143 humans. Furthermore, the lack of repetitive sampling of Mtb in mice or guinea pigs, which is the
144 standard approach in clinical trials, limits extrapolation from these models for time-to-event outcomes
145 and identification of the exact timing of drug resistance emergence. Finally, a formal study to examine
146 how accurate these models are at quantitative forecasting has not been performed, thus the in vivo
147 models cannot be used as baseline models.

148 The HFS-TB offers distinct advantages to current in vivo model systems for evaluating efficacy,
149 resistance potential and dose determination as described below.

- 150 • Simulation of human PK and PD

151 The primary advantage of the HFS-TB is its capacity to simulate human PK/PD of a drug or drug
152 combination. This is strengthened by the capability for iterative and repetitive sampling for
153 quantitative measurement of both organism and drug concentration simultaneously. This is a distinct
154 feature of this model as repetitive sampling of both drug and organism are not feasible in in vivo
155 models of infection based on limitations of access to infection sites for both organism and drug.
156 Therefore, providing a quantitative understanding of PK/PD relationships is the primary benefit of HFS-
157 TB in the drug development process. This allows measurement of efficacy and resistance suppression
158 providing a rational and efficient approach to explore novel combination therapies, which can then be
159 directly translated, into more effective clinical trial designs.

- 160 • Microbial response to drug(s)

161 The HFS-TB has been used to determine the bactericidal and sterilizing effect (i.e., microbial kill) rates,
162 likelihood of resistance emergence and effects of drug combinations, which are comparable to those
163 effects in the sputum cultures of patients. The HFS-TB model has the advantage that the microbial
164 sub-populations important in sterilizing effect (i.e., non-replicating persisters and semi-dormant bacilli)
165 can be separately studied from log-phase growth sub-populations. This allows for more accurate
166 identification of microbial kill rates, resistance emergence within each sub-population and differential
167 effects of antibiotics, all of importance in design of regimens that would shorten therapy duration.

- 168 • Complete eradication

169 The ability to culture the entire contents (usually 20 mL of culture) of the peripheral compartment of
170 the HFS-TB at the end of an experiment interval allows assessment of the potential for a compound's
171 ability to completely eradicate Mtb at early time points such as one or two months. The data from this
172 model can inform the likely time point in a clinical trial setting that can be considered for proof of
173 efficacy rather than relying on relapse rates (as is current practice) in in vivo models of infection.

- 174 • Assessment of resistance potential

175 As cited above, the HFS-TB has the advantage of accurately mimicking human PK. This is an
176 improvement to static in vitro models, which are typically employed to assess resistance potential for
177 drugs or drug combinations. Drug instability issues are obviated in the dynamic HFS-TB model as well.
178 As an example, rifampicin, a key component of the current standard of care for TB and the new anti-
179 TB drug ertapenem, are unstable in static medium incubator conditions used to assess MIC or
180 minimum bactericidal concentration profiles. These two aspects are important because the shape of
181 the concentration-time curve of some anti-tuberculosis drugs (i.e., the stressor) is an important
182 determinant of microbial effects such as acquired drug resistance. This reflects the evolutionary
183 principle in which oscillations in the intensity of environmental stressors result in higher mutation rates
184 than with constant stressor pressure. Comprehensive understanding of concentration-effect profiles is
185 needed to fully understand resistance potential. For the same reasons, the HFS-TB has advantages
186 over in vivo models, (i.e. rodent models) in which the half-life of many drugs differ greatly from that in
187 humans, thereby exposing Mtb to different shapes of the concentration-time curves, or intensity of the
188 chemical stressor.

- 189 • Repetitive sampling

190 Repetitive sampling from the same system offers another advantage over in vivo models, which rely
191 on terminal procedures to obtain samples to be cultured. Repetitive sampling vastly improves
192 statistical power, time-to-event analysis and repeated event analysis.

- 193 • Assessment of drug combinations

194 Anti-TB drugs exhibit peak and area under the concentration-time curve (AUC) concentration-
195 dependent synergy and antagonism in patients. Two and three drug combinations, at different doses
196 for each drug, can be performed in the HFS-TB allowing identification of dose and concentration-
197 dependent synergy or antagonism.

198 **Summary of the qualification exercise as submitted by the** 199 **applicant**

200 *Methods*

201 The sponsor performed comprehensive literature searches to identify all relevant publications that
202 were used to perform our analyses.

- 203 • Search A identified all HFS-TB studies and Monte Carlo simulation studies published in the
204 literature that utilized the HFS-TB output to make therapeutic predictions. HFS-TB generated
205 data from studies obtained in search A were then compared to clinical data from studies
206 obtained in searches B and C.
- 207 • Search B identified clinical studies that were used to examine therapeutic relevance of the
208 HFS-TB output through descriptive correlations. For each correlation, it was required that these
209 clinical studies were published prior to HFS-TB. Therefore they were not used for predictive
210 accuracy assessment.
- 211 • Search C identified clinical studies that were used to evaluate predictive or forecasting
212 accuracy of the HFS-TB study output. For each predictive evaluation, it was required that the
213 clinical study was published at least six months after the HFS-TB publication.

214 Standard evidence-based medicine criteria were used to evaluate the quality of clinical studies and are
215 described in the methods section (section 5) of the final dossier submitted. Data and information were

216 extracted from the relevant publications to enable several types of analyses. These analyses are
217 described within the results section of the final dossier submitted (section 6).

218 *Results*

- 219 • Search A identified 26 studies that reported the output of HFS-TB or used the HFS-TB output in
220 Monte Carlo simulations.
- 221 • Search B identified 17 clinical studies.
- 222 • Search C identified 20 clinical studies.

223

224 *Analyses conducted based upon search results*

225 *Analysis 1*

226 Mtb kill rates in patient sputum, patterns of microbial kill, cessation of effect and time to emergence of
227 drug resistance from the 17 clinical studies identified in search B were compared to the same
228 parameters in the 26 HFS-TB studies identified in search A. Descriptive correlations demonstrated
229 excellent concordance for these parameters for standard doses of isoniazid, rifampin, pyrazinamide,
230 ethambutol, ciprofloxacin and moxifloxacin.

231 *Analysis 2*

232 Predictive accuracy was examined using 20 clinical studies identified in search C, which were published
233 at least 6 months after HFS-TB studies identified in search A.

234 In analysis 2a, the forecasting accuracy of the pharmacokinetic/pharmacodynamic (PK/PD) indices or
235 dosing schedules associated with optimal microbial kill or resistance suppression was identified. The
236 PK/PD indices associated with microbial kill and resistance suppression by rifampin, isoniazid,
237 ethambutol and pyrazinamide were accurately predicted in the HFS-TB when compared with the clinical
238 studies.

239 In analysis 2b, the data from HFS-TB studies suggested new hypotheses relevant to therapeutic
240 strategies and contradicted some accepted therapeutic strategies. These HFS-TB study data were then
241 compared to clinical studies published at a later date. Six such hypotheses were subsequently
242 confirmed by the results of the clinical studies.

243 In analysis 2c, the quantitative predictive accuracy of several HFS-TB study generated parameters,
244 including optimal drug doses and PK/PD exposure values, was calculated. HFS-TB study results were
245 compared to results generated in clinical studies performed after the HFS-TB results were published.
246 The forecasting accuracy rate was 94.4% (95% confidence interval [CI]: 84.3-99.9). The bias was
247 1.8% (CI: -13.7 to 6.2) and thus crossed zero. Therefore, it is proposed that the HFS-TB model is a
248 drug development tool that is highly accurate for forecasting optimal drug exposures, drug doses,
249 dosing schedules and appropriate drug combinations for anti-TB drugs/drug regimens.

250 *Conclusion*

251 These data demonstrate a validated forecasting accuracy for HFS-TB and support its utility as a
252 valuable complementary, additional tool to existing methods for anti-TB drug dose selection and
253 regimen design. HFS-TB outputs can facilitate drug development strategies and be useful for more
254 informed design of phase I, phase II and phase III clinical studies.

255 **Scientific discussion based on the qualification team**
256 **assessment**

257 Hollow fiber systems are already in use, not only for the in-vitro study of antimycobacterial products,
258 but also for antivirals and for antibiotics acting against different bacterial species. The system is not
259 intended to replace other experiments or clinical trials, and it should not be a mandatory system for
260 implementation. Although expected to give supportive evidence to a marketing authorisation
261 application, the system should not be considered as pivotal to obtaining a marketing authorisation. It
262 will provide however important information with regard to dose selection, mechanism of action etc.
263 Most likely the data from the HFS-TB will be used in a regulatory context to support clinical trial
264 applications and scientific advice discussions where much of justification for the proof of concept and
265 the dosing regimen may well depend on the data obtained with this model.

266 The meta-analyses provided by the consortium combined with the totality of evidence in scientific
267 literature and the cases seen in regulatory submissions support the qualification of the HFS-TB system.

268 Some points of criticism on the qualification exercise that need further follow up from the consortium
269 are provided below:

- 270 • The data presented by the consortium are based on a relatively small number of published
271 studies. Although it remains possible that all studies were not identified by the searches and
272 that in particular non-published materials may not have been accessible, it appears that
273 reasonable efforts were done to avoid bias in the selection of sources. No raw data were
274 presented and all analyses were retrospective. Despite the fact that confirmatory clinical
275 studies should have been published at least 6 months after the predictive HFS-TB study before
276 being used this does not unequivocally reassure that both studies were independent from each
277 other. Also the presented analyses include different PK/PD indices and clinical outcomes. It
278 might be interesting when more data become available to present separate analyses per PK/PD
279 indices - clinical outcomes combinations. Additional prospective data with hypotheses to be
280 tested clearly formulated on beforehand would be of value to strengthen the predictive value of
281 the model. The consortium indicated that two such prospective studies are currently
282 conducted. One study is to establish the use of Moxifloxacin/rifampin/pyrazinamide + isoniazid
283 or ethambutol. The other study will investigate PA824 to document dose scheduling to induce
284 microbial kill and prevent resistance. Both will address log-phase growth, semi-dormant state
285 and intracellular mycobacteria. An evaluation of the hypotheses tested with the raw data being
286 generated would be welcomed. Sometime has elapsed since the last literature searches were
287 performed by the consortium and perhaps a new search could identify new data that could be
288 useful to better determine the value of the system. As it can be anticipated that more data will
289 become available independently from the consortium, it would be advisable to attempt
290 collecting these in order to integrate them in further analyses. The consortium is encouraged
291 to submit new data as indicated above in a follow up qualification opinion.
- 292 • It is considered very important to fully appreciate instances in which HFS-TB can over- or
293 under-estimate anti-Mtb activity or fail to predict the clinical situation and to understand the
294 reasons why this may occur (e.g. on the one hand the HFS-TB operates in an
295 “immunosuppressed” state but on the other hand it may not accurately mimic local factors that
296 could impact on antibacterial activity at the site of infection). Thus, it is necessary to consider
297 whether the HFS-TB could lead to a decision not to develop a promising agent because the
298 HFS-TB has underestimated its activity or could inadvertently over-estimate the activity of a
299 regimen against fully susceptible (DS-TB) or MDR/XDR-Tb. It is unlikely that cases of

300 underestimation will be known because they will likely result in premature termination of
301 development of a new product or a particular treatment regimen will not be pursued in a
302 clinical setting. It would however be problematic if useful treatments would not reach clinical
303 practice if the reason would be a shortcoming of the HFS-TB. Although cases of overestimation
304 are not reported for the time being they may exist and early recognition of such over-
305 estimated activity would be needed for the benefit of the patient in clinical trial. When more
306 data become available a better estimate of false negatives and false positives observed with
307 the system could be given. Therefore it remains important that data on this aspect would be
308 regularly updated also including examples that are not published but known to the consortium.
309 It would also be important to explain failures of the model to predict clinical outcome, because
310 it would allow a better understanding of the appropriate use of the model, and may indicate
311 those conditions and purposes when it should not be used. The consortium indicated that the
312 literature search is being continuously updated. Additional data generated both from within and
313 outside the CPTR consortium will be incorporated, in the spirit of maintaining the continuously-
314 evolving nature of quantitative drug development tools like HFS-TB.

- 315 • An important asset of the system is that it can mimic the pharmacokinetic behaviour of a
316 substance in humans. It is however not clear at the moment how the pharmacokinetics of
317 combinations will be mimicked at the same time for all mono-components, in particular if their
318 PK properties are markedly different. This will however need to be considered on a case by
319 case basis and can probably not be addressed in a general way. Another remaining concern is
320 the way the intracellular exposure to anti-Mtb substances is controlled in order to mimic
321 exposure in patients at the site of the infection. If the HFS-TB model is being used, information
322 should at least be available about the extent at which concentrations at the infection site and
323 in particular at intracellular (granuloma) sites are mimicked on a case by case basis. The
324 consortium has indicated that dynamic determinations of concentration-time profiles inside
325 macrophages are possible and mentioned an example with moxifloxacin. The control of
326 concentration-time curves intracellularly could also be systematically addressed in future work.
- 327 • The HFS-TB is a dynamic system that allows researchers to study relevant Mtb strains under
328 different metabolic conditions. Organisms in log-phase growth, dormant state or non-
329 replicating persistors can be studied separately. The HFS-TB system can also be used to study
330 the intracellular state of Mtb within macrophages or neutrophils. This possibility is considered
331 an important advantage of the system. It should be documented to a sufficient extent that the
332 organisms were indeed in the desired metabolic state during the experiment.
- 333 • Operational characteristics and reproducibility from laboratory to laboratory are important
334 aspects for a validated method. It is acknowledged that to some extent a case by case
335 adaptation may need to be applied. It is also recognised that usually methods to determine
336 pharmacodynamic actions may be less standardised than for instance safety pharmacology and
337 toxicology studies. Irrespective of the experimental procedures and materials the prediction of
338 the doses and regimens to be used clinically should be comparable. It is clear that at least
339 sufficient controls must be built into tests run with HFS-TB and that within a laboratory
340 procedures and materials should be well described and justified for the purpose of the study.
341 Nevertheless, the consortium will make a standard operating procedures lab manual available
342 and a limited series of experiments with Moxifloxacin and PA824 in triplicate are being planned
343 to assess inter- and intra-laboratory variability. This is a welcomed standardisation that may
344 contribute to wider usage of the HFS-TB system. The consortium is encouraged to submit
345 these data in a follow up qualification opinion.

346 Regardless of the limitations of the data submitted, the totality of evidence in scientific literature and
347 the cases seen in regulatory submissions support the qualification of the HFS-TB system.

348 **CHMP qualification opinion**

349 The HFS-TB is qualified to be used in anti-TB drug development programs as an additional and
350 complementary tool to existing methodology to inform selection of dose and treatment regimen,
351 including combination of 2 or more anti-Mtb drugs, to maximize bactericidal effects and minimize
352 emergence of drug resistance. HFS-TB can be used in regulatory submissions throughout the drug
353 development process for a product, especially for more informed design and interpretation of phase I,
354 phase II and phase III clinical studies. It should be noted that the qualification opinion does not
355 mandate the use of the HFS-TB or exclude the use of alternative methods in the confined setting.

356 More specifically CHMP recommends that the HFS-TB may be useful as follows:

- 357 • To provide preliminary proof of concept for developing a specific drug or combination to treat
358 tuberculosis
- 359 • To select the pharmacodynamic target (e.g. T/MIC, AUC/MIC)
- 360 • To provide data to support PK/PD analyses leading to initial dose selection for non-clinical and
361 clinical studies, with the aim of limiting the number of regimens that are to be tested in vivo; it
362 is anticipated that HFS-TB may be used to limit doses tested both in single drug and
363 combination regimen studies in vivo
- 364 • To assist in confirming dose regimens for later clinical trials taking into account the
365 accumulated human PK data in healthy volunteers and then patients as well as available
366 information on exposure-response relationships

367 **Annexes**

- 368 - Applicant submission – Request for CHMP Qualification Opinion
- 370 - Applicant submission – Response to Questions raised by the qualification team
- 371 - Applicant submission – Discussion Meeting for HFS-TB Qualification Opinion Request (Slides)

ⁱ All annexes mentioned under the Applicant's position refer to the documentation submitted with the request.