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Information for the package leaflet regarding polysorbates used as excipients in medicinal products for human use

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Executive summary

This document has been written in the context of the revision of the Annex of the European Commission Guideline on 'Excipients in the labelling and package leaflet of medicinal products for human use' (EC, 2018 [39]). Polysorbates were not listed in the latest Annex to the guideline dated 2022 (Annex, 2022 [3]), but in view of some safety issues (e.g., hepatotoxicity, potential cardiotoxicity, etc.) it was considered important to propose a warning for the package leaflet.

Polysorbate 80 (PS 80, polyoxyethylene sorbitan monooleate, also known as TweenTM 80) and polysorbate 20 (PS 20, polyoxyethylene sorbitan monolaureate, also known as TweenTM 20) are mixtures of the partial esters of sorbitol and its mono- and dianhydrides with oleic or lauric acid respectively, and condensed with approximately 20 moles of ethylene oxide per mole of sorbitol and its anhydrides. They are used as nonionic surfactants and as emulsifiers, being the most common surfactants used in biological medicinal products for protein stabilisation.

Although most available safety data is for PS 80 or 20, the package leaflet information should be used for all types of polysorbates unless otherwise justified.

Acute oral toxicity is low, which is probably attributed to the very low oral bioavailability of intact polysorbates. The acceptable daily intake (group ADI) for polysorbates as food additives (polysorbates 20, 80, 40, 60 and 65; E 432, E 433, E 434, E 435 and E 436, respectively) was set to 25 mg/kg body weight/day by EFSA in 2015 (EFSA ANS Panel, 2015 [32]).

In view of the estimated maximum oral dose of PS 80 or PS 20 in authorised medicinal products of about 1 mg/kg/day the oral exposure of PS 80 by oral formulations is estimated to be far below the ADI. Therefore, a warning on the effects of polysorbates as excipients by oral administration is not considered meaningful.

In contrast to the oral route, after intravenous administration (IV), the whole amount of intact polysorbates enters the bloodstream. As hypersensitivity reactions including anaphylactoid shock have been observed after IV, subcutaneous (SC) and intramuscular (IM) administration (including therapeutic proteins and vaccines) in patients showing positive Prick tests to PS 20 or 80, a warning about allergic reactions at threshold zero for the parenteral route is proposed.

It has to be noted that, after IV administration, metabolism of polysorbates may result in the formation of sorbitol, which itself is a source of fructose. This constitutes a potential risk for patients with hereditary fructose intolerance (HFI) and there is general advice for patients with HFI to avoid polysorbate as a potential source of fructose. The theoretical maximum amount of sorbitol is 14% (15%) of the polysorbate 80 (20) content. However, since to date neither PK nor clinical safety data has given evidence that sorbitol generation by polysorbates constitutes a definite risk for such patients, the decision was taken not to include the strict warning for HFI patients as required at threshold zero for parenteral medicinal products containing sorbitol (Information for the package leaflet regarding fructose and sorbitol, 2017 [58]).

A significant hemodynamic effect (short duration vasoplegia, left ventricular systolic decreased pressure) was observed in human adults after amiodarone IV bolus injection (Cordarone) containing 10 mg/kg PS 80 compared to a formulation without polysorbates and benzyl alcohol. In dogs, bolus injections ≥ 4.3 mg/kg of PS 80 alone lead to depression of the cardiac conduction and hypotension. By allometric scaling a human equivalent dose of about 3 mg/kg was estimated. From the totality of preclinical and clinical data a threshold of 3 mg/kg/day was derived for a cumulative daily dose, in worst case administered as a bolus injection, which triggers a warning regarding cardiovascular effects (e.g., low blood pressure) in humans.

The cardiovascular effects appeared to be related to the infusion rate rather than to the cumulative dose. This might also explain the apparent safe use of MVI Pediatric (Multi-Vitamins for Infusion), a US vitamin product for 24 h infusion resulting in relatively high cumulative PS 80 exposure (32.5 mg/kg/day in 1 kg neonates), but at a quite low infusion rate of 0.023 mg/kg/min. A small PK and safety study with anidulafungin infusions in infants and neonates with maximum PS 80 exposure of 7.7 mg/kg/day (max infusion rate over 60 min: 0.13 mg/kg/min) gives support that short-term exposure at low infusion rates of PS 80 < 8 mg/kg per day is safe even in infants and neonates.

Thus, a general recommendation for risk minimisation by lowering the rate of injection/infusion is given as a comment for consideration in the SmPC of parenteral products.

Risk for a cardiotoxic/torsadogenic potential of polysorbates is supported by in vitro data on hERG current inhibition, as well as from preclinical data showing an increase in effective refractory period (ERP) in guinea-pig cardiac preparations and in vivo in dogs. There is no evidence so far for depression of cardiac conduction from clinical data in humans, which would allow derivation of a safety threshold for cardiotoxicity. It is concluded that further (pre-clinical and) clinical electrophysiological studies are warranted to investigate the torsadogenic potential of polysorbate 80 in detail. Therefore, it is proposed that a warning on the risk of concomitant use of medications that prolong the QT/QTc interval is considered for the SmPC of all products containing polysorbates above the threshold of 3 mg/kg/day when given as bolus.

The hepatotoxic potential of polysorbates gained notoriety after the E-ferol tragedy in the 1980s when 38 infant deaths were reported after IV infusion of this Vitamin E formulation containing a mixture of polysorbate 80 (9%) and polysorbate 20 (1%) as solubilising agents. A clear dose-response relationship was found with an increased risk for severe hepatotoxicity in premature infants at a polysorbate dose of > 80 mg/kg/d. Data suggested that the cumulative doses over 6–45 days rather than short-term peak exposure levels appeared to be relevant for hepatotoxicity.

With regards to adults, case reports at exposures below 80 mg/kg/d may indicate an earlier onset of signs of hepatotoxicity: 35–40 mg/kg were calculated as the cumulative polysorbate dose within 24h identified in case reports of hepatotoxicity in adults after Amiodarone IV, e.g., showing abrupt elevation of liver enzymes. Such case reports are confounded by the fact that amiodarone itself is a hepatotoxic agent. However, the observation that subsequent oral amiodarone administrations in patients did not result in additional liver toxicity supports the association with the intravenous exposure of the excipient.

In conclusion, a parenteral threshold of 35 mg/kg/d for all age groups is suggested to trigger a warning for elevation of liver enzymes.

Polysorbates exposure via administration of therapeutic proteins and vaccines is very low (≤ 1.2 mg/kg) and therefore below all the proposed thresholds apart from zero. This is considered appropriate as it is in line with the absence of any signal of cardiotoxicity or hepatotoxicity after vaccines or therapeutic proteins exposure from epidemiology or pharmacovigilance.

The ability of polysorbates at high intravenous doses (> 3 mg/kg) or as a coat on drug nanoparticle formulations to enhance the uptake of drugs into the brain constitutes a potential interaction with drug substances and should be taken into account during drug development and benefit-risk evaluation of current and new parenteral products containing polysorbates.

Proposal for new information in the package leaflet

Name	Route of Administration	Threshold	Information for the Package Leaflet	Comments
Polysorbates e.g.: PS 20 (E 432) PS 80 (E 433) PS 40 (E 434) PS 60 (E 435) PS 65 (E 436) PS 85	Parenteral	Zero	This medicine contains x mg of polysorbate <type> in each <dosage unit><unit volume> <which is equivalent to x mg/<weight><volume>>. Polysorbates may cause allergic reactions. Tell your doctor if <you have><your child has> any known allergies.	
		3 mg/kg/day (total amount of all PS in the medicinal product)	Polysorbates can have an effect on your heart and blood circulation (e.g., irregular or abnormal heartbeat, or low blood pressure).	Risk minimisation by lowering the rate of injection/infusion is recommended for consideration in the SmPC of parenteral products. Due to a potential for QT prolongation and torsades de pointes in humans, for risk minimisation, a SmPC warning on the risk of concomitant use of medications that prolong the QT/QTc interval or for patients with congenital syndrome should be considered.
		35 mg/kg/day (total amount of all PS in the medicinal product)	Polysorbates may have an effect on the liver. If you have a liver disease ask your doctor or pharmacist for advice.	Case reports in adults indicate an onset of signs of hepatotoxicity (elevated liver enzymes) at a cumulative daily dose of 35–40 mg/kg. In neonates, doses > 80 mg/kg/day of polysorbate caused severe (fatal) hepatotoxicity.
	Topical	Zero	Polysorbates can cause allergic reactions.	

Scientific background

Introduction

Polysorbates are non-ionic surfactants widely used as excipients in oral, topical and injectable medicinal product formulations (Garidel et al., 2009 [46]). The focus in this report lies on polysorbates 20 and 80 as the most relevant polysorbates excipients, polysorbate 80 being by far the most used one (according to the EMA's electronic database of centralised products (SIAMED) checked in October 2023).

Polysorbates are also widely used as an emulsifier, dispersant or solubiliser in many foods (e.g., E 432–36) and in a variety of cosmetic products.

Polysorbates are not listed in the Annex to the 'Guideline on Excipients in the label and package leaflet of medicinal products for human use' dated 2022 [3]. However, it is now proposed that they are added to the list of excipients for which safety issues should trigger a warning in the package leaflet, notably on parenteral preparations for paediatric population (e.g., potential cardiotoxicity).

This report summarises updated toxicological and safety data on polysorbates 80 and 20 and provides a risk assessment concluding on thresholds for PIL warning.

1. Characteristics

1.1. Category (function)

Emulsifying agent; nonionic surfactant; solubilising agent; wetting, dispersing/suspending agent.

1.2. Physico-chemical Properties

Definition

Mixtures of partial esters of fatty acids, mainly oleic acid (PS 80) or lauric acid (PS 20), respectively, with sorbitol and its anhydrides ethoxylated with approximately 20 moles of ethylene oxide for each mole of sorbitol and sorbitol anhydrides (Polysorbates 20, 40, 60, 65, 80, and 85). Some polysorbates are ethoxylated with shorter polyoxyethylene chains (4–5 oxyethylene units), such as polysorbate 81, 21 and 61.

Chemical Names and CAS Registry Numbers

Polyoxyethylene (20) sorbitan monooleate 9005-65-6

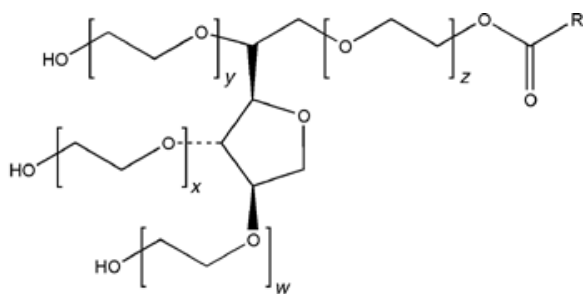
Polyoxyethylene (20) sorbitan monolaurate 9005-64-5

Empirical Formula and Molecular Weight

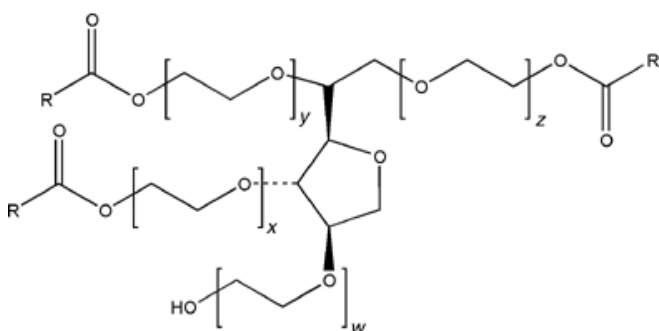
PS 80: $C_{64}H_{124}O_{26}$ Mr = 1310

PS 20: $C_{58}H_{114}O_{26}$ Mr = 1228

Structural Formula



Polyoxyethylene sorbitan monoester



Polyoxyethylene sorbitan triester

$w + x + y + z = 20$ (Polysorbates 20, 40, 60, 65, 80, and 85)

$w + x + y + z = 5$ (Polysorbates 81)

$w + x + y + z = 4$ (Polysorbates 21 and 61)

R = fatty acid (Polysorbate 80: > 58% oleic acid; Polysorbate 20: 40–60% lauric acid)

Typical Properties (Ph.Eur. monograph 01/2017: 0426 and 0428; Handbook of pharmaceutical excipients 2012 [53]; Wan and Lee; 1974 [132])

	Values (PS 80 / PS 20)
Acid value	≤ 2.0 for PS 80 and PS 20
Acidity/alkalinity	pH = 6.0–8.0 for a 5% w/v aqueous solution
Critical micelle concentration (CMC) at 25°C (µg/ml)	$\approx 14 / 60$
Flash point	149°C for PS 80
HLB value	15.0/16.7
Hydroxyl value	65–80/96–108
Moisture content	≤ 3.0 for PS 80
Saponification value	45–55/40–50

	Values (PS 80 / PS 20)
Solubility D = dispersible; I = insoluble; S = soluble; T = turbid; W = on warming.	Ethanol = S Mineral oil = I Vegetable oil = I Water = S
relative density at 20°C	1.08 / 1.11
Surface tension at 20°C (mN/m) for 0.1 % w/v solutions	42.5 for polysorbate 80
Viscosity (dynamic) (mPa s)	425 / 400

Stability and Storage Conditions

Polysorbates are stable in the presence of electrolytes and weak acids and bases; gradual saponification occurs with strong acids and bases. Polysorbates are hygroscopic and should be examined for water content prior to use and dried if necessary. Upon storage, polysorbates are prone to oxidation and formation of peroxides (the oleic acid esters are sensitive to oxidation notably due to photosensitivity).

Polysorbates should be stored in a well-closed container, protected from light, in a cool, dry place.

Incompatibilities

Discoloration and/or precipitation occur with various substances, especially phenols, tannins, tars, and tarlike materials. The antimicrobial activity of paraben preservatives is reduced in the presence of polysorbates. Also, publications indicate auto-oxidation of polysorbate (80 or 20) in aqueous solution resulting in hydroperoxide formation, reactive aldehydes including formaldehyde and acetaldehyde, or side-chain cleavage, that could influence the stability of proteins (e.g., Maggio, 2012 [78]).

Polysorbates are widely used as a stabiliser for formulation of proteins avoiding their aggregation. Their compatibility with the active substance should therefore be demonstrated before any approval.

Also, polysorbate 80 is known to increase the rate of di-(2-ethylhexyl) phthalate extraction from polyvinyl chloride (PVC) materials (Takehisa et al., 2005 [119]). Therefore, the demonstration of the suitability of, for example, the primary packaging by compatibility studies including studies on extractables and leachables during pharmaceutical development is required where appropriate (Sharma, 2007 [109]).

1.3. Use in medicinal products

Polysorbates 20 and 80 are non-ionic surfactants which are widely used as excipients in oral, topical, and injectable formulations (Garidel et al., 2009 [46]). For example, they are included in over 3000 medicinal products authorised in Germany (March 2017) and close to 2000 in The Netherlands. In medicinal products for oral use, polysorbates 80 and 20 are used in coated and uncoated tablets, capsules, oral solutions, and suspensions. The polysorbate content is variable and ranges from 0.02 to 66 mg per dose in centrally authorised solid formulations. Polysorbates also serve as solubilising

agents in many injectable formulations of poorly soluble active substances (e.g., docetaxel, amiodarone). The highest polysorbate exposure per kg BW was calculated as 55 mg/kg per dose (Taxotere, active substance docetaxel, 26 mg PS 80 per mg of drug (according to ten Tije et al. 2003b [121]), 75 mg docetaxel/m²; 60 kg adult). An infusion of 5 mg/kg amiodarone (usual dose) leads to a PS 80 exposure of 10 mg/kg (content of 2 mg PS 80 per mg of amiodarone in Cordarone).

Polysorbates are also added in certain multivitamin solutions to dissolve liposoluble and hydrosoluble vitamins in the same medium and are present in many biological medicinal products such as enzymes (alteplase), immunoglobulins and monoclonal antibodies for both preventing surface adsorption and as stabilisers against protein aggregation. The concentrations range varies from 0.0003% to 0.3% (w/v) (Kerwin, 2008 [63]). Low amounts of Polysorbate 80 (20 µg/ml) have been added to SCIG (Hizentra) to improve the visual appearance of the solution, as highly concentrated IgG solutions do not have a homogenous appearance (Maeder et al., 2011 [77]). Po exposure from therapeutic proteins is estimated to be much lower than from small molecules, i.e. < 1.2 mg/kg per dose.

Furthermore, polysorbates 80 and 20 are used in vaccines, either as stabiliser of the antigen preparation or as emulsifying agent in emulsion adjuvants. Examples for emulsion adjuvants are MF 59, an oil-in-water (o/w) emulsion containing droplets of squalene surrounded by a monolayer of non-ionic surfactants PS 80 and sorbitan trioleate (Span 85) (Shultze et al., 2008 [111]), and AS03, an oil-in-water emulsion adjuvant system containing α-tocopherol and squalene surrounded by PS 80 (4.86 mg per dose) (Langley et al., 2012 [72]).

Systemic exposure to polysorbates by most vaccines is very low due to their low amount (up to 0.75 mg per human dose corresponding to ≤ 0.01 mg/kg). Somewhat higher levels are reached with vaccines containing PS in the adjuvant system (up to 4.85 mg per dose (0.08 mg/kg), i.e. < 0.1 mg/kg).

1.4. Regulatory status in food

In the EU, polysorbates 20, 40, 60, 65 and 80 were approved as food additives and standards for their use were established (Annex to the Commission regulation (EU) No 1130/2011 [24]; Polysorbate 80 = E 433). In its scientific opinion, the EFSA Panel on Food Additives and Nutrient Sources Added to Food concluded that, based on the NOAEL of 2 500 mg/kg bw/day, identified from an oral carcinogenicity study with polysorbate 80 in rats, and applying an uncertainty factor of 100, a group ADI of 25 mg/kg bw/day for polysorbates 20, 80, 40, 60 and 65 (E 432, E 433, E 434, E 435 and E 436, respectively) could be established (EFSA 2015 [32]). It was further estimated that exposure of toddlers at the highest level was very close to the ADI (24.5 mg/kg bw/day).

1.5. Regulatory status in cosmetics

Polysorbates are used as hydrophilic, nonionic surfactants in a variety of cosmetic products. The Cosmetic ingredients expert panel reviewed the safety of polysorbates in 1984 (CIR [25]). The report states that these ingredients are used in numerous preparations without clinical reports of significant adverse effects. It was concluded that they are safe for use in cosmetics at present concentrations of use.

2. Pharmaco-toxicological data

Thorough reviews on the pharmacology/toxicology of polysorbates were conducted by the Joint FAO/WHO Expert Committee on Food additives (JECFA) in 1974 (Joint FAO/WHO [62]), the Cosmetic Ingredient Review Expert panel (US) in 1984 (CIR [25]), the EFSA Panel on Food Additives and

Nutrient Sources added to food (ANS) in 2015 (EFSA 2015 [32]) and the Japan Food Safety Commission in 2007 (FSC [43]). These reviews were used as a basis for the assessment.

2.1. Pharmacodynamics and safety pharmacology

Polysorbates have been shown to activate or inhibit numerous biochemical reactions in vitro (enzymes, cellular respiration, DNA replication etc.) which may not be necessarily indicative of the in vivo effects of the polysorbates (CIR, 1984 [25]).

The pharmacodynamics effects observed on the cellular level in vitro or in vivo and the effective concentrations/doses are summarised in Table 1 (see end of section 2.1).

2.1.1. Biological membranes

Due to the surface-active properties of the polysorbates and the physicochemical nature of cellular membrane bilayers, the polysorbates can affect the structure and function of biological membranes. Because of its dual hydrophobic/hydrophilic nature, polysorbate 80 in solution tends to orient itself so that the exposure of the hydrophobic portion of the molecule to the aqueous solution is minimised. Extensive studies have been made on the action of non-ionic surfactants using test systems ranging from artificial lipid monolayers to natural multilayer epithelia.

Whether the effect the polysorbates have on membranes is solely a function of their hydrophile-lipophile balance or whether the specific structure of the polysorbate molecule could determine its biological activity is unclear. For example, it was concluded that the lysis of erythrocytes by the polysorbates was caused not by the destruction of the membrane but by some rearrangement of the membrane structure accompanying adsorption of the surfactant. Electrophysiological studies in artificial membranes indicated that polysorbates lower the conductance of the membrane by making it less permeable to charged molecules and decrease membrane stability by becoming incorporated into the membrane structure (CIR, 1984 [25]). Recent investigations indicated that polysorbate 80 may increase the susceptibility of cells to oxidative stress (Tatsuishi et al., 2005 [120]).

Jelinek (2001 [60]) thoroughly investigated the cytotoxicity of several tensides in vitro: the lowest observed half-maximum cytotoxic concentration (CC50) for polysorbate 80 was 0.048 mg/ml determined by an XTT-assay with U937 cells (human monocytic cell line) after 24h incubation. As with other tensides, apoptosis inducing properties were predominant at low concentrations whereas at higher concentration necrosis and cell lysis become more prominent.

In agreement to this, other investigators observed cytotoxic effects in vitro at similar concentrations of 0.1 mg/ml (e.g., Ménard et al., 2011 [86], HUVEC model, derived from LDH release) respectively.

The critical micelle concentration (CMC) for polysorbate 80 has been reported in a range between 14 µg/ml (12 µM) and 26 µg/ml (about 20 µM) (Wan and Lee, 1974 [132]; Kerwin, 2008 [63]; Ménard et al., 2011 [86]). Comparison of these findings with the cytotoxicity data discussed above shows that both apoptosis induction and cell lysis induction occur at PS 80 concentrations above the CMC.

Many investigators have shown that PS 80 can interfere with the function of the transmembrane drug-export pump P-glycoprotein (P-gp, MDR1) either directly or through membrane perturbations, modulating multidrug resistance (MDR). Polysorbate 80 inhibits P-gp over a range from 0 to 1 mM, while it increased apical-to-basolateral permeability (AP-BL) and decreased basolateral-to-apical (BL-AP) permeability of the P-gp substrate rhodamine 123. These P-gp inhibition effects would appear to be related to these excipients' modulation of membrane fluidity, where PS 80 fluidises cell lipid bilayers. PS 80 also inhibits the peptide transporter, as measured by glycyl sarcosine permeability

(Rege, 2002 [102]). Polysorbate 20 has also been reported (Yang et al., 2012 [138]) to increase significantly intracellular accumulation of doxorubicin in vitro via a possible mechanism of inhibiting MDR1 function and expression.

This MDR1-reversing ability was used to develop a bioassay for polysorbate 80 (Webster et al. 1997 [134], see PK chapter). Complete reversal of MDR1 in vitro occurs at polysorbate 80 concentrations of 1–2 µl/ml, 50% inhibition occurs at levels of 0.2–0.3 µl/ml (corresponding to 0.22–0.33 mg/ml; Webster et al., 1997 [134]). Drori et al. (1995 [31]) demonstrated that Tween™ 80 alters membrane fluidity and increases membrane permeability and that these changes in the physical properties of biomembranes are important factors in achieving potentiation of anticancer-drug cytotoxicity.

A more recent review (Zhang et al., 2016 [141]) suggests that polysorbates may interfere with the function of also other efflux proteins such as BCRP or MRP2 as well as metabolic enzymes in the CYP family (e.g., CYP3A4, CYP2C9). Specific polysorbates may differ in their activity profile with regards to which efflux transporters and/or metabolic enzymes are affected (see also 3.3 Interactions).

Polysorbates produce various, seemingly disparate effects in neuromuscular systems. Both stimulation of colonic motility but also clear spasmolytic activity of Polysorbate 80 was found in animal studies (CIR, 1984 [25]).

2.1.2. Blood brain barrier

It has been known for a long time that polysorbate 80 increases the uptake of drugs into the brain (Azmin et al., 1985 [4]). Azmin et al. tried to investigate the mechanism by which polysorbate 80 enhances brain uptake of intravenous methotrexate (MTX) in mice. They could show that increased brain levels of MTX were observed after intravenous administration of MTX plus PS 80 compared to MTX alone, and the reverse was true for the MTX serum levels, indicating a direct effect of PS 80 on the BBB. This effect was observed with the lowest intravenous systemic dose of PS 80 investigated (3.2 mg/kg), lower doses of polysorbate 80 have not been tested. A possible enhancing effect by polysorbate 80 on the elimination of MTX from plasma was also discussed (Azmin et al. 1985 [4]). Calvo et al. (2001) [20] showed that a polysorbate 80 intravenous dose of 20 mg/kg in rats dramatically increased BBB permeability to sucrose. Polysorbate 80-coated nanoparticles can deliver drugs to the brain by a still debated mechanism (Kreuter, 2013 [69]). Gulyaev et al. (1999 [51]) had demonstrated that intravenous polysorbate 80-coated nanoparticles (coated by stirring in a 1% PS 80 solution) were able to deliver doxorubicin to the brain of rats. The highest levels were achieved between 2 and 4 h after drug administration. Administration of free doxorubicin in saline, or in 1% PS 80 solution or loaded to non-coated nanoparticles could not enhance brain uptake. These data correlate with the notion that coated nanoparticles reach brain endothelial cells essentially intact. Adsorbed on the particle surface, PS 80 may be delivered more efficiently to the brain endothelial cell. This could explain why the addition of PS 80 surfactant solution to free doxorubicin was totally inefficient.

Induction of endocytosis and/or transcytosis of the coated particles is favored as underlying uptake mechanism by polysorbate 80, but also membrane lipid solubilisation, opening of tight junctions or inactivation of the P-glycoprotein efflux pump could contribute to the effect (Kreuter, 2013 [69]). Polysorbate 80 stabilised nanoparticles adsorb preferentially apolipoproteins E or B that have been found responsible for the interaction with the BBB and the subsequent endocytosis/transcytosis (Göppert et al., 2005 [50]; Zensi et al., 2009 [139]). Later investigations by Koffie et al. further supported that PBCA nanoparticles coated with polysorbate 80 do not induce nonspecific BBB disruption but collaborate with plasma apolipoprotein E to facilitate BBB crossing (Koffie et al., 2011 [68]).

High accumulation of edelfosine in brain was reported by Estella-Hermoso de Mendoza et al. (2011 [38]). The authors suggested it was due to the inhibition of P-glycoprotein by Tween™ 80, as verified using a P-glycoprotein drug interaction assay. In vitro studies revealed that edelfosine-loaded lipid nanoparticles induced an antiproliferative effect in C6 glioma cell line.

2.1.3. Cardiovascular effects

Hemodynamics

There have been several studies on the hemodynamic effects of the polysorbates. The effects of the polysorbates vary from species to species, with a general trend toward a depression of cardiac output. When a 5% aqueous solution of polysorbate 80 was injected intravenously in doses of 1 ml/kg into cats, rabbits, and rhesus monkeys, there was a slight and transient fall in blood pressure; dogs exhibited a prolonged depressor response. This effect was never elicited by oral administration of the polysorbates (CIR, 1984 [25]).

Varma et al. found a dose-dependent fall in blood pressure of dogs with an average fall of 42 mmHg after the lowest IV dose of 5×10^{-3} ml/kg (corresponding to 5 mg/kg; Varma et al., 1985 [130]). Masini et al. demonstrated that histamine release is the main cause of the cardiovascular effects of polysorbate 80: histamine releasing properties have been demonstrated in vitro on isolated mast cells and in vivo in the dog. Administration of a dose of 10 mg/kg to a dog over 5 min (equals to an infusion rate of 2 mg/kg/min) produced severe hypotension accompanied by an increase in plasma histamine. H1- and H2-receptor blockade significantly reduced the cardiovascular effects. The authors concluded that the hypotension induced by the commercial intravenous amiodarone in dogs and humans is not due to amiodarone but to its solvent PS 80 (Masini et al., 1985 [83]).

More recent findings in dogs by Cushing et al. (2009 [30]) also support that the hypotensive effects observed after amiodarone IV result from the cosolvents used in its formulation (i.e. polysorbate 80/benzyl alcohol): no significant hemodynamic changes were found in dogs using a novel intravenous cyclodextrin-based formulation of amiodarone compared to the response observed with the commercial US formulation (Abraxis®) as well as with a vehicle formulation containing PS 80 and benzyl alcohol only (Cushing et al., 2009 [30]). The Abraxis® dose of 2.14 mg/kg amiodarone (equals to a PS dose of 4.28 mg/kg) was given as a bolus push or as a 10 min infusion (rate: 0.43 mg PS/kg/min).

Cardiotoxicity

Non-clinical in-vitro and in-vivo electrophysiology

Polysorbate 80 (Tween™ 80) inhibits hERG currents with a half-maximally inhibitory concentration of 0.02% (IC₂₀ 0.001%; Himmel 2007 [55]). According to the author the inhibitory effect by PS 20 is similar to (rather weaker than) PS 80. Part of the inhibitory effect is attributed to their interaction with lipid membranes, because hERG inhibition occurs close to critical micelle concentrations (Tween™ 20: ~ 0.007%).

Batey et al. (1997 [9]) found that 0.001% polysorbate 80 in combination with 1% DMSO (vehicle for halofantrine) increased the effective refractory period in guinea-pig right ventricular strips and left papillary muscles; the authors concluded that "The ability of the vehicle to prolong the effective refractory period in the ventricular preparations may be due to blockade of an outward K⁺ current such as I_{Kr} (...) it would seem likely that the observed increases in effective refractory period in ventricular preparations could be due to DMSO".

Torres-Arraut et al. (1984 [125]) studied the electrophysiological effects of polysorbate 80 in the cardiac conduction system of the dog and found that IV administration of 10 and 20 mg/kg

(cumulative) polysorbate 80 (equivalent to the amount of diluent in 5 and 10 mg/kg amiodarone respectively of commercial intravenous amiodarone) induced prolongation of the sinus node recovery time, depressed AV-nodal function and increased the atrial effective refractory period (ERP). Most importantly, at 20 mg/kg, polysorbate 80 increased ventricular ERP. The authors concluded that polysorbate 80 is a potent depressant of the cardiac conduction system in the dog and its electrophysiologic effects are similar to those of amiodarone.

In line with this, Varma et al. (1985 [130]) found a dose-dependent negative inotropic as well as negative chronotropic effect of PS 80 on guinea pig and rabbit paired atrial preparations.

After IV administration (1 h infusion) of polysorbate 80 at doses of 3–4.5 g, end of infusion plasma concentrations of polysorbate 80 in humans were about 0.1 µl/ml (i.e. in the 0.01% range; Webster et al. 1997 [134]). At similar concentrations, polysorbate 80 was found to inhibit hERG currents (IC₅₀ value of 0.02%; Himmel, 2007 [55]). A retrospective analysis of literature data indicated that block of hERG currents is associated with life-threatening Torsades de Pointes (TdP) cardiac arrhythmias if it occurs at concentrations close to those achieved in clinical use, and a 30-fold margin between free therapeutic plasma concentrations and IC₅₀ values for block of hERG currents appears to be a line of demarcation between the majority of drugs associated with Torsades de Pointes (TdP) arrhythmias and those which are not (Redfern et al., 2003 [101]).

The observation of Batey et al. (1997 [9]) that 0.001% polysorbate 80 in combination with 1% DMSO increased the effective refractory period in guinea-pig right ventricular strips and left papillary muscles might be due to block of I_{Kr} (which is encoded by hERG in humans) by polysorbate 80, and not due to DMSO as suggested by the authors. Himmel (2007 [55]) found that DMSO, at a concentration of 1%, inhibits hERG currents only by 16%, and similar weak or absent effects of DMSO on hERG currents have been observed (Zünkler et al., unpublished observations). In contrast, polysorbate 80 at a concentration of 0.001% (those tested by Batey et al., 1997 [9]) has been found to inhibit hERG currents by 20% (Himmel, 2007 [55]).

The preservative chlorobutanol and the hERG channel pore-blocker terfenadine synergistically inhibit hERG currents (Friemel and Zünkler, 2010 [44]), and it might be speculated that similar synergistic effects on hERG channels could occur after administration of polysorbate 80 in combination with other hERG channel blockers. Given the proposed mechanism of action of polysorbate 80 on hERG channels (an interaction with lipid membranes (Himmel, 2007 [55])), the similar potency for polysorbate 80 to inhibit both hERG currents and MDR1 and the effects observed in dogs after IV administration of PS 80 (depression of the AV-nodal function in addition to increasing the atrial and ventricular effective refractory periods), one may speculate that polysorbate 80 is a “multi-ion channel blocker” in the heart inducing cardiac electrophysiological effects not only via block of I_{Kr} (hERG channels).

2.1.4. Immune system

Complement activation/histamine release

Weiszhár et al. (2012 [135]) provided experimental evidence that polyethoxylated surfactants, such as polysorbate 80, activate the complement system in vitro, in normal human serum and plasma, generating the biologically active complement products, C3a, C5a and C5b-9. PS 80 appeared to be more efficient reactogenic than its structural homolog, PS 20. These results are consistent with the hypothesis that therapeutic side effects, such as acute hypersensitivity and anaphylactoid reactions, caused by intravenous medicines containing polyethoxylated detergents such as PS 80, can be attributed to complement activation-derived inflammatory mediators. However, in vitro complement activation was induced at high concentrations above 0.5% (equivalent to about 5 mg/mL; see

Table 1). Szebeni et al. (2005 [116]) have tentatively named such reactions as “Complement Activation-Related Pseudo Allergy (CARPA)” as a new class of drug induced toxicity including amphiphilic lipids. Coors et al. (2005 [26]) also identified polysorbate 80 as the causative agent for an anaphylactoid reaction of non-immunologic origin in a patient (see 4.1. Safety in adults)

Direct histamine releasing properties on isolated mast cells by polysorbate 80 have early been demonstrated in vitro (Masini et al., 1985 [83]). The mechanism of pseudo allergy caused by the polyoxyethylene nonionic surfactants (PNS) including polysorbates was investigated by Li et al. (2014 [75]). Based on in vitro cell analysis, it was assumed that PNS may enter mast cells via polyamine receptor-mediated endocytosis in a temperature-dependent and energy-dependent manner to induce mast cell degranulation followed by release of histamine. Data further suggest that isosorbide components of polysorbate 80 (polyoxyethylene isosorbide oleate) may be involved in causing pseudoallergy by polysorbate 80.

Polysorbate degradation products, e.g., free fatty acids, free radical species and other oxidation products such as aldehydes and ketones, have also been proposed as a cause of hypersensitivity reactions. They may function as haptens making degraded polysorbate a contact allergen (Singh et al. 2018 [113]).

Immunosuppressant effect

Mice given 0.3 ml intraperitoneal injections of 25% polysorbate 80 in saline solution prior to immunisation with ovalbumin absorbed to aluminium hydroxide demonstrated no primary IgE response, indicating that polysorbate 80 inhibited this response. Prior intraperitoneal injection of 0.3 ml of 25% polysorbate 80 in saline also caused a total suppression of the primary IgG response and a partial suppression of the passive haemagglutination response to ovalbumin in mice. Jerne plaque assays showed significant suppression of the primary antibody response. Mice treated with Tween™ 80 showed no significant decrease in contact sensitivity. Thus, the suppression caused by Tween™ 80 affected only the primary humoral immune response (Bryant and Barnett, 1979 [18]; Barnett and Bryant, 1980 [7]; Barnett, 1981 [8]; text excerpt from CIR review, 1984 [25]). Since very high doses of polysorbate 80 were used (intraperitoneal injection of 0.3 ml of 25% solution = 83.3 mg absolute dose of PS 80, corresponding to about 4167 mg/kg assuming a mouse weighing 20 g), the clinical relevance of these findings for the use as an excipient is questionable.

2.1.5. Tumor promotion/Tumor growth inhibition

Numerous reports are available on tumor promotion and co-carcinogenesis by polysorbates after application to the skin. Polysorbate effects on the skin that have been linked to tumor enhancement were the induction of epidermal hyperplasia (possibly due to its effect on biological membranes), inhibition of DNA repair, or facilitation of direct contact of a carcinogen with mucosal cell surfaces (CIR report 1984 [25]).

Several studies have shown that the polysorbates at higher concentrations also have tumor growth inhibition activity. Tumour growth inhibition by Tween™ 80 was reported in mice: intraperitoneal injection of polysorbate 80 into mice inoculated with carcinoma cells significantly reduced the formation and size of tumours and increased survival time of the animals (Witek et al. 1979 [137], Crispens et al. 1988 [27]). One author concluded that the cytotoxicity of polysorbate 80 for the tumour cells was related to the oleic acid component, since substitution of the polyoxyethylene sorbitan residue by diethanolamine did not eliminate the cytotoxic action (Witek et al., 1979 [137]).

Ng et al. (2004 [90]) demonstrated that the antiangiogenic property of taxanes can be significantly impaired by their formulation vehicles Cremophor EL and Tween™ 80, as well as serum binding

proteins. The underlying mechanistic basis is unclear. Tween™ 80 itself caused significant inhibition of angiogenesis at $\geq 5 \mu\text{l/ml}$ (corresponding to 5.5 mg/ml).

Table 1. Effective concentrations of PS 80 in vitro

PD parameter	Effective Concentration observed (LOEL, IC50)*	Equivalent concentration in mg/ml*	reference
Micelle association	CMC 20 μM	0.026 mg/ml	Kerwin, 2008 [63]
Cytotoxicity	Lowest CC50 48 $\mu\text{g/ml}$	0.048 mg/ml	Jelinek, 2001 [60]
P-gp (MDR1) inhibition	IC50: 0.2–0.3 $\mu\text{l/ml}$	0.22–0.33 mg/ml	Webster et al., 1997 [134]
Cardiotoxicity (hERG-channel inhibition)	IC ₅₀ 0.02%	0.2 mg/ml	Himmel, 2007 [55]
Haemolysis and cholestasis (isolated perfused rat liver)	1 $\mu\text{l/ml}$ in perfusate	1.1 mg/ml	Ellis, 1996 [33]
Histamine release (rat mast cells)	Lowest effect at 2.5 $\mu\text{l/ml}$ 50% release at 25 $\mu\text{l/ml}$	2.7 mg/ml 27 mg/ml	Masini et al., 1985 [83]
Complement activation	Significant effect $\geq 0.5\%$	5.5 mg/ml	Weiszhar et al., 2014 [135]
Antiangiogenic effect (rat aortic rings)	Significant effect $> 5 \mu\text{l/ml}$	5.5 mg/ml	Ng et al., 2004 [90]

* Relative density (20/20°C) of polysorbate 80 is about 1.1 g/ml (European Pharmacopoeia 8.1, 2014 [40]). Therefore, 1 μl of pure polysorbate solution equals 1.098 mg polysorbate.

Table 2. Effective doses of PS 80 in vivo

PD/Tox endpoint	Species	Admin route	NOAEL	Effective dose	reference
enhancement of brain uptake of other drugs	mice	IV		3.2 mg/kg (lowest dose tested; LOEL possibly lower)	Azmin et al., 1985 [4]
	rat	IV		20 mg/kg	Calvo et al., 2001 [20]
cardiac depression	dog	IV		10–20 mg/kg	Torres-Arraut et al., 1984 [125]
Hypotension	dog	IV		4.3 mg/kg $\geq 5 \text{ mg/kg}$	Cushing et al., 2009 [30] Varma et al., 1985 [130]

Hypotension (histamine release)	Dog	IV		10 mg/kg	Masini et al., 1985 [83]
Depression of primary immune response	mice	IV		> 4000 mg/kg	Barnett et al., 1980 [7]
neurobehavioral toxicity (PS 80)	rats	p.o.	2,013 mg/kg/d		Ema et al., 2008 [34]
diarrhea	rats	p.o.	4,500 mg/kg/d		EFSA, 2015 [32]

2.2. Toxicology

2.2.1. Single Toxicology

Polysorbates have a very low acute toxicity in rats and mice via oral, IP and IV routes with an LD50 > 2 g/kg/bw (BIBRA, 1992 [11]; Farkas et al., 1991 [41]).

2.2.2. Repeated Toxicology

Oral

Administration for three months with a dose of 10 mg/kg (0.2% wt/vol) PS 80 in mice, rats, dogs and monkeys was well tolerated (Thackaberry et al., 2010 [123]).

In a 13-week dietary administration study of PS 80 (0.31%, 0.62%, 1.25%, 2.5% and 5%) in F344 rats, no macroscopic or histological changes were observed in any organs.

In a similar conducted 13-week study in B6C3F1 mice, no effects were observed (Food Safety Commission, 2007 [43]).

Degenerative lesions were reported in the heart, liver and kidney after a daily oral administration of 1.5 ml of 1%, 2% and 3% PS 80 solution to rats for 3 months. However, no similar effects were seen in other studies (Food Safety Commission, 2007 [43]).

In mice given PS 80 in the diet for 10 weeks reversible liver damage developed. Unfortunately, no dose was specified (BIBRA, 1992 [11]).

No effects were reported in oral toxicity studies in rhesus monkeys who were given PS 80 0.1 g/kg/day for 10 months, and neither in rabbits given PS 80 3.6–55 g/kg/d for 65 days (BIBRA, 1992 [11]).

In repeated-dose toxicity studies diarrhea was observed as a major symptom. Based on the occurrence of diarrhea in rats fed polysorbates in subchronic studies, the lowest no observed adverse effect level (NOAEL) was calculated at 2% (PS 60, equivalent to 1,460 mg/kg body weight/day; EFSA, 2015 [32]).

Parenteral

Increased heart and kidney weight was observed without cellular damage after intramuscular injection of 0.6 g/kg/day for 1842 days in rats (BIBRA, 1992 [11]).

Intravenous administration for 65 days in rabbits of 1.3–2 g/kg/day of polysorbate 80 (as a 20% aqueous solution) resulted in kidney and spleen injury (BIBRA, 1992 [11]).

Local Tolerance

PS 80 stabilised NLC toxicity has been investigated using in vitro Eytex test and Draize test and no or minimal irritancy potential was scored indicating minimal toxicity or irritation potential to the external ocular tissues (Gonzalez-Mira et al., 2010 [49]).

Genotoxicity

In genotoxicity studies (in vitro and in vivo), it was concluded that polysorbate 80 was not mutagenic (Food Safety Commission, 2007 [43]).

Carcinogenicity

Hyperplastic lesions in B6C3F1 were increased at 50,000 ppm in male and females after a 2-year administration via diet, without a carcinogenic potential. No tumor genesis was identified in G57BL mice after 10 weeks of treatment up to 100 mg/mouse/d (Food Safety Commission, 2007 [43]).

After dermal administration in mice (skin-painting of 80 mg of undiluted PS 80 solution 6 times a week) for 52 weeks, only one mouse developed a benign skin tumor (Food Safety Commission, 2007 [43]).

A study in rats, after subcutaneous injection for 40 weeks of 2 ml of 6% polysorbate 80 solution (3 times a week) fibrosarcomas were identified at the injections site in 11 out 20 animals (Food Safety Commission, 2007 [43]).

No carcinogenic potential was demonstrated in hamster after intratracheal injection with 0.2 ml of 5% polysorbate 80 once a week for lifetime (Food Safety Commission, 2007 [43]).

Reproductive function toxicity

In reproductive studies performed with polysorbate 80 in rats (oral route), there were no effects on fertility, reproductive function and in morphological development, survival and growth of fetuses. Polysorbate 80 was not teratogenic (Food Safety Commission, 2007 [43]). The no observed adverse effect level (NOAEL) of polysorbate 80 for mother animals (rats) and the subsequent generation (F1) was considered to be 1.0 vol% (2,013 mg/kg body weight/day) as a level in drinking water.

In a previous study conducted by Enright et al. 2010 [36] in rats and rabbits after oral administration of 10 mg/kg of polysorbate 80 did not exhibit effects on fertility, or effects on early embryonic development in rats and no effects on embryo-fetal development in rabbits.

Neonatal/Juvenile Toxicity

Gajdova et al. (1993 [45]) investigated the influence of polysorbate 80 on the development of reproductive organs in neonatal female rats. Polysorbate 80 (1, 5 and 10% solution) was injected intraperitoneal for four days (days 4, 5, 6 and 7 after birth) and monitored up to 20 weeks. Statistically significant changes in vaginal opening time were observed for the medium and high dose group. In untreated animals the average length of the oestrous cycle was 4.3 compared to the range from 9.3 to 14 days in the treated animals. Further a decrease in relative weight of the ovaries in all

groups treated with polysorbate 80 was found compared to the control group. The authors concluded that the identified changes were similar to what have been seen after the administration of diethylstilbestrol, which was used as a positive control. Williams et al. (1997 [136]) could not confirm, after oral administration (up to 5 g/kg/d) for 3 days to immature rats (21 days after birth), the estrogenic effects of polysorbate 80 as it was observed in previous study by Gajdova et al. (1993 [45]), after intraperitoneal injection in neonatal female rats.

Investigation in young piglets (2 days after birth) that were treated with 2 to 4 ml/kg/day up to 13 days with a mixture of polysorbate 80 and polysorbate 20 after intravenous administration was found as not toxic. But the authors noted a recurring of subcutaneous edema in the neck region and several cases of necrotising enterocolitis at autopsy (Hale et al., 1995 [52]).

There are several reports on neurobehavioral toxicity of PS 80 available, conducted in rats, mice and cats (see Ema et al., 2008 [34]). To further evaluate the developmental neurotoxicity, including locomotor activity, of PS 80 a study in rats was conducted by Ema and Coworkers (Ema et al., 2008 [34]). Polysorbate 80 was given in their drinking water at concentration of 0.018, 0.13, 1 and 7.5% during pregnancy and lactation (day 0 of pregnancy until day 21 after delivery). For the locomotor activity no changes in male and female animals were observed during the treatment period. However, a decrease in successful conditioned avoidance responses was seen in the high dose group (7.5%), but no neurological changes were detected, including histopathological examination. The NOAEL considered being 1% (1,864 ml/kg/day) which is equivalent to 2,013 mg/kg/day. However, not enough information was provided if pups were directly exposed with PS 80 via maternal milk. The authors postulated that exposure took place partly via milk.

As pups gradually starts to consume food and water from around postnatal 14, it can be assumed that the pups were exposed with Polysorbate 80 onwards from this age via the drinking water (OECD, 2008 [91]).

The potential neurodevelopmental and maternal toxicity was studied in animal model e.g., by Brubaker et al. (1982 [17]). It occurred at the 7.5 vol% treated group. In a $\leq 1\%$ treated group, there was no effect on the mother rats and their subsequent generations (F1).

Farkas et al. conducted a study to assess the toxic effect of a 9:1 polysorbate 80: 20 mixture (2.5–4 mg/kg) in newborn rats, 2 days old, after a single intraperitoneal injection (Farkas et al., 1991 [41]). The LD50 (90-day) for neonatal rats was 3.5 g/kg. The main toxic effects observed in newborn rats were chylous ascites (milky fluid in the abdominal cavity), peritoneal fibrosis and severe tail inflammation. The authors concluded that the LD50 for neonatal rats is similar to that of adults.

Comparative toxicity study of IV administered alpha-tocopherol and alpha-tocopherol acetate and polysorbate vehicle containing formula (similar to that used in commercial preparations) was carried out on newborn rabbits by Rivera and co-workers. No toxicity could be attributed to the vitamin E or polysorbate treatment, but the pups treated with the polysorbate containing formula had microscopic evidence of mild bile stasis and elevated serum bilirubin levels, and lipidosis in the adrenal gland was primarily observed also in this group (Rivera et al., 1990 [104]).

2.3. Toxicokinetics

2.3.1. Oral administration

Studies have shown after oral administration in rats that the ester bond sites of polysorbate are hydrolyzed by pancreatic lipase and the free fatty acids then absorbed from the digestive tracts and oxidised. Excretion is mainly via exhaled breath as carbon dioxide. The kinetic is almost similar as

observed for the metabolism of ordinary fatty acids. The efficiency with which rats hydrolyzed and absorbed the labeled fatty acid portions of polysorbate 80 when fed at a dietary level of 10% was 100%. The polyoxyethylene sorbitan moiety left after hydrolysis of the ester is poorly absorbed from the rat's gastrointestinal tract. In one study with a radioactive carbon label in the polyoxyethylene portion of polysorbate 20, 90% was excreted in the feces and 8% in the urine. No radioactivity was found in the liver, carcass, or expired CO₂. When the sorbitol moiety of polysorbate 80 was labeled, 91% of the radioactivity was recovered in the feces, 2.1% in the urine, 1.6% in the carcass, and none in expired CO₂, liver, kidney, spleen, adrenals, brain, gonads, or fat. Since the radioactivity of both the sorbitol and polyoxyethylene labeled polysorbates is found largely in the feces and not in respired air, it is evident that there is no splitting of the ether bond between the oxyethylene group and the sorbitan moiety (Treon et al., 1967 [126]; CIR, 1984 [25]; Food Safety Commission, 2007 [43]).

These observations suggest that oral bioavailability of intact polysorbate 80 is extremely low. This is in line with data in mice where only about 3.2% of total oral administered polysorbate 80 was found to be excreted unchanged in the urine (Azmin et al., 1985 [4]). Also, the hydrolysed polyoxyethylene moiety appears to be poorly absorbed (< 10%) and is mainly excreted as such in the feces (Treon et al., 1967 [126]). In conclusion, after oral exposure almost solely the released fatty acid becomes systemically available. This substantiates the very low oral toxicity of polysorbate 80.

2.3.2. Intravenous administration

After intravenous injection into rats, the ester bond is also hydrolyzed by blood lipases. When polysorbate 20 was injected into rats, the labeled lauric acid moiety was metabolised and appeared mostly as expired CO₂ (68%; carcass, 22%; urine, 5%; feces and gastrointestinal contents, 2.5%; and liver 0.7%). The polyoxyethylene moiety was not catabolised since no radioactivity was recovered as CO₂ when this portion of the molecule was labeled. Most of the labeled polyoxyethylene (83%) appeared in the urine but some was present in the feces (11%) indicating biliary excretion (Treon et al., 1967 [126]).

Data in mice have shown that polysorbate 80 is also rapidly degraded after intravenous administration by esterases in plasma (van Tellinghen et al., 1999 [129]). The animals received an IV bolus dose of 3.3 µl of polysorbate 80:ethanol:saline (1:1:2, v/v/v) per g of body weight, corresponding to the amount of vehicle administered to animals receiving 33 mg/kg of docetaxel (0.83 µl/g, corresponding to about 0.9 g/kg). Within 15 min after bolus injection, the concentration of intact PS 80 measured by HPLC rapidly declined to levels < 0.05% (v/v) of the plasma volume. Parallel results obtained by studying the in vitro kinetics of PS 80 breakdown strongly suggest that esterases in plasma, catalyzing the cleavage of the oleic acid side chain from the PS 80 molecule, are responsible for this rapid decay.

3. Pharmacokinetics (in humans)

3.1. ADME (absorption, distribution, metabolism, elimination)

3.1.1. Oral administration

Clinical tests have shown that essentially the same pattern of metabolism is followed in humans as in the rat. The ethoxyl values of the urine and stools of four subjects fed 4.5 g of polysorbate 80 per day were determined to ascertain the amount of the polyoxyethylene portion excreted. The results showed that the polyoxyethylene fraction was excreted quantitatively; approximately 95% was excreted in the feces and 5% in the urine. Since there were no polyoxyethylenated fatty acids detected in the urine, it was concluded that the polyoxyethylene moiety in the urine represented polyoxyethylene sorbitan and

not the parent ester. The Polysorbate 80 was most likely hydrolysed by pancreatic lipase, with the liberated oleic acid following the normal metabolic pathways of unsaturated fatty acids. The source of the polyoxyethylene in the urine was that portion absorbed from the upper intestinal tract following hydrolysis of the ester bonds, since there was no carryover of the polyoxyethylene sorbitan in the urine during the postmedication control periods, there was no storage of this moiety in the body. The possibility of oxalic acid poisoning from the polyoxyethylene component would seem negligible in light of its quantitative excretion. Urinary studies for oxalate content in patients taking oral Polysorbate 80 indicated no increase in oxaluria (copied from: CIR, 1984 [25]).

3.1.2. Intramuscular administration

No data on the systemic bioavailability of polysorbate 80 after intramuscular injection (e.g., with vaccines) are currently available.

3.1.3. Intravenous administration

Webster et al. (1997 [134]) measured polysorbate 80 plasma concentrations in patients following IV administration of etoposide or docetaxel formulated with polysorbate 80 using a bioassay with MDR1-expressing cells (i.e. polysorbate 80 in plasma was determined on the basis of its ability to reverse MDR1). Patients received docetaxel containing 3.12–4.45 ml polysorbate 80 (corresponding to 3.4–4.9 g), and the median end-infusion (1 h) polysorbate 80 concentration was 0.1 µl/ml (corr. to 0.1 mg/ml) (range 0.07–0.41 µl/ml; only 1 patient had a level of > 0.2 µl/ml). Patients received IV etoposide containing 0.67–0.93 ml polysorbate 80, and in the end-infusion plasma sample PS 80 was not detectable (< 0.06 µl/ml). No time points other than “end of infusion” were investigated.

Sparreboom et al. have developed and published a method for determination of polysorbate 80 in plasma samples by liquid chromatography-tandem mass spectrometry (Sparreboom et al., 2002 [114]). By using this method, human PK data for PS 80 (Tween™ 80) after single dose IV infusion of different doses of docetaxel (Taxotere®) have been obtained from 39 cancer patients (ten Tije et al. 2003b [121]). Noncompartmental analyses yielded mean values of 7.7 L/h for total plasma CL, 3–8 L for Vss and 0.6 h for the terminal half-life. Plasma exposure (C_{max} and AUC) increased linearly with dose. After the lowest single dose of PS 80 (drug dose: 25 mg/m², Tween™ 80 content 26 mg/mg drug -> PS 80 dose 650 mg/m²) mean plasma C_{max} after 1 h infusion was 0.139 mg/ml (n=3), and after the highest individual dose (drug dose 75 mg/m² -> polysorbate 80 dose 1950 mg/m²) mean plasma C_{max} (n=19) was 0.457 mg/ml.

These intravenous human data provided evidence that polysorbate 80 has a high plasma clearance, a very short half-life (< 1h), and distributes mainly inside the blood compartment. Apart from the data described above by the Sparreboom group no further human PK data of any polysorbate excipient after intravenous administration have been published to date.

However, discrepancy is noted between the duration of 4 h for 50% breakdown by human plasma or pure esterase determined in vitro (van Tellingen et al., 1999 [129]) and the much more rapid decline observed in vivo (Sparreboom et al., 2002 [114]). Recent in vitro data show that polysorbate 80 is also metabolised by human liver carboxylesterase-1 (Zhang et al., 2020 [142]) suggesting that the liver may also be involved in the clearance.

In principle, metabolism of polysorbates in the plasma and liver could potentially result in the formation of sorbitol, which itself is a source of fructose. This constitutes a potential risk for patients with hereditary fructose intolerance (HFI), and polysorbates are mentioned among agents to be considered (Gaughan et al., 2021 [47]; Maiorana et al., 2020 [79]). The theoretical maximum amount

of sorbitol is 14% (15%) of the polysorbate 80 (20) content. There are no PK data on sorbitol generation in vivo which could support estimation of velocity and relevance of sorbitol generation.

3.2. Pharmacokinetics in children

In general, for many PK variables (also plasma esterase activity) there are clear differences between neonates and older infants and children (Morselli, 1976 [87]). Ester hydrolysis is low in newborns, and it appears to be significantly related to the developmental stage. Reduced activity of acetylcholinesterases and arylesterases in premature and full-term newborns are reported. The progressive increase in esterase activity with age was paralleled by the increment in plasma proteins; both parameters achieving adult values at 1 year of age (Morselli, 1976 [87]). Whether this holds true also for plasma esterase/lipase hydrolysing PS 80 is not clear. Age-dependent development in liver carboxylesterase activity, as described by Hines et al. (2016 [56]), is likely to further contribute to potential differences the clearance of polysorbates in particular for neonates.

As a conservative conclusion in analogy to other enzyme activities, in children < 1 year of age intravenous polysorbate 80 might be expected to be metabolised more slowly bearing a higher risk for adverse effects.

Pesce and McKean (1989 [98]) reported cases of death of several neonates after parenteral administration of a vitamin-E preparation (E-ferol), which contained 9% of polysorbate 80 and 1% polysorbate 20. Analysis of peritoneal fluid from a baby given E-ferol, a vitamin E supplement, revealed levels as high as 100 µg/ml polysorbate (McKean and Pesce, 1985 [85]). According to the authors the reason for this appears to be in the inability of the neonate to metabolise the compound, however, they do not provide any data supporting this assumption. Of note, the authors state that studies in rat pups are invalid to study neonatal toxicity, since most experiments are done on rat pups which are much more mature compared to human preterm neonates.

Interestingly, based on the published case report of 36 low-birth weight, premature neonates who experienced toxicity and death following receipt of high doses of an intravenous vitamin E product (E-Ferol) containing polysorbate 80 for a period ranging from 6 to 45 days, the toxicity (e.g., the development of hepatic lesions) and death could occur several months after the cessation of E-Ferol (Martone et al., 1986 [80]). Therefore, monitoring plasma concentrations of polysorbate 80 (e.g., during a study treatment period) may not necessarily help identify early signs of toxicity.

3.3. Interactions

Polysorbate 80 is reported to influence the pharmacokinetics of other drugs (reviewed by Schwartzberg and Navari 2018 [107]).

Surface active agents are thought to produce micellar solutions in the intestinal lumen in much the same way as bile salts, thus enhancing the uptake of fatty acids. When fed to rats for 1 week at 0.1% and 1% of the diet, Polysorbate 80 augmented the absorption of fats present at 10 to 33% of the diet (CIR report, 1984 [25]).

It has been known for a long time that polysorbate 80 has the potential to increase gastrointestinal absorption of other drugs as well as the uptake of drugs into the brain (Azmin et al., 1985 [4]). However, the amount of polysorbate 80 applied in this investigation (6, 12, 24%) is much higher than the amount typically used in oral medicinal products. This ability is utilised in the coating of nanoparticles with PS 80 for intravenous drug delivery to the brain (Kreuter, 2013 [69]).

At 0.01% in human serum, PS 80 decreased the binding of atropine sulfate to serum albumin (CIR report 1984 [25]).

There is increasing (in vitro, pre-clinical and clinical) research in the ability of surfactants to potentially modulate oral absorption through interaction with multiple transporters. Polysorbate 20 and 80 inhibit P-glycoprotein (P-gp/MDR1) thereby influencing intracellular accumulation of drugs and modulating multi-drug resistance. Furthermore, polysorbates may interfere with the function of also other efflux proteins such as BCRP or MRP2 as well as metabolic enzymes in the CYP family (see chapter 2.1.1. Biological membranes, and review by Zhang et al., 2016 [141]) In animals, surfactants such as pluronic® P85 and polysorbate 20 have been shown to enhance the oral absorption of P-gp and BCRP substrates (Al-Ali et al. 2019 [2]). Lee et al. (2021 [73]) demonstrated that Tween® 80 was an efficient inhibitor of BCRP function, as evidenced by the increased topotecan accumulation in BCRP-overexpressing cells. In vivo, Tween® 80 (1–2.5%) improved oral topotecan bioavailability (over 1.5-fold). For this type of products, the potential interaction should be considered during drug development and benefit risk assessment.

Effect of polysorbate 80 on metabolic activity of CYP3A4 and CYP2C9 in human liver microsomes has been revealed by Christiansen et al (Christiansen et al., 2011 [22]). Effect of polysorbate 80 on the expression of CYP3A4 mRNA and CYP3A4 protein has been studied by Tompkins et al in HepG2 and Fa2N4 human liver cell lines, human primary hepatocytes and intestinal LSI74T cell model (Tompkins et al., 2010 [124]). Polysorbate 80 tended to decrease CYP3A4 mRNA and protein expression in the above-mentioned model systems.

The ability of polysorbate 80 to form micelles leads to drug entrapment, significantly altering the disposition of the formulated drugs (ten Tije et al., 2003a [122]; Loos et al., 2003 [76]). In patients who received the same amount of polysorbate 80 that was present in 100 mg/m² of intravenous etoposide, both the volume and the clearance of doxorubicin were increased (Cummings et al., 1986 [28]).

The pharmacokinetic study by Wang et al. (2012 [133]) showed that the polysorbate 80 coated poly (-caprolactone)–poly (ethylene glycol)–poly (-caprolactone) micelles altered the biodistribution pattern and increased paclitaxel concentration in the brain significantly compared to the uncoated micelles and the free drug after intravenous injection in rats.

However, PK investigations of Docetaxel and PS 80 in mice by van Tellingen et al. (1999 [129]) indicated that the vehicle was not able to interfere in the disposition of docetaxel due to the rapid degradation of polysorbate 80 by esterases in plasma (in contrast to Cremophor EL, which was found to be causative for the observation of nonlinear kinetics of Paclitaxel). Because patients receive docetaxel by a 1-h IV infusion instead of a bolus injection (mice), the plasma levels of PS 80 remain much lower. Therefore, the authors concluded that in patients interactions by polysorbate 80 are even less likely.

On the other hand, Baker et al. (2005. [6]) observed an association between polysorbate AUC and unbound clearance of the drug docetaxel in patients with normal liver function.

There are two studies on the mechanism of the inhibitory effect of PS 80 on the intramuscular absorption of drugs. The inhibition of absorption could not be attributed to a direct or indirect effect on the capillary wall. It was concluded that the effect was mainly due to its influence on the extracellular space and the permeability of connective tissue (Kobayashi et al., 1974 [66]; Kobayashi, et al. 1977 [67]).

In a study in 8 patients with recurrent stage pTa or pT1 transitional cell carcinoma of the bladder, the rate of absorption of thioTEPA was not influenced by Tween™ 80, but it did cause statistically significant increases in mean peak plasma levels (from 101 to 154 ng/ml) and mean AUC values (from 0.376 to 0.496 micrograms h per ml) and a decrease in the mean half-life (from 1.83 to 1.25 h). The authors concluded that to obtain plasma levels similar to those achieved after instillation with thioTEPA alone, the dose should be reduced with Tween™ 80 (Masters et al., 1990 [84]).

4. Clinical safety data

4.1. Safety in adults

Hypersensitivity, pseudoallergy

The potency of polysorbate 80 as a type IV allergen is well-known. Tuberculin type hypersensitivity to PS 80 has been reported after water base formulated retinol injection to psoriatic patients and contact sensitivity to PS 80 by patch testing patients with eczema. Also, a high sensitisation rate to emulsifiers like polysorbate in patients with chronic leg ulcers was found (Pasche-Koo et al., 1994 [94]). Similar reactions were observed after IM injection of Vit. K (Aquamnophyton) with polysorbate 80 and not with preparation free of emulsifiers (Shelley et al., 1995 [110]). Few reports on polysorbate-induced contact allergy exist (cited by Coors et al., 2005 [26] and Palacios Castaño et al., 2016 [92]).

Coors et al. (2005 [26]) identified Polysorbate 80 as the causative agent for an immediate-type allergic shock reaction occurring in a patient after infusion of a multivitamin preparation containing polysorbate 80 (Multibionta N). A positive skin Prick test reaction to PS 80 but no Polysorbate-specific IgE antibodies were identified, suggesting a non-immunologic nature of the anaphylactoid reaction.

Early in the clinical development of docetaxel, it became clear that docetaxel infusion is associated with the occurrence of unpredictable (acute) hypersensitivity reactions, neutropenia, neurotoxicity, musculoskeletal toxicity and cumulative fluid retention. These side-effects have been attributed, in part, to the presence of high levels of the surfactant polysorbate 80 (Engels et al., 2007 [35]; Zhang et al., 2014 [140]). Also, the high rate of infusion related hypersensitivity reactions with fosaprepitant (an intravenous neurokinin-1 receptor antagonist for chemotherapy-induced nausea) is attributed to the high polysorbate 80 content in the formulation (Schwartzberg and Navari 2018 [107]; Boccia et al. 2019 [15]). Szebeni et al. (2005 [116], Szebeni 2014 [117]) have named such non-IgE-mediated hypersensitivity reactions to the intravenous injection of a variety of nanomedicines comprising liposomal, micellar, or polymer-conjugated drugs as "Complement activation-related pseudoallergy (CARPA)" (see 2.1.4. Immune system).

Owing to polysorbate 80 in its formulation, even prophylactic pre-medications were administered to prevent hypersensitivity reactions (Hennenfent and Govindan, 2005 [54]).

Regarding non-IV administrations, Palacios Castano et al. (2016 [92]) report 2 cases of anaphylaxis due to sensitisation to polysorbate 80 following intramuscular administration of a corticosteroid (Inzitan). A positive prick test performed with polysorbate 80 has indicated the role of this substance in the development of urticaria in a 28 year old adult after subcutaneous injection of Humira® and Stelara® (Perez-Perez et al., 2011 [96]). Further case reports on anaphylactic reactions to subcutaneous injections of therapeutic proteins like monoclonal antibodies or epoetins (Perino et al. 2018 [97]; Price et al. 2007 [99]; Bergmann et al. 2020 [10]; Steele et al. 2005 [115]) have been reported. Perino et al. (2018 [97]) describe the case of a teenager who presented with an anaphylaxis to the first injection of Xolair (omalizumab), with positive skin-prick tests to both Xolair and its excipient polysorbate 20, and cross-reactivity with polyethylene-glycol (PEG)/macrogol. Price et al.

(2007 [99]) describe two patients who experienced adverse reactions minutes after omalizumab administration after more than a year of uneventful treatment. None of the patients were IgE- or IgG-sensitised to omalizumab, one developed a positive intradermal test to polysorbate. Bergmann et al. (2020 [10]) report of anaphylaxis to the 13th injection of mepolizumab (including PS 80) and to a subsequent administration of omalizumab (PS 20) in a single patient who showed a positive skin prick test to polysorbate, probably due to sensitisation during long-term treatment with mepolizumab.

Furthermore, one case report of a hypersensitivity reaction due to polysorbate 80 in a human vaccine was identified: A 17-year-old girl reported generalised urticaria, eyelid angioedema, rhinoconjunctivitis, dyspnoea and wheezing 1 h after the third intramuscular administration of quadrivalent human papilloma virus vaccine (Gardasil; Badiu et al. 2012 [5]). Prick test performed with PS 80 and another PS 80-containing vaccine resulted positive. During the Covid-19 pandemic a huge number of people have been intramuscularly vaccinated with vaccines containing PS 80 (e.g., the vector-based vaccines Vaxzevria (AstraZeneca) and JCOVDEN (Janssen)). Allergic/anaphylactic reactions after Covid-19 vaccination due to PS 80 sensitisation have been, although very rarely, reported (Paoletti et al 2021 [93]); Ieven et al. 2021 [57]; Burlando et al. 2021 [19]).

Most authors suggest that the reactions were rather non-IgE-mediated (pseudoallergic) responses to the excipients PS 20 or 80, respectively, however, in some cases, esp. with occurrence after the third (or more) administration, an IgE-mediated genesis cannot be excluded Badiu et al. 2012 [5]; Palacios Castaño et al. 2016 [92]). Furthermore, people having pre-existing anti-PEG-IgE (prevalence 0.1% estimated from screening in normal human sera; Zhou et al. (2020 [143])) might also react positive in skin prick tests using polysorbates and polysorbate-containing medicinal products. Evidence from Covid-19-vaccinations during 2020/21 shows that clinical cross-reactivity between PEG and polysorbates occurs (Troelnikov et al. 2021 [127]; Ieven et al. 2021 [57]), but overall, the risk appears to be lower than expected. The PS 80 containing Covid-19 vaccine Vaxzevria has been shown to be tolerated by PEG allergic patients (Sellaturay et al. 2021 [108]); Ieven et al. 2021 [56]).

Hepatotoxicity/cardiovascular effects

Amiodarone

Rhodes et al. (1993 [103], case report of a 72y old adult) were the first to suggest polysorbate 80 as the hepatotoxic component in the IV formulation of amiodarone. Further case reports were published (Fonseca et al., 2015 [42]; Paudel et al., 2016 [95]; Ratz Bravo et al., 2005 [100]; Chen et al., 2016 [21]; Giannattasio et al., 2002 [48]).

In one case the patient was loaded with amiodarone 150 mg IV followed by amiodarone drip (1 mg/min for first 6 hours and then 0.5 mg/min for next 18 hours), a total dose of 1050 mg amiodarone was calculated (Paudel et al., 2016 [95]). This amiodarone dose corresponds to a cumulative dose of 2100 mg PS 80, i.e. 35 mg/kg in a 60 kg adult (content of 2 mg PS 80 per mg amiodarone assumed as in Cordarone®). A similar dose level is identified in a second case report (Fonseca et al., 2015 [42]): The patient was started on intravenous amiodarone with a bolus dose (injection over 3 min) of 300 mg followed by a continuous infusion of 900 mg over 24 h (1200 mg total dose amiodarone corresponding to 2400 mg PS 80, i.e. 40 mg/kg in a 60 kg adult). 18 h after starting amiodarone he showed an abrupt elevation of aminotransferases. As already shown in other cases, introduction of oral amiodarone in this patient did not result in any additional liver injury. Based on this observation, Rhodes et al. (1993 [103]) had proposed that polysorbate 80, the solvent of intravenous formulation of amiodarone, could be involved in this adverse effect since it is present in the intravenous but not in the oral form of amiodarone.

Munoz et al. (1988 [88]) investigated in 20 patients undergoing coronary arteriography the hemodynamic effects of an experimental preparation of IV amiodarone 5 mg/kg without Tween™ 80 (N) (10 patients) with those of the commercial form with Tween™ 80 (A; PS 80 dose: 10 mg/kg) (10 patients). Both A and N caused similar bradycardia, increase in ventricular filling pressure, vascular resistance and decrease in cardiac and contractility indexes. Amiodarone blood levels were similar after A or N. The data document a significant initial short duration vasoplegia, mainly related to Tween™ 80, after A, when amiodarone itself after producing a similar very slight effect causes bradycardia, and a moderate and progressive negative inotropic effect. Both preparations were injected as 3 min bolus, thus rate of PS 80-injection was 3.33 mg/kg/min. It was concluded that while the experimental form would be of interest, the risk of severe hypotension after IV Cordarone can be largely avoided by using a slower rate of infusion, especially in patients with hypovolemic status (Munoz et al., 1988 [88]). The observations are supported by earlier studies from Sicard et al. (1977 [112]) who found vasodilatation with associated tachycardia when injecting five patients with a quantity of pure Tween™ 80 equivalent to the amiodarone formulation.

Docetaxel

Some adverse effects occurring in the majority of cancer patients receiving Taxotere®, such as severe hypersensitivity reactions and fluid retention, are considered attributable to the excipients polysorbate 80 and ethanol. For that reason, many polysorbate-free formulations are currently under development (e.g., Li et al. 2014 [75]). Schwartzberg and Navari 2018 [107] report a diminished potential for hypersensitivity reactions with novel docetaxel formulations reducing the requirement of premedication with corticosteroids.

Tagawa et al. (2017 [118]) compared the adverse event profiles following injection of original or generic docetaxel in breast cancer patients. Significant product-related differences were observed in the following non-hematological adverse events: injection site reaction ($P = 0.0012$), hand-foot syndrome (\geq grade 3) ($P = 0.0003$), and oral mucositis (\geq grade 3) ($P = 0.0080$). Multivariate logistic regression analyses identified significant negative associations with the amounts of polysorbate 80 and ethyl alcohol present (Tagawa et al., 2017 [118]).

Taxotere® leads to the highest polysorbate exposure of all parenteral products (55 mg/kg; see Table 1). Impairment of liver function is among the common side effects. However, since docetaxel itself is potentially hepatotoxic, clinical cases of severe hepatotoxicity after docetaxel would not be attributed solely to polysorbate 80.

The potential metabolism of polysorbates to sorbitol after IV treatment (see chapter 3. Pharmacokinetics) could elicit hepatotoxicity due to sorbitol in HFI patients. It cannot be ruled out that single cases might be due to this rare disease (Curran and Havill, 2002 [29]).

Vaccine-induced narcolepsy

In 2012, observational studies in Finland Sweden reported an association between the occurrence of narcolepsy (chronic sleep disorder with excessive daytime sleepiness) and vaccination with a European A(H1N1) pandemic vaccine (Pandemrix®) during the H1N1 influenza pandemic 2009. In the following, several large epidemiological studies in other European countries confirmed an increased risk of narcolepsy in children, adolescents and adults after vaccination with AS03-adjuvanted pandemic vaccine Pandemrix. In search of possibly causative ingredients, also a contribution of the PS 80 containing emulsion adjuvant (AS03) has been addressed. Vaarala et al. (2014 [128]) found detergent-induced antigenic changes of viral nucleoprotein (NP), that are recognised by antibodies from children with narcolepsy, these results moved the focus from adjuvant(s) onto the H1N1 viral proteins. Since in contrast to Pandemrix® after vaccination with Arepanrix® (also adjuvanted with

polysorbate 80 containing AS03) or Focetria® (adjuvanted with polysorbate 80 containing MF59) only few cases of narcolepsy were reported, the difference between these vaccines came into focus (Jacob et al., 2015 [59]). Recent data indicate that Pandemrix-induced narcolepsy might be caused by an immune response against NP which is present in much higher amounts in Pandemrix than in Focetria (Ahmed and Steinman, 2016 [1]). Besides, Saariaho et al. (2015 [105]) found that patients with Pandemrix-associated narcolepsy had more frequently (14.6%) anti-GM3 antibodies than vaccinated healthy controls (3.5%) ($P = 0.047$). The data suggest that autoimmunity against GM3 is a feature of Pandemrix-associated NC and that autoantibodies against gangliosides were induced by Pandemrix vaccination. Altogether, there is currently no evidence for a causative contribution of polysorbate to Pandemrix®-induced narcolepsy.

It has to be noted that the content of polysorbate is markedly higher in vaccines where it is used as emulsifier in its oil-in-water adjuvant (1.175–4.85 mg/kg) than in many other vaccines where it is used as protein stabiliser (< 1 mg/kg).

4.2. Safety in children

Parenteral Vitamin solutions (E-Ferol tragedy)

The hepatotoxic potential of polysorbate gained notoriety after the E-ferol tragedy in the 1980s. E-ferol was an IV formulation of vitamin E marketed in December 1983 as antioxidant therapy for premature infants. The formulation contained a mixture of polysorbate 80 (9%) and polysorbate 20 (1%) as solubilising agents. After only 4 months of use, 38 infant deaths were reported in 11 states. While hepatic histology results from infants receiving E-ferol suggested a more cytotoxic than steatotic process, few investigations supported vitamin E content as the responsible culprit, thus leaving the mixture of polysorbate as suspect. Neonates whose deaths were attributed to E-Ferol administration received 100 to 548 mg/kg polysorbates per day (25 to 137 vitamin E U/kg/day), the duration of therapy was from 6 to 45 days, and the cumulative dose of polysorbates ranged from 1508 to 12000 mg/kg (377 to 3000 vitamin E U/kg) (Bove et al. 1985 [16]). A clear dose-response relationship was found: the attack rate (no. cases/no exposed %) increased at average daily doses > 20 U/kg/d E Ferol. This corresponds to an increased risk at a polysorbate dose of > 80 mg/kg/d (consisting of 72 mg PS 80 plus 8 mg PS 20) (Martone et al., 1986 [80]).

Polysorbates are still used as solubilising agents in marketed parenteral vitamin products in the US (M.V.I. Pediatric®: Multi-Vitamin for Infusion) that are administered to neonates. For example, the recommended daily doses of M.V.I. pediatrics® for infants < 1 kg are 30% (1.5 mL) of a single full dose (5 mL), and for infants weighing 1 to 3 kg 65% (3.25 mL) of a single full dose (5 mL) (M.V.I. pediatrics® label; 5 mL of reconstituted product provides 50 mg PS 80 and 0.8 mg PS 20 per 5 mL dose, in sum 50.8 mg polysorbates per 5 mL dose). This would yield an amount of 15.24 mg polysorbate per dose for infants < 1 kg, which is equal to 30.5 mg/kg/d for an infant weighing 500 g. The PS 80 dose for infants weighing 1–3 kg would be in the same range (11–33 mg/kg/d).

Amiodarone

Kicker et al. (2012 [65]) report of hepatotoxicity in a 34-week-old female infant with Down syndrome (2.6 kg) after amiodarone infusion. The reported patient received 270 mg of polysorbate 80 (103 mg/kg) in addition to 135 mg of IV amiodarone. The short-terminal half-life was discussed to explain the rapid resolution in hepatic injury after discontinuation of parenteral amiodarone.

Masi et al. (2009 [82]) published a case report of a 4-day-old newborn with cardiogenic shock after receiving by mistake a high “oral” loading dose (47 mg/kg) of amiodarone IV. Considering that the injectable product has a ratio of 2 mg polysorbate 80 for every 1 mg of amiodarone, the newborn

received ca. 100 mg/kg PS 80 IV with the amiodarone loading dose over a 30 min period (polysorbate infusion rate ca. 3.3 mg/kg/min): Measured plasma concentrations of amiodarone never reached toxic levels. Unfortunately, polysorbate levels were not measured. Of note, amiodarone IV solutions also contain the excipient benzyl alcohol (Masi et al., 2009 [81]).

In a study evaluating the efficacy and safety of intravenous amiodarone in infants and children, amiodarone loading doses at 1 mg/kg, 5 mg/kg and 10 mg/kg plus maintenance doses of 2, 5, and 10 mg/kg during the subsequent 47 hours were used (Saul et al., 2005 [106]). Adverse events such as hypotension, bradycardia and atrioventricular block appear more common for the two higher dose groups. The cumulative amiodarone dose levels correspond to polysorbate doses of 6 mg/kg, 20 mg/kg and 40 mg/kg administered over 48h (corresponding to a max rate of 0.2, 0.25 and 0.5 mg/kg/min, resp.). From the infusion schedule given it is deduced that the lowest dose group received about 4 mg/kg polysorbate on day 1 plus 2 mg/kg on day 2. The potential contribution of polysorbate to adverse events was not discussed by the authors.

Anidulafungin

Cohen-Wolkowicz et al. (2011 [23]) investigated the pharmacokinetics and safety of anidulafungin in infants and neonates. Anidulafungin was administered intravenously to 15 infants and neonates over 3 to 5 days as a loading dose of 3 mg/kg infused over 60 min on day 1 and daily maintenance dosages of 1.5 mg/kg infused over 60 min. Two anidulafungin presentations were used in the study. Infants received an intravenous alcohol (20%) based presentation whereas neonates received an alcohol-free, water for injection presentation. Only one of the two anidulafungin presentations is still approved commercially (Ecalta®). No drug related serious events were observed. Eight out of 15 subjects (53%) experienced at least one adverse event; most of these events were mild or moderate in severity (Table 2, see end of section 2.1). All but 2 adverse events were considered by the investigator to be unrelated to anidulafungin. The most reported non-serious adverse event was worsening hyperbilirubinemia.

From the content of polysorbate 80 in one of the products used (Eraxis® label) the exposure of PS 80 in this study was calculated to be 7.7 mg/kg/day on day 1 and 3.8 mg/kg/day over 3–5 days. As administrations were over 60 minutes these doses are equivalent to a rate of 0.064–0.13 mg/kg/min.

Parenteral nutrition

Total parenteral nutrition (TPN) is widely used. Although mechanical, septic, and metabolic complications are well known, hypersensitivity skin reactions are rare. The report of Levy and Dupuis (1990 [74]) describes a 16-year-old boy with Burkitt's lymphoma who developed an urticarial skin rash when treated with TPN and vitamins. The adverse skin reaction was probably caused by the inactive component of excipient, polysorbate.

5. Safety information relevant for the package leaflet

With respect to derivation for thresholds triggering a warning statement in the PI, a risk assessment for PS 80 (20) is warranted. For that purpose, the potential hazard and the corresponding doses/concentrations with regard to different administration routes are summarised.

Topical exposure

Delayed hypersensitivity reactions including contact dermatitis and contact urticarial have been reported after administration of creams containing polysorbates. Therefore, a warning on allergic reactions at threshold zero is proposed.

Oral exposure

In contrast to the parenteral route the oral application appears to be much less toxic. This is probably attributed to the very low oral bioavailability of the intact polysorbate: Only small amounts of polyoxyethylene sorbitans are absorbed intact. Enzymatic cleavage in the gut leads to the fact that after oral exposure almost solely the released fatty acid becomes systemically available (see chapter 3. Pharmacokinetics).

In its recent re-evaluation EFSA sums up that similar toxicokinetics would be expected for all polysorbates based on their similarities in structure and metabolic fate. The acute toxicity is very low. There is no concern regarding genotoxicity, carcinogenicity or developmental toxicity. From a limited number of studies, there is no indication of reproductive toxicity (EFSA, 2015 [32]). In the re-evaluation by EFSA in 2015 the acceptable daily intake (group ADI) for polysorbates as food additives (polysorbates 20, 80, 40, 60 and 65; E 432, E 433, E 434, E 435 and E 436, respectively) was set to 25 mg/kg body weight/day.

In view of the maximum oral doses of PS 80 or 20 in authorised medicinal products the oral exposure of PS 80 by oral formulations is estimated to be far below ADI.

Polysorbate 80 may increase gastrointestinal absorption of other drugs (see 3.3. Interactions). However, evidence for interactions between *oral* polysorbate containing medicinal products and the PK of concomitant oral medicinal products is currently too low to justify a general warning in SmPC/PIL for all oral medicinal products with polysorbates. This potential PK interaction may be provided in the SmPC/PIL on a case-by-case basis.

Future use of excipients as absorption enhancers may warrant reconsidering this type of warning. There are some fairly recent papers exploring this and a review that highlights that interactions with surfactants may be very complex (see 3.3. Interactions).

In conclusion, a threshold for oral administration of polysorbates as excipients is currently not considered meaningful.

Parenteral exposure

Hypersensitivity reactions including anaphylactic shock have been observed after IV as well as IM and SC administration of polysorbate containing medicinal products (see 4.1. Safety in adults). These include therapeutic proteins and vaccines with very low amounts of PS 20 and 80 (exposure as low as 0.01 mg/kg) with patients tested positive for PS 20 or 80 in skin tests. Although a non-IgE-mediated pseudoallergic mechanism is discussed, the case reports do not allow to define a threshold, and the possibility of IgE-mediated anaphylaxis cannot be excluded. Therefore, a respective warning of allergic/hypersensitivity reactions at threshold zero after parenteral exposure (all routes, not only IV) is considered justified.

In contrast to the oral route, after intravenous administration intact polysorbates enter the bloodstream. But even after IV administration polysorbates are rapidly cleared from plasma (half-life < 1h) probably due to hydrolysis by blood and liver lipases and esterases. The resulting fatty acid moieties are probably catabolised following the normal metabolic pathways of unsaturated fatty acids whereas the polyoxyethylene moiety is mainly excreted unchanged by the kidney (see chapters 2.3. Toxicokinetics and 3. Pharmacokinetics).

A potential risk for patients with HFI was identified due to the potential generation of free sorbitol by metabolism in the liver after IV application of polysorbates. Patients with HFI are advised to avoid polysorbate as a potential source of fructose (Gaughan et al. 2021 [47]). In one case study the

authors speculate that the hepatic and renal failure of the patient with HFI might have been caused by polysorbate 80 in the amiodarone infusion (Curran and Havill, 2002 [29]).

However, since to date neither PK nor clinical safety data give evidence that sorbitol generation by polysorbates constitutes a definite risk for such patients, the decision was taken not to include the strict warning for HFI patients as it is required for sorbitol-containing products for parenteral use at threshold zero (see Annex of the excipients guideline [3]).

It has been known for a long time that polysorbate 80 can enhance the uptake of drugs into the brain. Enhancement of brain uptake of other drugs is observed after IV doses of 3.2 mg/kg/d in mice and 20 mg/kg in rats (Azmin et al. 1985 [4], Calvo et al. 2001 [20]). In the case of coated particles, induction of endocytosis and/or transcytosis is favored as underlying uptake mechanism by polysorbate 80, but also membrane lipid solubilisation, opening of tight junctions or inactivation of the P-glycoprotein efflux pump could contribute to the effect. This ability is utilised in the coating of nanoparticles with PS 80 for drug delivery to the brain (Kreuter, 2013 [69]). However, simple addition of polysorbate 80 surfactant solution to doxorubicine was totally inefficient compared to coated nanoparticles. Nevertheless, the ability of polysorbates to affect distribution and uptake constitutes a potential interaction with drug substances, which should be taken into account during drug development and benefit-risk evaluation of current and new parenteral products containing polysorbates.

Evidence for a cardiotoxic/torsadogenic potential of polysorbates comes from in vitro data on hERG current inhibition as well as from preclinical data showing an increase in effective refractory period (ERP) in guinea-pig cardiac preparations and in vivo in dogs (see chapter 2.1.3. Cardiovascular effects). Block of I_{Kr} (hERG channels) by polysorbate 80 might explain the observation of increased ventricular ERP in the dog after IV administration of 20 mg/kg polysorbate 80 (Torres-Arraut et al., 1984 [125]). Some data indicate that polysorbate 80 is a "multi-ion channel blocker" in the heart inducing cardiac electrophysiological effects not only via block of I_{Kr} . The electrophysiological studies performed in guinea-pig cardiac preparations (Batey et al. 1997 [9]) and in-vivo in dogs (Torres-Arnault et al. 1984 [125]) were published several years ago and, therefore, the methods do not seem to be completely state-of-the-art of the year 2017.

Precautionary safety limits with regard to cardiotoxicity could be approximated from in vitro IC₅₀ of PS 80 for inhibition of hERG currents which is reported to be 0.2 mg/ml (0.02%; IC₅₀ of PS 20 similar, see 2.1.3. Cardiovascular effects). According to Redfern et al. 2003, a 30-fold margin between free therapeutic plasma concentrations and IC₅₀ values for block of hERG currents appears to be a line of demarcation between the majority of drugs associated with Torsades de Pointes (TdP) arrhythmias and those which are not. Division of IC₅₀ by 30 results in a plasma concentration of 0.007 mg/ml (0.0007%) polysorbate 80 which should not be exceeded in vivo. Following this, an IV bolus dose of 0.35 and 0.7 mg/kg for adults and infants, respectively, could be regarded as safe, because it will not exceed this initial plasma concentration. This derivation is valid for bolus injections only. When infusions are administered more slowly (e.g. over 1h), much lower polysorbate levels were measured at the end of infusion than expected from calculation for a bolus dose: humans receiving very high polysorbate 80 doses of 3–4.5 g (50–75 mg/kg) via Taxotere® infusions (1h) yielded end of infusion plasma concentrations of about 0.01% (Webster et al. 1997 [134]). These are tenfold lower than expected from a 3 g bolus dose distributing in 3 L Plasma (0.1%). This is in line with the rapid plasma clearance of PS 80 observed in adults (7.7 L/h, see chapter 3.1.2. Intramuscular administration).

For cases of administration as continuous infusion, alternatively, an infusion rate (R_{inf}) can be estimated which would not exceed a steady state concentration in plasma (C_{ss}) of 0.007 mg/ml (using equation $R_{inf} = CL \cdot C_{ss}$). From this a "safe" continuous infusion rate of < 0.015 mg/kg/min for an

adult (60 kg) is calculated. This corresponds to a safe cumulative dose of 21 mg/kg/d when given as continuous infusion. With respect to slower metabolism of polysorbates, i.e. lower CL/kg, expected in infants compared to adults (see 3.2. Pharmacokinetics in children), an additional safety factor for infants could be discussed.

In vivo bolus doses ≥ 10 mg/kg of PS 80 alone lead to depression of the cardiac conduction (prolongation of the sinus node recovery time, depressed AV-nodal function and increased atrial effective refractory period (ERP)) and hypotension in dogs (Torres-Arraut et al., 1984 [125], Masini et al., 1985 [83]). The authors concluded that polysorbate 80 is a potent depressant of the cardiac conduction system in the dog and its electro physiologic effects are similar to those of amiodarone. Infusion rate was 2 mg/kg/min (over 5 min) in the Masini study (Masini et al., 1985 [83]) when hypotension/Histamine release was observed. Recent preclinical data in dogs (Cushing et al., 2009 [30]) unequivocally confirmed that the hypotensive effects of commercial amiodarone IV result from the co-solvents (PS 80 and benzyl alcohol) in the formulation. Polysorbate 80 exposure was even lower in this study (4.3 mg/kg, given as a bolus or as 10 min infusion; rate: 0.43 mg/kg/min). It cannot be excluded that benzyl alcohol also contributed to the hypotensive response due to its negative inotropic effects (see Cushing et al., 2009 [30]).

There is no evidence so far for depression of cardiac conduction from clinical data in humans. Some authors speculated that the reported cases of death of several neonates after parenteral administration of a vitamin-E preparation (E-ferol) containing 9% polysorbate 80 and 1% polysorbate 20 might be due to block of I_{Kr} by polysorbate 80 (Pesce and McKean 1989). However, cases occurred at much higher cumulative dose levels (> 80 mg/kg/day) where hepatotoxicity is predominant (see below).

In human adults a significant hemodynamic effect (short duration vasoplegia, left ventricular systolic pressure decreased) was observed after amiodarone IV injection (Cordarone®) compared to a formulation without polysorbate and benzyl alcohol (Munoz et al. 1988 [88]). Polysorbate dose (10 mg/kg) was given as a 3 min bolus (rate for PS 80: 3.33 mg/kg/min). Effects occurred immediately during the injection and were short-lived. The authors concluded that "the risk of severe hypotension after IV Cordarone® can be largely avoided by using a slower rate of infusion". (Current dose recommendation for the loading rapid infusion of IV Cordarone (US label): 150 mg amiodarone over the first 10 minutes, which equals to a PS 80 dose of 4.3 mg/kg at a rate of 0.43 mg/kg/min). This is in contrast to dosage recommendations for amiodarone products in the EU (e.g., Amiodaron-ratiopharm®, DE) which include a bolus injection of 5 mg/kg (corresponding to 10 mg/kg PS 80) over ≥ 3 min).

Of note, after an accidental high total PS 80 dose (100 mg/kg) infused at a similar rate of 3.3 mg/kg/min (via amiodarone IV), a cardiogenic shock was observed in a 4-day-old newborn (Masi et al., 2009 [82]).

Maximum plasma concentration of polysorbate in humans after IV injection of a bolus injection of 3 mg/kg is roughly estimated to be 0.03–0.07 mg/ml (plasma volume of 60 ml/kg assumed). This is in the range of the concentrations needed to elicit cytotoxic effects of PS 80 on cells in vitro (0.05 mg/ml) and below IC₅₀ at hERG channels (0.2 mg/ml). These considerations might add mechanistically support for a 3 mg/kg bolus dose as a plausible threshold limit for a warning.

Support for the safe short term exposure of PS 80 < 3 mg/kg per day in infants and neonates comes from a small PK and safety study in infants and neonates (Cohen-Wolkowicz et al., 2011 [23]): Anidulafungin has been given to neonates (8) and infants (9) at 3 mg/kg/day loading dose (day 1) and subsequently at 1.5 mg/kg/day for 3–5 days (according to the posology of Ecalta®). These doses correspond to PS 80 doses of 7.7 mg/kg/day on Day 1 and 3.8 mg/kg/day over 3–5 days. Polysorbate

infusion rates (over 60 min) were 0.064–0.13 mg/kg/min. There were no product related serious adverse effects. From this a short-term exposure limit for PS 80 of ≤ 8 mg/kg/d given at an infusion rate of < 0.15 mg/kg/min could cautiously be deduced for infants and neonates > 1 months of age. (Of note, non-serious events included elevation of liver enzymes).

MVI paediatric is used in the US and has been used over a long period also in Europe even though not authorised without any apparent safety issues. The cumulative dose of polysorbate 80 in MVI paediatric is very high, maximally 32.5 mg/kg/day in a 1 kg neonate/infant given as an infusion over 24 hours. The safe use of this product is apparently contradictory to the high cumulative daily dose. However, it is given as a continuous infusion, and at the maximal dose the hourly rate is 1.35 mg/kg/h for a 1 kg neonate. This equals to a rate of 0.023 mg/kg/min, which is markedly below the injection rate of PS 80 via amiodarone bolus injection in adults leading to hemodynamic effects (3.33 mg/kg/min) and below the rate of 0.13 mg/kg/min considered as safe in neonates and infants from the Cohen-study (see above). This could well explain why this product is safely used and further supports that rate of injection (peak exposure, C_{max}) might be more important than the cumulative dose, at least for cardiovascular/cardio toxic effects.

In the Saul study in infants and children (Saul et al., 2005 [106]) even the lowest dose of 4 mg/kg on day 1 (6 mg/kg over 48 h, max rate: 0.2 mg/kg/min) did have some effects on blood pressure and heart rate, and higher doses (20 and 40 mg/kg over 48 h) more often. This study supports the assumptions of rate and dose both contributing and also a threshold of 3 mg/kg/day for risk of effects on blood pressure and heart rate.

In summary, the hemodynamic (and perhaps also the potential cardiotoxic) effects appear to be rather related to the infusion rate (peak exposure) than to the total dose (cumulative exposure). An infusion rate of 0.015 mg/kg/min is theoretically considered as safe (from IC₅₀ of hERG inhibition), a 10-fold higher rate of 0.06–0.13 mg/kg/min (up to a total dose of 4–7.7 mg/kg/day) has been proven to be safe in infants and neonates (anidulafungin, Ecalta®, Cohen-Study, see above). On the other hand, short infusions at rates of 0.43–2 mg/kg/min lead to hypotension/histamine release in dogs (Cushing et al., 2009 [30]; Masini et al., 1985 [83]), and a rate of 3.3 mg/kg/min (for 3 min) was shown to be associated with hemodynamic effects in adults and (for 30 min) with a cardiogenic shock in a 4 day old new-born (Munoz et al., 1988 [88]; Masi et al., 2009 [82]).

Cushing et al. (2009 [30]) reviewed that polysorbate 80 alone (at doses between 2 and 20 mg/kg) or in combination with benzyl alcohol produced profound reductions in arterial blood pressure in dogs and other animal models. However, a proof of the alleged effect at 2 mg/kg could not be found in the literature cited. The lowest effect levels leading to hypotension in dogs identified were 0.43 mg/kg/min (10 min infusion at this rate) or 4.3 mg/kg bolus dose (Cushing et al. 2009 [30]). This is supported by Varma et al. 1985 [130] who demonstrated a blood pressure lowering effect in dogs after the lowest applied bolus IV dose of 5 mg/kg PS 80. The lowest effective dose of 4.3 mg/kg in dogs is considered equal to a human dose of about 3 mg/kg (by application of an allometric factor of 1.4 for dogs). Clinical data (Saul et al. 2005 [106]) also support this threshold.

In conclusion, from the totality of preclinical and clinical data a threshold dose of 3 mg/kg/day is proposed. This is a cumulative dose which in worst case is administered as a bolus injection. Values equal to or above should trigger a warning regarding cardiovascular effects (hypotension/cardiac depression). Secondly, a general recommendation for lowering the infusion rate as much as feasible is suggested for consideration in the comments column.

In addition, it is concluded that further (pre-clinical and) clinical electrophysiological studies are warranted to investigate the torsadogenic potential of polysorbate 80 in detail (according to the ICH

S7B and E14 Guidelines, e.g. measurement of action potential parameters in isolated cardiac preparations, measurement of proarrhythmic effects in isolated cardiac preparations, evaluation of polysorbate 80 regarding the TRIAD concept; a “thorough QT study” in humans according to the ICH E14 Guideline). Due to the potential effect on hERG channels by polysorbates synergistic effects might occur after administration of polysorbate 80 in combination with other hERG channel blockers. Therefore, a warning on the risk of concomitant use of medications that prolong the QT/QTc interval should be considered for the SmPC/PIL of all products containing polysorbates above this threshold.

Furthermore, a parenteral threshold triggering a warning of hepatotoxicity is deemed necessary. From the E-Ferol tragedy a cumulative dose limit of 80 mg/kg/day (corresponding to an infusion rate of 0.055 mg/kg/min when applied as continuous infusion) for severe hepatotoxicity in premature infants was deduced, because no cases (defined as illness following the clinical diagnosis of ascites or occurrence of at least two clinical laboratory abnormalities) occurred at doses that were below 20 units of alfa-tocopherol, which corresponded to doses below 72 mg/kg/day and 8 mg/kg/day for PS 80 and PS 20, correspondingly (Martone et al., 1986 [80]). The toxicities reported were after infusions administered continuously over 24 hours, and they occurred after a longer time of infusion or even after administration has been stopped. This suggests that cumulative doses rather than short term peak exposure levels appear to be relevant for hepatotoxicity. But it is still unclear whether toxicities might be related to peak concentrations or cumulative concentrations, and whether the toxicity might depend on rate of administration.

Case reports in adults at exposures below 80 mg/kg/d may indicate an earlier onset of signs of hepatotoxicity: 35–40 mg/kg were calculated as the cumulative polysorbate dose within 24 h identified in case reports of hepatotoxicity in adults after Amiodarone IV, e.g., showing abrupt elevation of liver enzymes (see 4.1. Safety in adults). Such case reports are confounded by the fact that amiodarone itself is a hepatotoxic agent. However, the observation that oral amiodarone administrations in such patients do not result in additional liver toxicity supports the association with the intravenous exposure of the excipient.

In conclusion, a threshold of 35 mg/kg/d for all age groups is suggested to trigger a warning for elevation of liver enzymes. This threshold would be supported by the fact that it is above the exposure expected from MVI paediatrics (maximally 33 mg/kg/day in a 1 kg neonate/infant) which has been used over a long period in the US (as well as in Europe) without any apparent safety issues. However, it would be below the expected polysorbate exposure from Taxotere® (55 mg/kg). As with amiodarone, docetaxel itself is potentially hepatotoxic, it is thus not possible to attribute cases of severe hepatotoxicity after docetaxel solely to polysorbate 80. However, in recent studies comparing Taxotere® with new polysorbate-free formulations, hepatotoxicity was not among the differences identified (Tagawa et al., 2017 [118]).

Recently, Kriegel et al. (2019 [70]) proposed the application of a progressive paediatric safety factor (PPSF) on the maximum acceptable excipient dose (MAED) in adults in order to estimate the amount of excipient in formulations that will avoid adverse events. By use of this PPSF a MAED of 1.4 mg PS 80 for neonates was calculated from the MAED of PS 80 in adults (300 mg IV, i.e. the lowest PS 80 amount in authorised amiodarone formulations, which is considered as being the tolerable maximum). This “maximum tolerated” dose corresponds to a body-weight-related dose for neonates of ≤ 1 mg/kg. This is in line with the parenteral threshold of 3 mg/kg/d triggering the first warning statement aside from hypersensitivity.

Impact of the proposals on the labelling of parenteral medicinal products in EU:

Small molecules

- Anidulafungin (Ecalta®): polysorbate dose: 8.5 mg/kg/d (continuous infusion; above zero and 8 mg/kg/day cumulative dose threshold: labelling of content/allergy and cardiovascular effects, e.g., hypotension);
- Amiodarone (e.g., Amiodaron-ratiopharm®, DE): polysorbate exposure from bolus injection of 5 mg amiodarone/kg: 10 mg/kg (over 3 to 20 min) (above first and second threshold: labelling of content/allergy and "cardiovascular effects, e.g., hypotension");
- Docetaxel (Taxotere®): polysorbate exposure by single dose: 55 mg/kg (above all thresholds; labelling: content/allergy, cardiovascular effects, liver toxicity).

Proteins

Polysorbate exposure by administration of therapeutic proteins is low (≤ 1.2 mg/kg). The bolus dose (SC or IV) is below all thresholds apart from zero. Even if administered as slow infusion, the infusion rates are expected to be below 0.15 mg/kg/min which could be regarded as safe even in neonates and infants. For example, Herceptin® loading dose (including polysorbate exposure dose of 1 mg/kg) is given as a 90 min infusion. Thus, infusion rate is calculated as 0.01 mg/kg/min.

Vaccines

The highest polysorbate content is 0.75 mg/vaccine dose which is equivalent to $0.75/60 \text{ kg} = 0.0125 \text{ mg/kg}$ in adults (worst case: 16 year-old/30 kg $\rightarrow 0.025 \text{ mg/kg}$). The highest content in a vaccine authorised for use in infants/neonates is 0.1 mg/dose which corresponds to 0.03 mg/kg in a 3 kg infant. A higher polysorbate content per vaccine dose is observed in vaccines containing oil adjuvants. The seasonal influenza vaccine Fludac is authorised for elderly adults (> 65 years) only. The polysorbate dose of 1.175 mg/vaccine dose is equivalent to about 0.02 mg/kg for a 60 kg person. All derived bolus IM/SC dose levels of PS 80 would be below the first threshold apart from zero (3 mg/kg/d).

This is considered appropriate as it is in line with the absence of any signal of cardiotoxicity/hepatotoxicity after vaccine exposure from epidemiology or pharmacovigilance.

Further support is derived from the following considerations: considering a worst case scenario of complete systemic availability of the total dose of PS 80 administered with one vaccine dose (max. dose assumed: 0.75 mg), maximum plasma concentrations of intact PS 80 of about 0.25 $\mu\text{g/ml}$ in adults (70 kg) or 1.5 $\mu\text{g/ml}$ in children (10 kg) or 3 $\mu\text{g/ml}$ in neonates (3 kg) can be conservatively estimated (assuming intravenous injection of the total dose into plasma volumes of 3 l, 0.5 l and 0.25 l, respectively). These concentrations are not expected to have any effect, they are even below the precautionary limit of 0.0007 mg/ml (7 $\mu\text{g/ml}$) with regard to potential QT prolongation (see above) and far below concentrations eliciting membrane/cytotoxic effects on cells in vitro (CC50 for cytotoxicity in vitro: 48 $\mu\text{g/ml}$; see Table 1 at the end of section 2.1).

For the previous pandemic vaccine Pandemrix® (market authorisation expired 2015 in EU) which contained a very high polysorbate content in its emulsion adjuvant (4.85 mg/dose), the estimated worst case maximum plasma concentrations after one vaccine dose (assuming sudden 100% bioavailability) would calculate as 1.6, 9.7, and 19.4 $\mu\text{g/ml}$ in adults, children, and neonates, respectively.

With respect to vaccine-induced narcolepsy, it is noted that there is scientific evidence indicating that a special antigen ingredient is more likely to be causative for the development of narcolepsy than the polysorbate 80 containing adjuvant (Ahmed and Steinman, 2017 [1]).

References – Bibliography

1. Ahmed SS and Steinman L. Mechanistic insights into influenza vaccine-associated narcolepsy. *Human vaccine & Immunotherapeutics* 2016, 12:3196-3201 Anonymous. Final Report on the Safety Assessment of Polysorbates 20, 21, 40, 60, 61, 65, 80, 81, and 85. 1984, *Journal of the American College of Toxicology* 3 (5), p. 1–82.
2. Al-Ali Ahmed A. Abdulhusein, Nielsen Rasmus Blaaholm, Steffansen Bente, Holm René, Nielsen Carsten Uhd. Nonionic surfactants modulate the transport activity of ATP-binding cassette (ABC) transporters and solute carriers (SLC): Relevance to oral drug absorption. *International Journal of Pharmaceutics*. Volume 566, 20 July 2019, Pages 410–433.
3. Annex of the European Commission guideline 'Excipients in the labelling and package leaflet of medicinal products for human use' (EMA/CHMP/302620/2017 rev. 2).
4. Azmin MN, Stuart JF, Florence AT. The distribution and elimination of methotrexate in mouse blood and brain after concurrent administration of polysorbate 80. *Cancer Chemother Pharmacol*. 1985; 14(3), p. 238–42.
5. Badiu I, Geuna M, Heffler E, Rolla G. Hypersensitivity reaction to human papillomavirus vaccine due to polysorbate 80. *BMJ Case Rep*. 2012 May 8; 2012:bcr0220125797. doi: 10.1136/bcr.02.2012.5797. PMID: 22605841; PMCID: PMC3351639.
6. Baker SD, Li J, ten Tije AJ, Figg WD, Graveland W, Verweij J, Sparreboom A. Relationship of systemic exposure to unbound docetaxel and neutropenia. *Clin Pharmacol Ther*. 2005 Jan; 77(1), p. 43–53.
7. Barnett JB. and Bryant RL., Adjuvant and immunosuppressive effects of retinol and Tween 80 on IgG production in mice. *Int Arch Allergy Appl Immunol*. 1980; 63(2), p. 145–52.
8. Barnett JB. Immunosuppressive effects of tween 80 on mice. *Int Arch Allergy Appl Immunol*. 1981;66(2), p. 229–32.
9. Batey AJ, Lightbown ID, Lambert JP, Edwards G, Coker SJ. Comparison of the acute cardiotoxicity of the antimalarial drug halofantrine in vitro and in vivo in anaesthetized guinea-pigs. *Br. J. Pharmacol*. 1997, 122:563–569.
10. Bergmann KC, Maurer M, Church MK, Zuberbier T. Anaphylaxis to Mepolizumab and Omalizumab in a Single Patient: Is Polysorbate the Culprit? *J Investig Allergol Clin Immunol*. 2020 Aug; 30(4):285–287. doi: 10.18176/jiaci.0492. PMID: 32723701.
11. BIBRA Toxicology International. Toxicity profile, polysorbate 80. 1992 - Available from: <http://legacy.library.ucsf.edu/tid/wmw45d00/pdf>.
12. Black S, Della Cioppa G, Malfroot A, Nacci P, Nicolay U, Pellegrini M, Sokal E, Vertruyen A Safety of MF59-adjuvanted versus non-adjuvanted influenza vaccines in children and adolescents: An integrated analysis. *Vaccine* 2010 28 7331–6.
13. Blasi P, Schoubben A, Romano GV, Giovagnoli S, Di Michele A, Ricci M. Lipid nanoparticles for brain targeting II. Technological characterization. *Colloids Surf B Biointerfaces*. 2013 Oct 1;110:130–7.
14. Block SL, Ruiz-Palacios GM, Guerrero L, Beygo J, Sales V, Holmes SJ, Dose-range study of MF59-adjuvanted versus nonadjuvanted monovalent A/H1N1 pandemic influenza vaccine in six-to less than thirty-six-month-old children. *Pediatric Infectious Disease Journal*, 2012 Jul 31(7) 92–8.

15. Boccia R, Geller RB, Clendeninn N, Ottoboni T. Hypersensitivity and infusion-site adverse events with intravenous fosaprepitant after anthracycline-containing chemotherapy: a retrospective study. *Future Oncol.* 2019 Jan;15(3):297–303. doi: 10.2217/fon-2018-0662. Epub 2018 Oct 10. PMID: 30301373.
16. Bove KE, Kosmetatos N, Wedig KE, Frank DJ, Whitlatch S, Saldivar V, Haas J, Bodenstein C, Balistreri WF. Vasculopathic hepatotoxicity associated with E-Ferol syndrome in low-birth-weight infants. *JAMA.* 1985 Nov 1; 254(17), p. 2422–30.
17. Brubaker CM, Taylor DH, Bull RJ Effect of Tween 80 on exploratory behavior and locomotor activity in rats. *Life Sci.* 1982 Jun 7; 30(23), p. 1965–71.
18. Bryant, R.L., and Barnett, J.B. (1979). Adjuvant properties of retinol on IgE production in mice. *Int. Arch. Allergy Appl. Immunol.* 59(1), p. 69–74.
19. Burlando M, Herzum A, Cozzani E, Parodi A. Acute urticarial rash after COVID-19 vaccination containing Polysorbate 80. *Clin Exp Vaccine Res.* 2021 Sep; 10(3):298–300. doi: 10.7774/cevr.2021.10.3.298. Epub 2021 Sep 30. PMID: 34703816; PMCID: PMC8511582.
20. Calvo P, Gouritin B, Chacun H, Desmaële D, D'Angelo J, Noel JP, Georgin D, Fattal E, Andreux JP, Couvreur P. Long-circulating PEGylated polycyanoacrylate nanoparticles as new drug carrier for brain delivery. *Pharm Res.* Aug 2001; 18(8), p. 1157–66.
21. Chen CC, Wu CC. Acute Hepatotoxicity of Intravenous Amiodarone: Case Report and Review of the Literature. *Am J Ther.* Jan-Feb 2016; 23(1):e260-3.
22. Christiansen A, Backensfeld T, Denner K. Weitschies W, Effect of non-ionic surfactants on cytochrome P450-mediated metabolism *in vitro*. *Eur J Pharm Biopharm* 2011 May;78(1):166–72. doi: 10.1016/j.ejpb.2010.12.033. Epub 2011 Jan 8. PMID: 21220010.
23. Cohen-Wolkowicz et al., Safety and Pharmacokinetics of Multiple-Dose Anidulafungin in Infants and Neonates. *Clin Pharmacol Ther.* May 2011, 89(5), p. 702–707.
24. Commission Regulation (EU) No 1130/2011 of 11 November 2011 amending Annex III to Regulation (EC) No 1333/2008 of the European Parliament and of the Council on food additives by establishing a Union list of food additives approved for use in food additives, food enzymes, food flavourings and nutrients.
25. Cosmetic Ingredient Review (CIR) Expert panel, Final Report on the Safety Assessment of Polysorbates 20, 21, 40, 60, 61, 65, 80, 81, and 85. *J Am Coll Toxicol.* 3, 1984.
26. Coors EA, Seybold H, Merk HF, Mahler V. Polysorbate 80 in medical products and nonimmunologic anaphylactoid reactions. *Ann Allergy Asthma Immunol.* 2005 Dec; 95(6), p. 593–9.
27. Crispens C. G., Jr., Sorenson J. R. Treatment of reticulum cell sarcoma in SJL/J mice with Tween 80. *Anticancer Res.*, 1988(8), p. 1341–1343.
28. Cummings J, Forrest GJ, Cunningham D, Gilchrist NL, Soukop M. Influence of polysorbate 80 (Tween 80) and etoposide (VP-16-213) on the pharmacokinetics and urinary excretion of adriamycin and its metabolites in cancer patients. *Cancer Chemother Pharmacol.* 1986; 17(1), p. 80–4.
29. Curran BJ, Havill JH. Hepatic and renal failure associated with amiodarone infusion in a patient with hereditary fructose intolerance. *Crit Care Resusc.* 2002 Jun; 4(2):112–5. PMID: 16573414.

30. Cushing DJ, Kowey PR, Cooper WD, Massey BW, Gralinski MR, Lipicky RJ. PM101: a cyclodextrin-based intravenous formulation of amiodarone devoid of adverse hemodynamic effects. *Eur J Pharmacol.* 2009 Apr 1; 607(1-3):167–72. doi: 10.1016/j.ejphar.2009.02.009. Epub 2009 Feb 14. PMID: 19232340.
31. Drori S., Eytan G. D., Assaraf Y. G. Potentiation of anticancer-drug cytotoxicity by multidrug-resistance chemosensitizers involves alterations in membrane fluidity leading to increased membrane permeability. *Eur. J. Biochem.*, 1995(228), p. 1020–1029.
32. EFSA ANS Panel (EFSA Panel on Food Additives and Nutrient Sources Added to Food), 2015. Scientific Opinion on the re-evaluation of polyoxyethylene sorbitan monolaurate (E 432), polyoxyethylene sorbitan monooleate (E 433), polyoxyethylene sorbitan monopalmitate (E 434), polyoxyethylene sorbitan monostearate (E 435) and polyoxyethylene sorbitan tristearate (E 436) as food additives. *EFSA Journal* 2015; 13(7):4152, 74 pp.
33. Ellis AG, Crinis NA, Webster LK. Inhibition of etoposide elimination in the isolated perfused rat liver by Cremophor EL and Tween 80. *Cancer Chemother Pharmacol.* 1996; 38(1), p. 81–7.
34. Ema M, Hara H, Matsumoto M, Hirata-Koizumi M, Hirose A and Kamata E. Evaluation of developmental neurotoxicity of polysorbate 80 in rats. *Reproductive Toxicology* 2008; 25(1), p. 89–99.
35. Engels FK, Mathot RA, Verweij J. Alternative drug formulations of docetaxel: a review. [Anticancer Drugs](#). Feb 2007, 18(2), p.95–103.
36. Enright BP, McIntyre BS, Thackaberry EA, Treinen KA and Kopytek SJ. Assessment of hydroxypropyl methylcellulose, propylene glycol, polysorbate 80, and hydroxypropyl- β -cyclodextrin for use in developmental and reproductive toxicology studies. *Birth Defects Res B Dev Reprod Toxicol.* 2010 Dec; 89(6), p. 504–16.
37. Eraxis® label:
https://www.accessdata.fda.gov/drugsatfda_docs/nda/2006/21948s000_Eraxis_PrntLbl.pdf
38. Estella-Hermoso de Mendoza A, Pr  at V, Mollinedo F, Blanco-Prieto MJ In vitro and in vivo efficacy of edelfosine-loaded lipid nanoparticles against glioma. *J Control Release.* 2011 Dec 20; 156(3), p. 421–6.
39. European Commission guideline on 'Excipients in the label and package leaflet of medicinal products for human use, March 2018. Rev. 2, Notice to Applicants, Volume 2C Guidelines, SANTE-2017-11668.
40. European Pharmacopoeia 8.1, 2014.
41. Farkas WR, Lorch V, Conover WR 3rd, al-Ansari HM, Abney LK, Painter PC 3rd, Reyniers JP, Congdon CC Polysorbate toxicity in neonatal rats and mice. *Pharmacol Toxicol.* Feb 1991, 68(2), p. 154–6.
42. Fonseca P, Dias A, Gon  alves H, Albuquerque A, Gama V. Acute hepatitis after amiodarone infusion. *World J Clin Cases.* Oct 2015, 3(10), p. 900–3.
43. Food Safety Commission. Evaluation report of food Additives. Polysorbates (Polysorbates 20, 60, 65 and 80). 2007; Original: Japanese- Available from:
https://www.fsc.go.jp/english/evaluationreports/foodadditive/polysorbate_report.pdf
44. Friemel A, Zunkler BJ. Interactions at human ether-a-go-go-related gene channels. *Toxicol Sci* 2010; 114: 346–355.

45. Gajdova M, Jakubovsky J and Valky J: Delayed effects of neonatal exposure to Tween 80 on female reproductive organs in rats; *Food Chem. Toxicol.*, 31 (1993), p. 183–190.
46. Garidel, P., Claudia Hoffmann, C., Alfred Blume, A., 'A Thermodynamic Analysis of the Binding Interaction Between Polysorbate 20 and 80 with Human Serum Albumins and Immunoglobulins: A Contribution to Understand Colloidal Protein Stabilisation', *Biophysical Chemistry*, Elsevier, 2009, 143 (1–2), p. 70.
47. Gaughan S, Ayres L, Baker PR II. Hereditary Fructose Intolerance. 2015 Dec 17 [Updated 2021 Feb 18]. In: Adam MP, Mirzaa GM, Pagon RA, et al., editors. *GeneReviews®* [Internet]. Seattle (WA): University of Washington, Seattle; 1993–2023. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK333439/>
48. Giannattasio F, Salvio A, Varriale M, Picciotto FP, Di Costanzo GG, Visconti M. Three cases of severe acute hepatitis after parenteral administration of amiodarone: the active ingredient is not the only agent responsible for hepatotoxicity. *Ann Ital Med Int.* 2002 Jul-Sep;17(3):180–4.
49. Gonzalez-Mira E1, Egea MA, Garcia ML, Souto EB. Design and ocular tolerance of flurbiprofen loaded ultrasound-engineered NLC. *Colloids Surf B Biointerfaces.* 2010 Dec 1; 81(2), p. 412–21.
50. Göppert TM and Müller RH, Polysorbate-stabilized solid lipid nanoparticles as colloidal carriers for intravenous targeting of drugs to the brain: Comparison of plasma protein adsorption patterns, *Journal of Drug Targeting*, 2005, 13:3, 179–187, DOI: 10.1080/10611860500071292
51. Gulyaev AE, Gelperina SE, Skidan IN, Antropov AS, Kivman GY, Kreuter J. Significant transport of doxorubicin into the brain with polysorbate 80-coated nanoparticles. *Pharm Res.* 1999 Oct; 16(10), p. 1564–9.
52. Hale TW, Rais-Bahrami K, Montgomery DL, Harkey C and Habersang RW. Vitamin E toxicity in neonatal piglets. *J Toxicol Clin Toxicol.* 1995; 33(2), p. 123–30.
53. *Handbook of Excipients*, 7th edition 2012 (eds.: Rowe RC, Sheskey PJ, Cook WG, Fenton ME) access on 12.02.2014.
54. Hennenfent KL and Govindan R. Novel formulations of taxanes: a review. *Old wine in a new bottle? Annals of Oncology* 2005, 17, p. 735–749.
55. Himmel HM. Suitability of commonly used excipients for electrophysiological in-vitro safety pharmacology assessment of effects on hERG potassium current and on rabbit Purkinje fiber action potential. *Journal of Pharmacological and Toxicological Methods* 2007, 56, p. 145–158.
56. Hines RN, Simpson PM, McCarver DG. Age-Dependent Human Hepatic Carboxylesterase 1 (CES1) and Carboxylesterase 2 (CES2) Postnatal Ontogeny. *Drug Metab Dispos.* 2016 Jul;44(7):959–66. doi: 10.1124/dmd.115.068957. Epub 2016 Jan 29. PMID: 26825642.
57. Ieven T, Van Weyenbergh T, Vandebotermiet M, Devolder D, Breynaert C, Schrijvers R. Tolerability of polysorbate 80-containing COVID-19 vaccines in confirmed polyethylene glycol-allergic patients. *J Allergy Clin Immunol Pract.* 2021 Dec; 9(12):4470–4472.e1. doi: 10.1016/j.jaip.2021.09.039. Epub 2021 Oct 6. PMID: 34626857; PMCID: PMC8492825.
58. Information for the package leaflet regarding fructose and sorbitol used as excipients in medicinal products for human use (EMA/CHMP/460886/2014), 2017.

59. Jacob L, Leib R, Ollila HM, Bonvalet M, Adams CM, Mignot E. Comparison of Pandemrix and Arepanrix, two pH1N1 AS03-adjuvanted vaccines differentially associated with narcolepsy development. *Brain Behav Immun*. Jul 2015(47), p. 44–57.
60. Jelinek A, In-vitro-Toxizität grenzflächenaktiver Substanzen. Dissertation Martin-Luther-Universität Hall-Wittenberg 2001. <http://sundoc.bibliothek.uni-halle.de/diss-online/01/01H171/index.htm>).
61. Jennifer S. Kicker, MD, Julie A. Haizlip, MD, and Marcia L. Buck: Hepatotoxicity After Continuous Amiodarone Infusion in a Postoperative Cardiac Infant. *J Pediatr Pharmacol Ther*. 2012 Apr-Jun; 17(2), p. 189–195.
62. Joint FAO/WHO (Food and Agriculture Organization/World Health organisation) Expert Committee on Food additives (JECFA): Toxicological Evaluation of some food additives including anticaking agents, antimicrobials, antioxidants, emulsifiers and thickening agents, WHO Food Additive Series No 5. 1974.
63. Kerwin BA, Polysorbates 20 and 80 used in the formulation of protein biotherapeutics: Structure and degradation pathways. *J Pharm Sci* 2008; 97(8):2924–2935.
64. Kevin E. Bove, MD; Niki Kosmetatos, MD; Kathryn E. Wedig, MD; Donald J. Frank, MD; Stephen Whitlatch, MD; Victor Saldivar, MD; Joel Haas, MD; Carl Bodenstein, MD; William F. Balistreri, MD: *Vasculopathic Hepatotoxicity Associated With E-Ferol Syndrome in Low-Birth-Weight Infants*. *JAMA*. 1985;254(17):2422–2430.
65. Kicker JS et al. Hepatotoxicity After Continuous Amiodarone Infusion in a Postoperative Cardiac Infant. *J Pediatr Pharmacol Ther*. 2012 Apr-Jun; 17(2), p. 189–195.
66. Kobayashi H, Nishimura T, Okumura K, Muranishi S, Sezaki H. Effect of polysorbates on absorption rates of water-soluble, micelle-free drugs administered intramuscularly in the rat. *J Pharm Sci*. Apr. 1974, 63(4), p. 580–4.
67. Kobayashi H, Peng TC, Kawamura R, Muranishi S, and Sezaki H, Mechanism of the inhibitory effect of polysorbate 80 on intramuscular absorption of drugs. *Chem. Pharm. Bull. (Tokyo)* 1977, 25(4), p. 569–74.
68. Koffie et al. Nanoparticles enhance brain delivery of blood–brain barrier-impermeable probes for in vivo optical and magnetic resonance imaging. *Proc natl Acad Sci USA* 2011 108(46), p. 18837–18842.
69. Kreuter J, Mechanism of polymeric nanoparticle-based drug transport across the blood brain barrier (BBB). *Journal of Microencapsulation* 2013; 30:49–54.
70. Kriegel C, Festag M, Kishore RSK, Roethlisberger D, Schmitt G. Pediatric Safety of Polysorbates in Drug Formulations. *Children (Basel)*. 2019 Dec 20; 7(1):1. doi: 10.3390/children7010001. PMID: 31877624; PMCID: PMC7022221.
71. Lam XM, Lai WG, Chan EK, Ling V, Hsu ChC Site-specific tryptophan oxidation induced by autocatalytic reaction of polysorbate 20 in protein formulation. *Pharm Res*, 2011(28), p. 2543–55.
72. Langley JM, Reich D, Aggarwal N, Connor D, Lebel, MH, Gupta A, Garfield H, Ping L, Madan A, Vaughn D, Randomized, multicenter trial of a single dose of AS03-adjuvanted or unadjuvanted H1N1 2009 pandemic influenza vaccine in children 6 months to <9 years of age: Safety and immunogenicity. *Pediatric Infectious Disease Journal*, 2012 Aug; 31(8) 848–58.

73. Lee J., Kang J., Kwon N-Y, Sivaraman A., Naik, Jin S-Y, Oh A.R., Shin J-H, Na Y., Lee K., Lee H-J. Dual Inhibition of P-gp and BCRP Improves Oral Topotecan Bioavailability in Rodents. *Pharmaceutics*. 2021 Apr; 13(4): 559. Available at <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8071537/>
74. Levy M, Dupuis LL. JPEN: Parenteral nutrition hypersensitivity. *J Parenter Enteral Nutr*. 1990 Mar-Apr;14(2), p. 213–5.
75. Li C, Sun C, Li S, Han P, Sun H, Ouahab A, Shen Y, Xu Y, Xiong Y, Tu J. Novel designed polyoxyethylene nonionic surfactant with improved safety and efficiency for anticancer drug delivery *Int J Nanomedicine*. Apr 2014(9), p. 2089–100.
76. Loos W. J., Baker S. D., Verweij J., Boonstra J. G., Sparreboom A. Clinical pharmacokinetics of unbound docetaxel: role of polysorbate 80 and serum proteins. *Clin. Pharmacol. Ther.*, 2003(74), p. 364–371.
77. Maeder W, Lieby P, Sebald A, Spycher M, Pedrussio R, Bolli R. Local tolerance and stability up to 24 months of a new 20% proline-stabilized polyclonal immunoglobulin for subcutaneous administration. *Biologicals*, Jan 2011, 39(1), p. 43–9.
78. Maggio E. T. Polysorbates, peroxides, protein aggregation and immunogenicity – a growing concern. (Review article) *J Excipients Food Chem*, Jun 2012, 3(2), p. 45–53.
79. Maiorana A, Sabia A, Corsetti T, Dionisi-Vici C. Safety of vaccines administration in hereditary fructose intolerance. *Orphanet J Rare Dis*. 2020 Oct 1;15(1):274. doi: 10.1186/s13023-020-01552-z. PMID: 33004052; PMCID: PMC7528578.
80. Martone WJ et al. Illness with fatalities in premature infants: Association with an intravenous Vitamin E preparation, E-Ferol. *Pediatrics* 1986; 78, p. 591–600.
81. Martos A, Koch W, Jiskoot W, Wuchner K, Winter G, Friess W, Hawe A. Trends on Analytical Characterization of Polysorbates and their Degradation Products in Biopharmaceutical Formulations. *J Pharm Sci*. 2017; pii: S0022-3549(17)30157-0. doi: 10.1016/j.xphs.2017.03.001.
82. Masi S, de Cléty SC, Anslot C, and Dettaille T. Acute amiodarone toxicity due to an administration error: could excipient be responsible? *Br J Clin Pharmacol*. Jun 2009; 67(6), p. 691–693.
83. Masini et al. Histamin-releasing properties of polysorbate 80 in vitro and in vivo: correlation with its hypotensive action in the dog. *Agents Actions*. 1985 Sep; 16(6): 470–7.
84. Masters JR, McDermott BJ, Jenkins WE, Fenwick E, Shah PJ, Mundy AR, Loadman PM, Bibby MC. ThioTEPA pharmacokinetics during intravesical chemotherapy and the influence of Tween 80. *Cancer Chemother Pharmacol*. 1990; 25(4), p. 267–73.
85. McKean and Pesce, Determination of polysorbate in ascites fluid from a premature infant. *J Anal Toxicol* (1985) 9(4), p. 174–176.
86. Ménard N, Tsapis N, Poirier C, Arnauld T, Moine L, Lefoiulon F, Péan J-M, and Fattal E Physicochemical characterisation and toxicity evaluation of steroid-based surfactants designed for solubilisation of poorly soluble drugs. *Eur J Pharm Sci* 2011; 44:595–601.
87. Morselli PL, Clinical Pharmacokinetics in neonates. *Clin Pharmacokin* 1976(1), p. 81–98.
88. Munoz A, Karila P, Gallay P, Zettelmeier F, Messner P, Mery M, Grolleau R. A randomized hemodynamic comparison of intravenous amiodarone with and without Tween 80. *Eur Heart J*. Feb 1988, 9(2), p. 142–8.

89. M.V.I. pediatrics® label
<https://dailymed.nlm.nih.gov/dailymed/fda/fdaDrugXsl.cfm?setid=7589f8f3-e82a-43b0-868a-cbb991e2183f>
90. Ng SSW, Figg WD and Sparreboom A, Taxane-mediated antiangiogenesis in vitro – influence of formulation vehicles and binding proteins. *Cancer Res.* Feb 2004 , 64(3), p. 821–4.
91. OECD Environment, Health and Safety Publications Series on Testing and Assessment, No. 43, Guidance document on mammalian reproductive toxicity testing and assessment, July 24, 2008.
92. Palacios Castaño MI, Venturini Díaz M, Lobera Labairu T, González Mahave I, Del Pozo Gil MD, Blasco Sarramián A. Anaphylaxis Due to the Excipient Polysorbate 80. *J Investig Allergol Clin Immunol.* 2016;26(6):394–396. doi: 10.18176/jiaci.0109. PMID: 27996954.
93. Paoletti G, Racca F, Piona A, Melone G, Merigo M, Puggioni F, Ferri S, Azzolini E, Lagioia M, Lamacchia D, Cataldo G, Cecconi M, Canonica GW, Heffler E. Successful SARS-CoV-2 vaccine allergy risk-management: The experience of a large Italian University Hospital. *World Allergy Organ J.* 2021 May;14(5):100541. doi: 10.1016/j.waojou.2021.100541. Epub 2021 Apr 8. PMID: 33850601; PMCID: PMC8030995.
94. Pasche-Koo F, Piletta PA, Hunziker N, Hauser C, High sensitization rate to emulsifiers in patients with chronic leg ulcers, *Contact Dermatitis* 1994, 31(4), p. 226–228.
95. Paudel R, Dogra P, Suman S, Acharya S, Matta J. Acute Liver and Renal Failure: A Rare Adverse Effect Exclusive to Intravenous form of Amiodarone. *Case Rep Crit Care.* 2016; 2016:5232804.
96. Perez-Perez L, Garcia-Gavin J, Pineiro B, Zulaica A, Biologic induced urticaria due to polysorbate 80: usefulness of prick test, *British Journal of Dermatology*, 2011, 164(5), p. 1119–1120.
97. Perino E, Freymond N, Devouassoux G, Nicolas JF, Berard F. Xolair-induced recurrent anaphylaxis through sensitization to the excipient polysorbate. *Ann Allergy Asthma Immunol.* 2018 Jun; 120(6):664–666. doi: 10.1016/j.anai.2018.02.018. Epub 2018 Feb 23. PMID: 29481891.
98. Pesce AJ1, McKean DL. Toxic susceptibilities in the newborn with special consideration of polysorbate toxicity. *Ann Clin Lab Sci.* 1989 Jan-Feb; 19(1), p. 70–3.
99. Price KS, Hamilton RG. Anaphylactoid reactions in two patients after omalizumab administration after successful long-term therapy. *Allergy Asthma Proc.* 2007 May-Jun; 28(3):313–9. doi: 10.2500/aap.2007.28.3003. PMID: 17619560.
100. Ratz Bravo AE, Drewe J, Schlienger RG, Krahenbuhl S, Pargger H, Ummenhofer W., Hepatotoxicity during rapid intravenous loading with amiodarone Description of three cases and review of the literature, *Critical Care Medicine*, 2005, 33(1), p. 128–134.
101. Redfern et al. 2003, *Cardiovasc. Res.* 58, p. 32–45.
102. Rege B.D., Kao J.P.Y., Polli J.E. Effects of nonionic surfactants on membrane transporters in Caco-2 cell monolayers. *European Journal of Pharmaceutical Sciences* 2002, 16:237–246.
103. Rhodes A, Eastwood JB, Smith SA. Early acute hepatitis with parenteral amiodarone: a toxic effect of the vehicle? *Gut.* 1993; 34(4), p. 565–6.
104. Rivera A Jr, Abdo KM, Bucher JR, Leininger JR, Montgomery CA, Roberts RJ. Toxicity studies of intravenous vitamin E in newborn rabbits. *Dev Pharmacol Ther.* 1990; 14(4), p. 231–7.

105. Saariaho AH, Vuorela A, Freitag TL, Pizza F, Plazzi G, Partinen M, Vaarala O, Meri S., Autoantibodies against ganglioside GM3 are associated with narcolepsy-cataplexy developing after Pandemrix vaccination against 2009 pandemic H1N1 type influenza virus. *J Autoimmun.* 2015 Sep; 63, p. 68–75.
106. Saul JP., Scott W. A., Brown S., Marantz P., Acevedo V., Etheridge SP., Perry JC., Triedman J. K., Burriss S. W., Cargo P., Graepe J., Koskelo E-K., Wang R. Intravenous Amiodarone for Incessant Tachyarrhythmias in Children. A Randomized, Double-Blind, Antiarrhythmic Drug Trial. *Circulation* 2005 Nov 29; 112(22):3470–7.
107. Schwartzberg LS, Navari RM. Safety of Polysorbate 80 in the Oncology Setting. *Adv Ther.* 2018 Jun; 35(6):754–767. doi: 10.1007/s12325-018-0707-z. Epub 2018 May 23. PMID: 29796927; PMCID: PMC6015121.
108. Sellaturay P, Gurugama P, Harper V, Dymond T, Ewan P, Nasser S. The Polysorbate containing AstraZeneca COVID-19 vaccine is tolerated by polyethylene glycol (PEG) allergic patients. *Clin Exp Allergy.* 2022 Jan;52(1):12-17. doi: 10.1111/cea.14064. Epub 2021 Dec 9. PMID: 34822190.
109. Sharma B., Immunogenicity of therapeutic proteins. Part 2: Impact of container closures. *Biotechnology Advances* 25, 2007, p. 318–324.
110. Shelley WB, Talanin N, Shelley ED. Polysorbate 80 hypersensitivity. Letters to the Editor. *The Lancet* Volume 345, Issue 8960, 20 May 1995, p. 1312–1313.
111. Shultze V, D’Agosto V, Wack A, Novicki D, Zorn J, Henning R. Safety of MF59™ adjuvant. *Vaccine*, 2008(26), p. 3209–22.
112. Sicard M, Besse P, Choussat A, Bricaud H. Action hémodynamique de l’amiodarone intraveineuse chez l’homme. *Arch Mal Coeur* 1977(3), p. 219–227.
113. Singh SK, Mahler HC, Hartman C, Stark CA. Are Injection Site Reactions in Monoclonal Antibody Therapies Caused by Polysorbate Excipient Degradants? *J Pharm Sci.* 2018 Nov; 107(11):2735–2741. doi: 10.1016/j.xphs. 2018.07.016. Epub 2018 Jul 25. PMID: 30055223.
114. Sparreboom A et al., Determination of the docetaxel vehicle, polysorbate 80, in patient samples by liquid chromatography-tandem mass spectrometry. *J Chromatogr B Analyt Technol Biomed Life Sci.* 2002 Jun 25; 773(2), p. 183–90.
115. Steele RH, Limaye S, Cleland B, Chow J, Suranyi MG. Hypersensitivity reactions to the polysorbate contained in recombinant erythropoietin and darbepoietin. *Nephrology (Carlton).* 2005 Jun; 10(3):317–20. doi: 10.1111/j.1440-1797.2005.00389.x. PMID: 15958049.
116. Szebeni J. Complement activation-related pseudoallergy: a new class of drug-induced acute immune toxicity. *Toxicology.* 2005 Dec 15;216 (2–3): p. 106–21.
117. Szebeni J. Complement activation-related pseudoallergy: a stress reaction in blood triggered by nanomedicines and biologicals. *Mol Immunol.* 2014 Oct;61(2):163-73. doi: 10.1016/j.molimm.2014.06.038. Epub 2014 Aug 12. PMID: 25124145.
118. Tagawa N, Sugiyama E, Tajima M, Sasaki Y, Nakamura S, Okuyama H, Shimizu H, Sato VH, Sasaki T, Sato H. Comparison of adverse events following injection of original or generic docetaxel for the treatment of breast cancer. *Cancer Chemother Pharmacol.* 2017 Sep 1.

119. Takehisa H. et al., Release behavior of diethylhexyl phthalate from the polyvinyl-chloride tubing used for intravenous administration and the plasticized PVC membrane. *International Journal of Pharmaceutics*, 2005, p. 30–37.
120. Tatsuishi T, Oyama Y, Iwase K, Yamaguchi JY, Kobayashi M, Nishimura Y, Kanada A, Hirama S. Polysorbate 80 increases the susceptibility to oxidative stress in rat thymocytes, *Toxicology*, 2005 Feb 1; 207(1), p. 7–14.
121. ten Tije AJ et al., Disposition of polyoxyethylated excipients in humans: implications for drug safety and formulation approaches. *Clin Pharmacol Ther* 2003b, 74(5):509–10.
122. ten Tije AJ, Verweij J., Loos W. J., Sparreboom A. Pharmacological effects of formulation vehicles: implications for cancer chemotherapy. *Clin. Pharmacokinet.*, 2003a, 42, p. 665–685.
123. Thackaberry EA, Kopytek S, Sherratt P, Trouba K and McIntyre B. Comprehensive investigation of hydroxypropyl methylcellulose, propylene glycol, polysorbate 80, and hydroxypropyl-beta-cyclodextrin for use in general toxicology studies. *Toxicol Sci*. Oct 2010, 117(2), p. 485–92.
124. Tompkins L, Lynch C, Haidar S. Polli J, Wang H Effects of commonly used excipients on the expression of CYP3A4 in colon and liver cell *Pharm Res* 2010, 27, p. 1703–12.
125. Torres-Arraut E, Singh S, Pickoff AS. Electrophysiologic effects of Tween 80 in the myocardium and specialized conduction system of the canine heart. *J Electrocardiol*. 1984; 17, p. 145–51.
126. Treon JF, Gongwer LE, Nelson MF and Kirschman JC. Physiologic and metabolic patterns of non-ionic surfactants. In: *Chemistry, physics and application of surface active substances* (4th edition). Ed. Asinger F. Gordon and Breach, London, UK, 1967, p. 381–395.
127. Troelnikov A, Perkins G, Yuson C, Ahamdie A, Balouch S, Hurtado PR, Hissaria P. Basophil reactivity to BNT162b2 is mediated by PEGylated lipid nanoparticles in patients with PEG allergy. *J Allergy Clin Immunol*. 2021 Jul; 148(1):91–95. doi: 10.1016/j.jaci.2021.04.032. Epub 2021 May 12. PMID: 33991580.
128. Vaarala O, Vuorela A, Partinen M, Baumann M, Freitag TL, Meri S, Saavalainen P, Jauhiainen M, Soliymani R, Kirjavainen T, Olsen P, Saarenpää-Heikkilä O, Rouvinen J, Roivainen M, Nohynek H, Jokinen J, Julkunen I, Kilpi T. Antigenic differences between AS03 adjuvanted influenza A (H1N1) pandemic vaccines: implications for pandemrix-associated narcolepsy risk. *PLoS One*. 2014 Dec 15; 9(12):e114361.
129. van Tellingen et al., Rapid Esterase-sensitive Breakdown of Polysorbate 80 and Its Impact on the Plasma Pharmacokinetics of Docetaxel and Metabolites in Mice. *Clin Cancer Res* 1999(5), p. 2918–2924.
130. Varma RK, Kaushal R, Junnarkar AY, Thomas GP, Naidu MU, Singh PP, Tripathi RM, Shridhar DR. Polysorbate 80: a pharmacological study. *Arzneimittelforschung*. 1985; 35(5):804–8. PMID: 4026903.
131. Villa M, Black S, Groth N, Rothman KJ, Apolone G, Weiss NS, Aquino I, Boldori L, Caramaschi F, Gattinoni A, Malchiodi G, Crucitti A, Della Cioppa G, Scarpini E, Mavilio D, Mannino S Safety of MF59-adjuvanted influenza vaccination in the elderly: Results of a comparative study of MF59-adjuvanted vaccine versus nonadjuvanted influenza vaccine in Northern Italy *Am J Epidemiol* 2013 178(7), p. 1139–45.
132. Wan LS and Lee PF, CMC of polysorbates. *J Pharm Sci*. 1974; 63(1):136–137.

133. Wang Y, Wang C, Gong C, Wang Y, Guo G, Luo F, Qian Z *Int J Pharm.* 2012 Sep 15; 434(1–2):1–8. doi: 10.1016/j.ijpharm.2012.05.015. Epub 2012 May 15.)
134. Webster LK, Linsenmeyer ME, Rischin D, Urch ME, Woodcock DM, Millward MJ. Plasma concentrations of polysorbate 80 measured in patients following administration of docetaxel or etoposide. *Cancer chemotherapy and pharmacology* 1997(39), p. 557–60.
135. Weiszhar Z, Czucz J, Révész C, Rosivall L, Szebeni J, Rozsnyay Z. Complement activation by polyethoxylated pharmaceutical surfactants: Cremophor-EL, Tween-80 and Tween-20. *Eur J Pharm Sci.* 2012 Mar 12; 45(4), p. 492–498.
136. Williams J, Odum J, Lewis RW and Brady AM: The oral administration of polysorbate 80 to the immature female rat does not increase uterine weight. *Toxicol Lett.* 1997 Mar 14; 91(1), p. 19–24.
137. Witek R., Krupa S., Kubis A. Cytotoxic action of diethanolamine oleate on Ehrlich exudative carcinoma in mice, compared with the action of polyoxyethylene sorbitan mono-oleate . . Polysorbate 80 coated poly (ϵ -caprolactone)-poly (ethylene glycol)-poly (ϵ -caprolactone) micelles for paclitaxel delivery. (Tween 80). *Arch. Immunol. Ther. Exp. (Warsz)*, 1979(27), p. 321–324.
138. Yang S, Liu J, Chen Y, Jiang J. Reversal effect of Tween-20 on multidrug resistance in tumor cells in vitro. *Biomed Pharmacother.* 2012 Apr; 66(3):187–94. doi: 10.1016/j.biopha.2011.10.007. Epub 2012 Jan 18.
139. Zensi et al 2009, *J Control. Release* 137, p. 78–86.
140. Zhang H, Dou J, Zhai Y, Liu A, Zhai G, Advances in the formulations of non-injection administration of docetaxel, *Journal of Drug Targeting* Volume 22, 2014(2), p. 87–94.
141. Zhang W, Li Y, Zou P, Wu M, Zhang Z, Zhang T. The Effects of Pharmaceutical Excipients on Gastrointestinal Tract Metabolic Enzymes and Transporters-an Update. *AAPS J.* 2016 Jul; 18(4):830–43.
142. Zhang Sisi, Zhang, Hui Xiao, Rosalynn Molden, Haibo Qiu, Ning Li, Rapid Polysorbate 80 Degradation by Liver Carboxylesterase in a Monoclonal Antibody Formulated Drug Substance at Early Stage Development. *Journal of Pharmaceutical Sciences* 109 (2020) 3300–3307.
143. Zhou ZH, Stone CA Jr, Jakubovic B, Phillips EJ, Sussman G, Park J, Hoang U, Kirshner SL, Levin R, Kozlowski S. Anti-PEG IgE in anaphylaxis associated with polyethylene glycol. *J Allergy Clin Immunol Pract.* 2021 Apr; 9(4):1731–1733.e3. doi: 10.1016/j.jaip.2020.11.011. Epub 2020 Nov 17. PMID: 33217616; PMCID: PMC8090930.